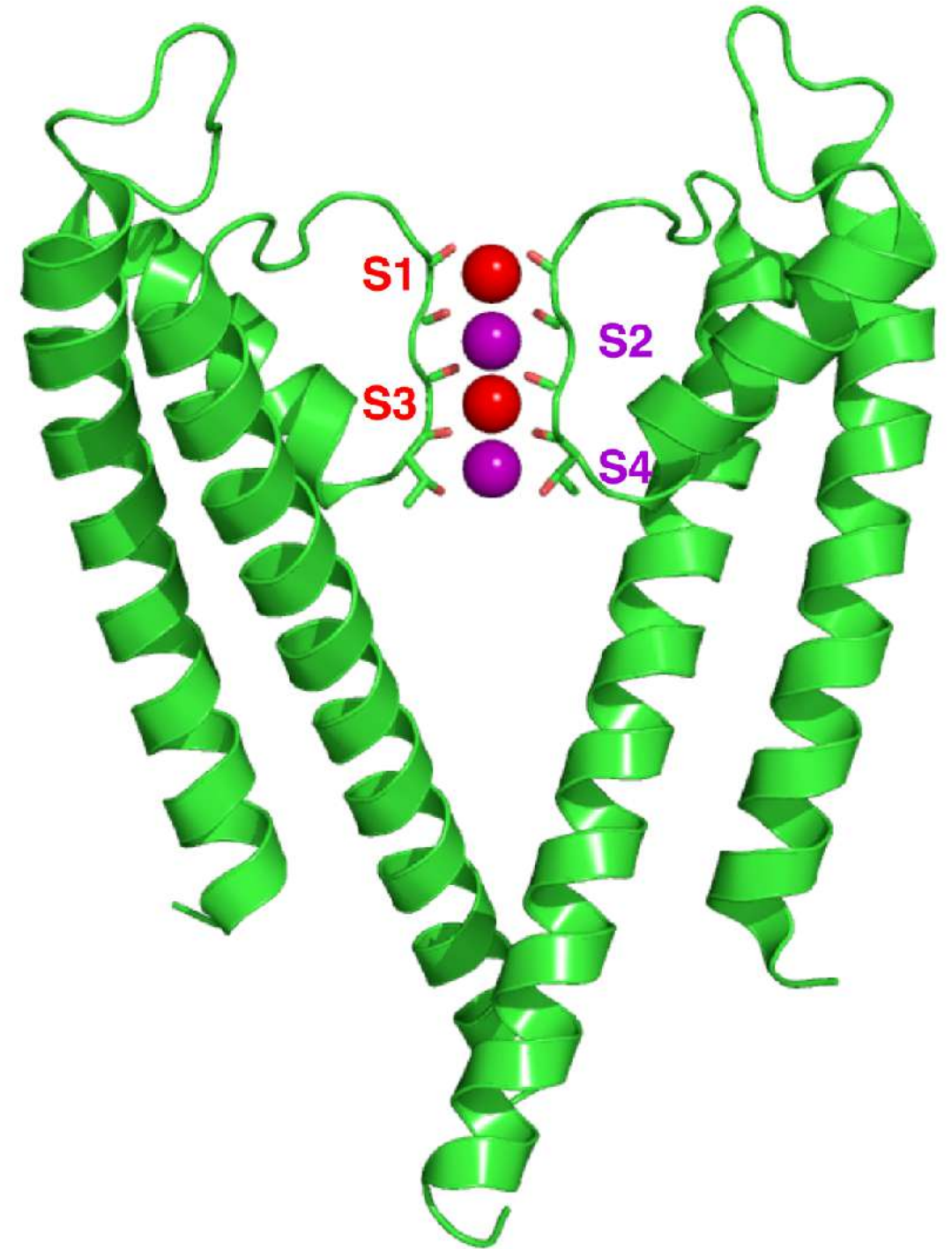


Interpretation and limitation of Single Particle Cryo-EM analysis

Rich Hite
