

BIOGRAPHICAL SKETCH

Provide the following information for the Senior/key personnel and other significant contributors.
Follow this format for each person. **DO NOT EXCEED FIVE PAGES.**

NAME: Taylor, Lily

eRA COMMONS USER NAME (credential, e.g., agency login): lilytaylor

POSITION TITLE: Graduate Student Research Assistant

EDUCATION/TRAINING *(Begin with baccalaureate or other initial professional education, such as nursing, include postdoctoral training and residency training if applicable. Add/delete rows as necessary.)*

INSTITUTION AND LOCATION	DEGREE (if applicable)	Start Date MM/YYYY	Completion Date MM/YYYY	FIELD OF STUDY
University of Arizona	BS	08/2016	05/2020	Biochemistry
UC Los Angeles (UCLA)	PHD	07/2020	In Progress	Biochemistry, Molecular and Structural Biology

A. Personal Statement

My primary research interests involve the application of methods in synthetic and structural biology to assess the relationship between protein sequence and function. My interest in ascribing the functional properties of a protein to its encoded sequence parallels my undergraduate research experiences in which I modeled the structural properties of small molecular dimers under the guidance of Professor Stephen Kukolich at the University of Arizona to define key interactions between the molecules of interest. In doing so, I became well-versed in computational chemistry methods and appreciative of the vast information gained from a thorough structural understanding of a particular system. Upon the completion of my undergraduate degree, I entered the Biochemistry, Molecular and Structural Biology PhD program at UCLA where I now apply computational and structural methods to biological systems as an avenue to study how viruses exploit sequence-function relationships to gain broader pathogenic properties, and how this information can be used to predict the scope of their functional sequence landscapes. Under the advisement of Professor Jose Rodriguez at UCLA, I plan to explore the functional and structural effects of sequence variation of viral machinery implicated in serious human disease - information that will ultimately be utilized for the development of therapeutics and to gain a greater understanding of the molecular properties that govern their pathogenicity.

B. Positions, Scientific Appointments and Honors**Positions and Scientific Appointments**

2018 – 2020 Undergraduate Student Researcher, University of Arizona
2020 – Present Graduate Student Researcher, UC Los Angeles

Honors

2016 – 2020 Honors Program Member, University of Arizona
2016 – 2020 Arizona Excellence Scholarship Award, University of Arizona
2017 – 2020 Silver and Sage Scholarship Award, University of Arizona
2022 –2024 Cellular and Molecular Biology Training Fellowship, UC Los Angeles

C. Contributions to Science

Undergraduate Research: As an undergraduate at the University of Arizona I worked in the lab of Professor Stephen Kukolich where I studied the properties of concerted proton-tunneling interactions in hydrogen-bonded networks. I worked towards defining the tunneling frequencies of proton exchanges between symmetrically hydrogen-bonded acids using microwave spectroscopy and high-performance computing. I also applied these methods to obtain precise structural models of transition metal complexes and small molecular dimers.

Graduate Research: My ongoing predoctoral research is focused on studying the functional consequences of sequence variation of New World Hemorrhagic Fever Mammarenavirus (NWM) glycoproteins. While all pathogenic NWMs bind the same cellular receptor to initiate infection, the sequences across the receptor-binding domains of NWM glycoproteins differ substantially, with sequence identities ranging from 25-46%. This sequence variability curtails efforts to develop broadly neutralizing therapeutics against these deadly pathogens and limits our ability to prepare for and identify emerging strains that may pose a serious risk to human health. My work aims to elucidate the functional effects of sequence variation of the NWM glycoproteins to build a comprehensive understanding of the sequence-level factors which regulate their pathogenic properties. This information will be extracted by first mapping the paths that traverse the functional sequence landscape between human infection-causing strains, determining the sites of structural variability across these functional variants, and using this information to identify key sites that can be exploited to inform the rational design of proteins and inhibitors that modulate viral infectivity.

