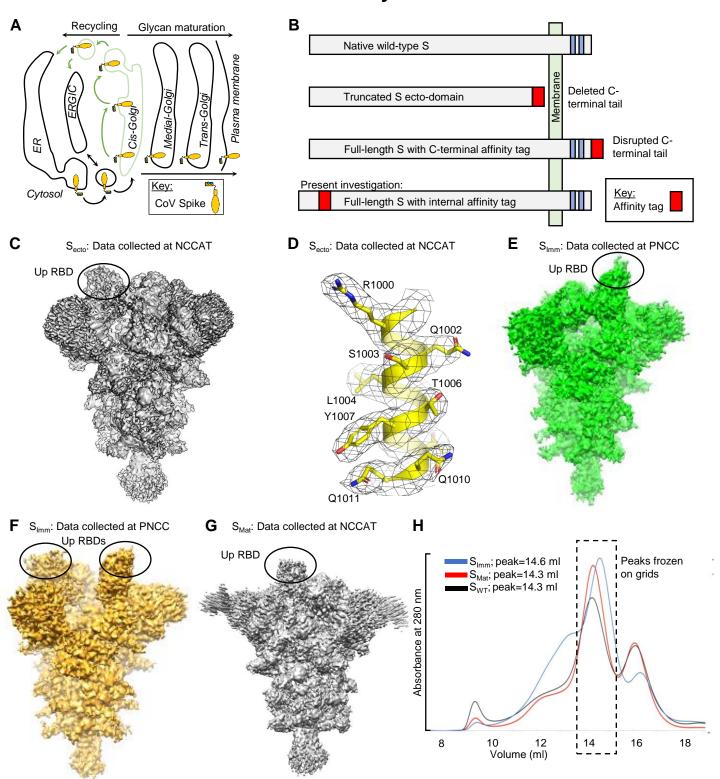
## Preliminary data on feasibility:

Our team has the expertise to handle all data processing on our own as shown below and in our biosketches. Arresting S maturation intermediates in distinct ER-Golgi-plasma membrane compartments (Fig. A) has been made possible by an innovative construct design that retains the native trafficking signals in the cytosolic tail by inserting an affinity purification tag in an internal site rather than the tail C-terminus (Fig. B). Using this approach, we have generated an S construct that is retained in ER and cis-Golgi and displays immature, high-mannose glycans (S<sub>imm</sub>), a construct that is rapidly exported to the plasma membrane without repeated recycling in the glycan modifying compartments and displays mature, complex-type glycans (S<sub>Mat</sub>), the S<sub>WT</sub> construct that recycles in these compartments prior to export to plasma membrane, and an S ecto-domain (S<sub>ecto</sub>) as a control to compare with massive amounts of published data on S<sub>ecto</sub>. The S<sub>WT</sub> construct differs from  $S_{lmm}$  at one residue (His1271 in  $S_{WT}$  substituted by Lys1271 in  $S_{lmm}$ ) whereas it differs from  $S_{Mat}$  at two residues (Lys1269/His1271 in  $S_{WT}$  substituted to Ala1269/Ala1271 in  $S_{Mat}$ ). As shown in our initial GUP1 application (NCCAT-GUP1-SH220622), glycan analysis by mass spectrometry validates the processing of these S maturation constructs. Single particle  $\it cryoEM \ data$  have been collected on  $S_{ecto}$  and  $S_{Mat}$  at NCCAT and on  $S_{lmm}$  at PNCC, on grids *prepared in-house*. The S<sub>ecto</sub> reconstruction is at a resolution of 2.5Å (Fig. C, D). The structure of this internally tagged S<sub>ecto</sub> construct is consistent with previously published Cterminally tagged S<sub>ecto</sub> structures, which *validates* our internal tagging approach in maintaining the tertiary and quaternary organization of the spike. This dataset displays a predominantly one-up state of the RBDs upon separation of Sector conformational states by heterogeneous refinement, 3D classification, and 3D variability analysis. In contrast, the S<sub>Imm</sub> dataset from PNCC, reconstructed to a resolution of 3.2Å, displays a wide conformational *diversity* in the RBDs (Fig. E, F). As S<sub>imm</sub> has ~1.7 fold more immature glycans than S<sub>ecto</sub>, this suggests that glycan maturation changes during inter-organelle trafficking play a critical role in *modulating spike RBD conformations*. This glycan maturation difference is *larger* between full-length  $S_{lmm}$ ,  $S_{WT}$ , and  $S_{Mat}$ , especially at regulatory glycan sites such as at Asn343. An on-the-fly S<sub>Mat</sub> reconstruction by the NCCAT staff during the cryosparc-live session shows an architecture of characteristic of the spike protein (Fig. G). These  $S_{Mat}$  data are currently in early stages of processing in our lab. Hence, the S<sub>WT</sub> data collected under this RAP proposal will complete the entire panel of spike glycan maturation intermediates.

**Status of ready-to-go grids for S**<sub>WT</sub>: The S<sub>WT</sub> protein has been purified under the conditions established for S<sub>Imm</sub> and S<sub>Mat</sub> (**Fig. H**), which are the two spike constructs we have characterized both biochemically and by single particle cryoEM. Freezing of grids for S<sub>WT</sub> have also been prepared under the same conditions as the grids for S<sub>Imm</sub>, S<sub>Mat</sub>, and S<sub>ecto</sub>. Our data show that all the full-length spike constructs demonstrate similar ice thickness and particle distribution on the cryoEM grids whereas S<sub>ecto</sub> has a higher particle density. **The S**<sub>WT</sub> **grids are in storage and ready to be shipped within a 24-hour notice.** 

## **Preliminary Data**



**Figure: Preliminary data. (A)** S recycling and export pathway. **(B)** S construct designs contrasting our strategy of internal tagging with prior C-terminal truncation and C-terminal tagging strategies. **(C and D)** Unsharpened 2.5Å resolution cryoEM map of  $S_{ecto}$ . **(E and F)** CryoEM reconstructions of  $S_{lmm}$  showing two different RBD conformations. **(G)** Preliminary cryoEM reconstruction of our recently collected  $S_{Mat}$  dataset from NCCAT. **(H)** Chromatograms showing purification of  $S_{lmm}$ ,  $S_{Mat}$ , and  $S_{WT}$  in detergent on a superose 6 size-exclusion chromatography column.