Memorial Sloan Kettering Cancer Center

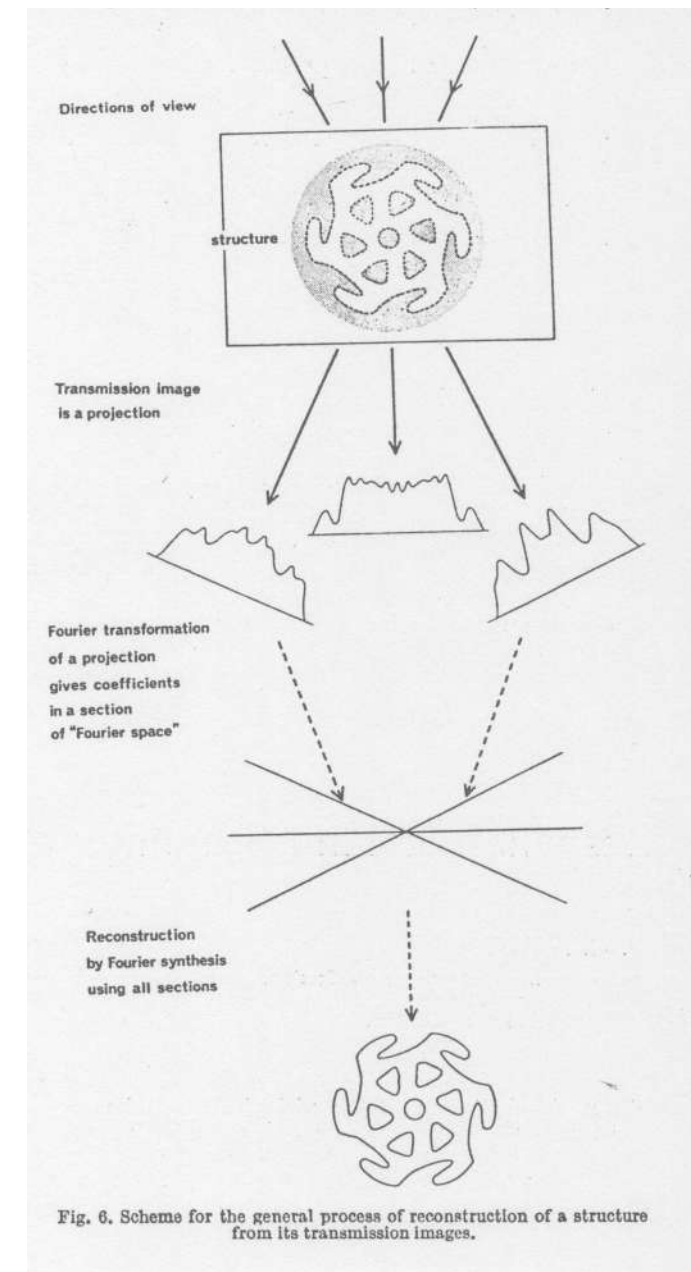
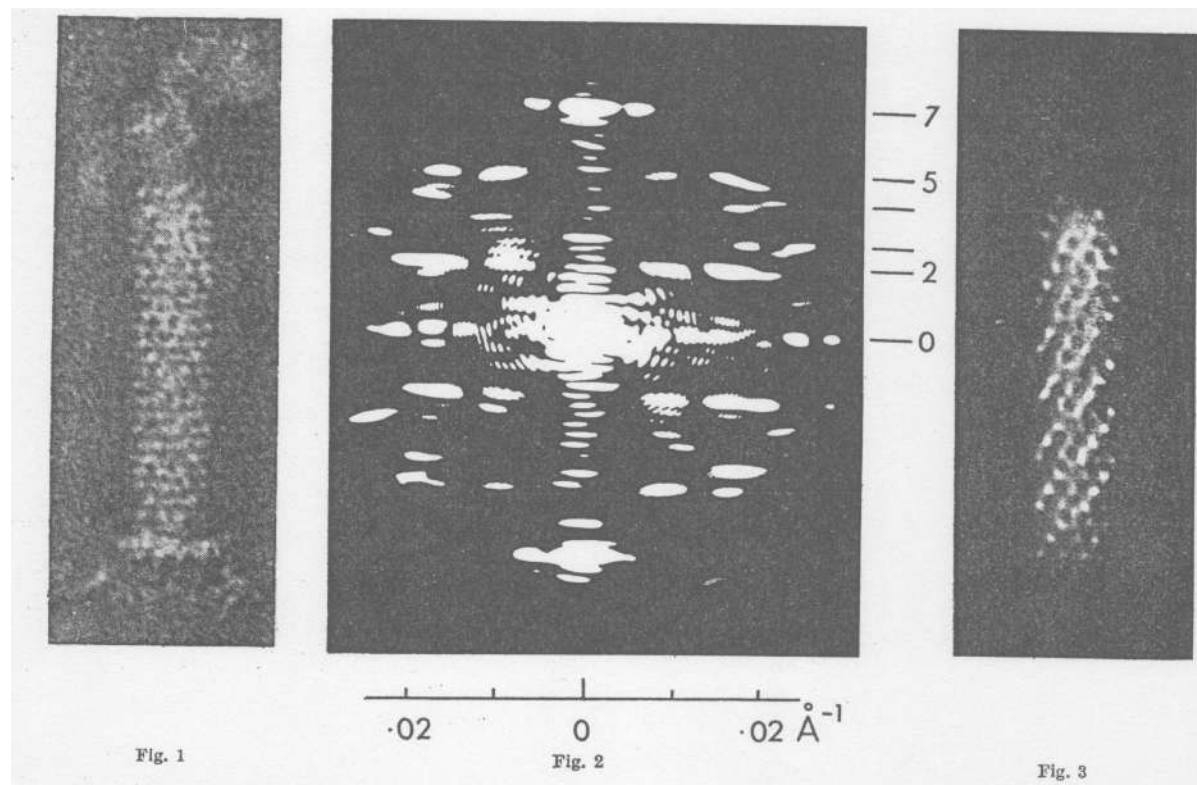
Overview