1. Taylor, L. J., Rodriguez, J. A. (2024) Approaches to Characterize *In Situ* Viral Glycoprotein Structure and Function. Poster presented at: American Crystallographic Association; July 2024; Denver, OH.
2. Taylor, L. J., Rodriguez, J. A. (2024) Evaluating the Functional Implications of Sequence Variation Across New World Hemorrhagic Fever Mammarenavirus Glycoproteins. Poster presented at: American Society for Virology; June 2024; Columbus, OH.
3. Taylor, L. J., Rodriguez, J. A. (2023) Toward Structural Insights into New World Hemorrhagic Fever Mammarenavirus Glycoprotein-Mediated Infection. Poster presented at: West Coast Structural Biology Workshop; 2023 March; Pacific Grove, CA.
4. Taylor, L. J., Ferrero, S., Helguera, G., Rodriguez, J. A. (2022) Predicting the Function of New World Hemorrhagic Fever Mammarenavirus Glycoprotein Variants. Poster presented at: Seaborg Symposium; 2022 August; Los Angeles, CA and Sigman Symposium; 2022 October; Los Angeles, CA.

D. Scholastic Performance

YEAR	COURSE TITLE	GRADE
UNIVERSITY OF ARIZONA		
2016	Honors Fundamentals of Chemistry	B
2016	Honors Fundamental Techniques of Chemistry	B
2016	Honors Advanced First-Year Composition	A
2016	Honors Special Topics in Science	A
2016	Calculus 1	B
2016	Spanish 1	A
2017	Honors Quest	A
2017	Honors Colloquium	A
2017	Calculus 2	C
2017	Spanish 2	A
2017	Minority Biomedical Research Colloquium	A
2017	Honors Organic Chemistry	B
2017	Honors Organic Chemistry Lab	A

2017	Mathematical Physics for Chemistry	B
2017	General Biology Lab	A
2017	General Biology	C
2017	The Pursuit of Happiness	A
2018	Introduction to Biochemical Research	A
2018	Honors Organic Chemistry 2	B
2018	Honors Organic Chemistry Lab 2	A
2018	Dance Appreciation	A
2018	General Biology 2	A
2018	General Biology 2 Lab	A
2018	Nutrition, Food and You	A
2018	Introduction to African American Literature	A
2018	Biochemistry 1	A
2018	Ecology	A
2018	Medical Ethics	A

YEAR	COURSE TITLE	GRADE
2018	Introductory Mechanics	A
2019	Biochemistry 2	A
2019	Biochemistry Lab Techniques	A
2019	Honors Colloquium	A
2019	Environmental Ethics	A
2019	Introductory Electricity and Magnetism	B
2019	Physical Chemistry 1	A
2019	Word Roots: Science and Medical Terms	A
2019	Introductory Biostatistics	B
2019	Introductory Biotechnology	A
2020	Honors Water Use and Sustainability	A
2020	Biochemistry of Nucleic Acids	A
2020	Honors Thesis 2	A
2020	Physical Chemistry 2	A
2020	Plants and Our World	A
UC LOS ANGELES		
2020	Biochemistry Research Seminar	A
2020	Protein Structure	A
2020	Biocatalysts and Bioenergetics	A
2020	Nucleic Acid Structure	A
2020	Gene Expression	A
2020	Biomolecular Structure and Regulation	A
2021	Structural Biology	A
2021	Structural Molecular Biology	A
2021	Structural Molecular Biology Lab	A
2021	Protein Mass Spectroscopy	A
2021	Stem Cell Seminar	S
2022	Chemistry of Biology	S
2022	Organic Colloquium	A
2023	Core Principles in Cell and Molecular Biology	A
2023	Skills Development for Cell & Molecular Biologists	S
2023	Advanced Topics and Approaches in Cell and Molecular Biology	S
2023	Ethics and Accountability in Biomedical Research	S

BIOGRAPHICAL SKETCH

Provide the following information for the Senior/key personnel and other significant contributors.
Follow this format for each person. **DO NOT EXCEED FIVE PAGES.**

NAME: Rodriguez, Jose A

eRA COMMONS USER NAME (credential, e.g., agency login): JRODRIGUEZ2

POSITION TITLE: Associate Professor, Department of Chemistry & Biochemistry, UCLA

EDUCATION/TRAINING *(Begin with baccalaureate or other initial professional education, such as nursing, include postdoctoral training and residency training if applicable. Add/delete rows as necessary.)*

INSTITUTION AND LOCATION	DEGREE (if applicable)	Completion Date MM/YYYY	FIELD OF STUDY
University of California, Los Angeles (UCLA)	B.S.	06/2007	Biophysics
University of California, Los Angeles (UCLA)	Ph.D.	09/2012	Molecular Biology
University of California, Los Angeles (UCLA)	Postdoctoral	07/2016	Biological Chemistry

A. Personal Statement

My research is aimed at understanding fundamental aspects of molecular structure. This includes an improved understanding of the relationship between biological structure and function, the development of new tools and approaches that advance our knowledge of biochemistry, and the discovery of new treatments to combat devastating and incurable diseases.

My formative scientific experiences have allowed me to collaboratively contribute to several areas of biomedical science including (1) characterization of the millisecond timescale swimming behavior of *Trypanosoma brucei*, the parasitic pathogen responsible for African sleeping sickness, (2) use of high brilliance X-ray sources to uncover the three dimensional structure of the *Neospora caninum* cell (3) development of antibody-based immunotherapies to treat cancer, (4) development of inhibitors of infection by New World Arenaviruses – etiological agents of deadly South American hemorrhagic fevers, (5) determination of the structures of insecticidal toxins critical for future pesticide development and vector control for tropical disease eradication, and (6) determination of structures of toxic segments of amyloid forming proteins, including those that cause Parkinson's, Alzheimer's and Prion disease.

My scientific vision has been shaped by an early introduction to methods development in imaging including the use and application of high-powered instruments including brilliant X-ray and electron sources. I have applied these instruments to address pressing questions in aging research. As an example of this, in 2015 I helped advance the new cryo-electron microscopy technique known as electron micro-diffraction (MicroED) by determining a structure of the toxic core of alpha-synuclein fibrils, the cause of Parkinson's disease - the first novel structure determined by MicroED. Since 2015, my group has been at the forefront of electron diffraction developments; we have established ED as a growing scientific field for both small molecule and protein nanocrystals. My group has further leveraged other cryoEM and cryoET approaches to answer open questions in infectious disease and amyloid biology.

B. Positions, Scientific Appointments, and Honors**Positions and Employment**

2021-present Associate Professor, Department of Chemistry and Biochemistry, UCLA
 2016-2021 Assistant Professor, Department of Chemistry and Biochemistry, UCLA
 2014-2017 Visiting Scientist, Biophysics with Tamir Gonen in Janelia Research Campus
 2013-2016 Giannini Post-Doctoral Fellow, Biological Chemistry with David Eisenberg at UCLA
 2013-2015 Lecturer, Physics in the UCLA Department of Physics and Astronomy
 2013-2014 Visiting Scientist, Spring-8 in RIKEN Harima Institute

2012-2013 HHMI Post-Doctoral Associate, Biological Chemistry with David Eisenberg at UCLA
 2010-2012 Graduate Student Mentor, Undergrad Research Center, Tama Hasson at UCLA
 2007-2012 Graduate Student Researcher, Molecular Biology with Manuel L. Penichet at UCLA

Other Experience and Professional Memberships

2020-present USNC/Cr EM representative
 2018-present American Chemical Society, Member
 2017-present Biophysical Society, Member
 2006-2012 American Association for Cancer Research (AACR), Student Member

Honors

2020 Sloan Fellowship
 2019-2021 Packard Fellowship
 2019 UCLA Chem & Biochemistry McCoy Award
 2018 American Chemical Society, Chemistry & Engineering News Talented 12
 2018-2022 Pew Scholar Award
 2017-2020 Searle Foundation Scholar Award
 2017-2021 Beckman Young Investigator Award
 2015 Parvin Foundation Award for Post-Doctoral Research
 2013-2016 Giannini Foundation Post-Doctoral Fellowship
 2011 UCLA AMGEN Molecular Biology Institute Dissertation Year Award
 2007-2007 NSF-AGEP Competitive Edge Graduate Summer Research Fellowship
 2007-2012 Howard Hughes Medical Institute Gilliam Fellowship for Graduate Studies
 2007-2011 UCLA Whitcome Fellowship for Graduate Study in Molecular Biology
 2006-2007 American Association for Cancer Research Thomas J. Bardos Award
 2005 HHMI Excellent Research Opportunities Program
 2005-2007 Minority Access to Research Careers (MARC) trainee, UCLA

C. Contributions to Science

1. *Cryo Electron Microscopy and Electron Crystallography*

The following list of articles outlines my group's efforts in advancing the field of electron diffraction, as well as my application of micro electron diffraction (MicroED) and cryoEM to pressing biomedical problems through the determination of novel macromolecular structures.

- a. Richards LS, Flores MD, Zink S, Schibrowsky NA, Sawaya MR, **Rodriguez JA.** (2023) Cryo-EM Structure of a Human LECT2 Amyloid Fibril Reveals a Network of Polar Ladders at its Core. **Structure** 31, 1386–1393.
- b. Richards LS, Flores MD, Millán C, Glynn C, Zee C, Sawaya MR, Gallagher-Jones M, Borges RJ, Usón I, **Rodriguez JA.** (2023) Fragment-Based Ab Initio Phasing of Peptidic Nanocrystals by MicroED. **ACS Bio Med Chem Au.** doi: 10.1021/acsbiochemau.2c00082.
- c. Kim, L.J., Ohashi, M., Tan, D., Asay, M., Cascio, D., **Rodriguez, J.A.**, Tang, Y., Nelson, H. (2021) Prospecting for natural products by genome mining and microcrystal electron diffraction. **Nature Chem Bio.** 17, 872–877.
- d. Jones, C.G., Martynowycz, M.W., Hattne, J., Fulton, T.J., Stoltz, B.M., **Rodriguez, J.A.**, Nelson, H. and Gonen, T. (2018) The CryoEM Method MicroED as a Powerful Tool for Small Molecule Structure Determination. **ACS Central Sci.** 4 (11), 1587–1592.

2. *Biophysics of Infectious Diseases*

The following select list of articles describes my efforts in novel approaches for structural and biophysical investigation of immunological and infectious diseases. These include the structural elucidation of prion assemblies, the cause of incurable prion disease.

- a. Glynn, C., Hernandez, E., Gallagher-Jones, M., Miao, J., Sigurdson, C. and **Rodriguez, J.A.** (2022) Structural consequences of sequence variation in mammalian prion b2a2 loop segments. **Frontiers in Neuroscience**, 16.
- b. Ferrero, S., Flores, M.D., Short, C., Vazquez, C., Clark, L.E., Ziegenbein, J., Zink, S., Fuentes, D., Payes, C., Batto, M.V., Collazo, M., García, C.C., Abraham, J., Cordo, S.M., **Rodriguez, J.A.**, Helguera, G. (2021) Antibody-Based Inhibition of Pathogenic New World Hemorrhagic Fever

Mammarenaviruses by Steric Occlusion of the Human Transferrin Receptor 1 Apical Domain. **Journal of Virology** 95(17):e0186820.

- c. Glynn, C., Sawaya, M.R., Ge P., Gallagher-Jones, M., Short, C.W., Bowman, R., Apostol, M., Zhou, Z.H., Eisenberg, D.S., **Rodriguez, J.A.** (2020) Cryo-EM structure of a human prion fibril with a hydrophobic, protease-resistant core. *Nature SMB*. doi: 10.1038/s41594-020-0403-y.
- d. Gallagher-Jones, M., Glynn, C., Boyer, D.R., Martynowycz, M.W., Hernandez, E., Miao, J., Zee, C., Novikova, I.V., Goldschmidt, L., McFarlane, H.T., Helguera, G.F., Evans, J.E., Sawaya, M.R., Cascio, D., Eisenberg, D.S., Gonen, T. and **Rodriguez, J.A.** (2018) Sub-ångström cryo-EM structure of a prion protofibril reveals a polar clasp. *NSMB*. 25, 131-134.

3. Structures Relevant to Neuroscience and Neurodegenerative Disease

My group has been engaged in a number of collaborative projects related to amyloid structures. My role has been to provide assistance and leadership in the area of structure determination by frontier methods including MicroED. The work has been fruitful, resulting in a number of original research articles including the four highlighted below.

- a. Flores MD, Sawaya MR, Boyer DR, Zink S, Tovmasyan S, Saucedo A, Zee C, Cardenas J, Fioriti L, **Rodriguez JA**. Structure of a reversible amyloid fibril formed by the CPEB3 prion-like domain reveals a core sequence involved in translational regulation. **bioRxiv** [Preprint]. 2022 bioRxiv 2022.12.07.519389; doi: <https://doi.org/10.1101/2022.12.07.519389>
- b. Hughes, M.P., Sawaya, M.R., Boyer, D.R., Goldschmidt, L., **Rodriguez, J.A.**, Cascio, D., Chong, L., Gonen, T., Eisenberg, D.S. (2018) Atomic structures of low-complexity protein segments reveal kinked β sheets that assemble networks. *Science*. 359(6376), 698-701.
- c. Seidler, P.M., Boyer, D.R., **Rodriguez, J.A.**, Sawaya, M.R., Cascio, D., Murray, K., Gonen, T. and Eisenberg, D.S. (2017) Structure-based inhibitors of tau aggregation. *Nature Chemistry*. doi: 10.1038/nchem.2889
- d. Krotee, P., **Rodriguez, J.A.**, Sawaya, M.R., Cascio, D., Reyes, F.E., Shi, D., Hattne, J., Nannenga, B.L., Oskarsson, M.E., Philipp, S., Griner, S., Jiang, L., Glabe, C.G., Westermarck, G.T., Gonen, T., and Eisenberg, D.S. (2017) Atomic structures of fibrillar segments of hIAPP suggest tightly mated β -sheets are important for cytotoxicity. *Elife*. 6. pi: e19273.

4. X-ray Crystallography & Diffraction Methods

The following select list of articles describes my efforts in frontier crystallographic experiments to advance methodology and/or reveal novel structures.

- a. Gallagher-Jones M., Ophus C., Bustillo KC., Boyer DR., Panova O., Glynn C., Zee C., Ciston J., Mancina KC., Minor AM., and **Rodriguez JA**. (2019) Nanoscale mosaicity revealed in peptide microcrystals by scanning electron nanodiffraction. *Nat. Comms. Biol.* 2:26.
- b. Gallagher-Jones, M., Baraldi-Dias, C.S., Pryor, A.J., Bouchmella, K., Zhao, L., Lo, Y.H., Cardoso, M.B., Shapiro, D., **Rodriguez, J.**, Miao, J. (2017) Correlative cellular ptychography with functionalized nanoparticles at the Fe L-edge. *Nat. Sci. Reports*. 7: 4757.
- c. Colletier, J.P., Sawaya, M.R., Gingery, M., **Rodriguez, J.A.**, Cascio, D., Brewster, A.S., Boutet, S., Williams, G.J., Messerschmidt, M., DePonte, D.P., Sierra, R.G., Laksmono, H., Koglin, J., Hunter, M.S., Park, H.W., Uervirojnangkoorn, M., Bideshi, D.K., Brunger, A.T., Federici, B.A., Sauter, N.K., Eisenberg, D.S. (2016) A potent mosquito larvicide revealed by *de novo* phasing with an X-ray free-electron laser. *Nature*. DOI 10.1038/nature19825.

Comprehensive List of Published Work:

<https://www.ncbi.nlm.nih.gov/sites/myncbi/1RGFUfHRv5957/collections/51179380/public/>