1. What is structural biology?
2. Why is cryo-electron microscopy currently so popular among structural biologists?
3. What advances have allowed cryo-electron microscopy to become so widely adopted?
4. What are the limitations of Cryo-EM?



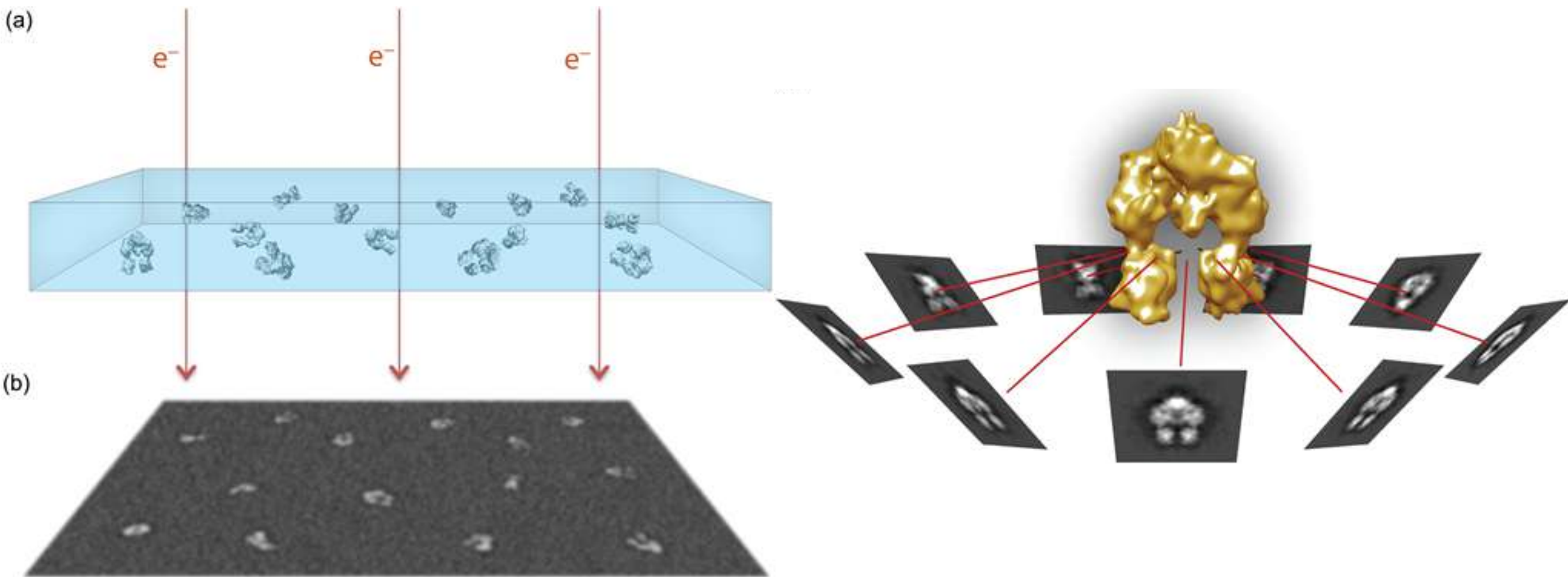
Molecular electron microscopy

- First EM structure -bacteriophage T4 tail - De Rosier and Klug (1968)
 - Applied helical averaging techniques to resolve the structure at $\sim 35 \text{ \AA}$
 - Described the general principles for Fourier synthesis of EM images



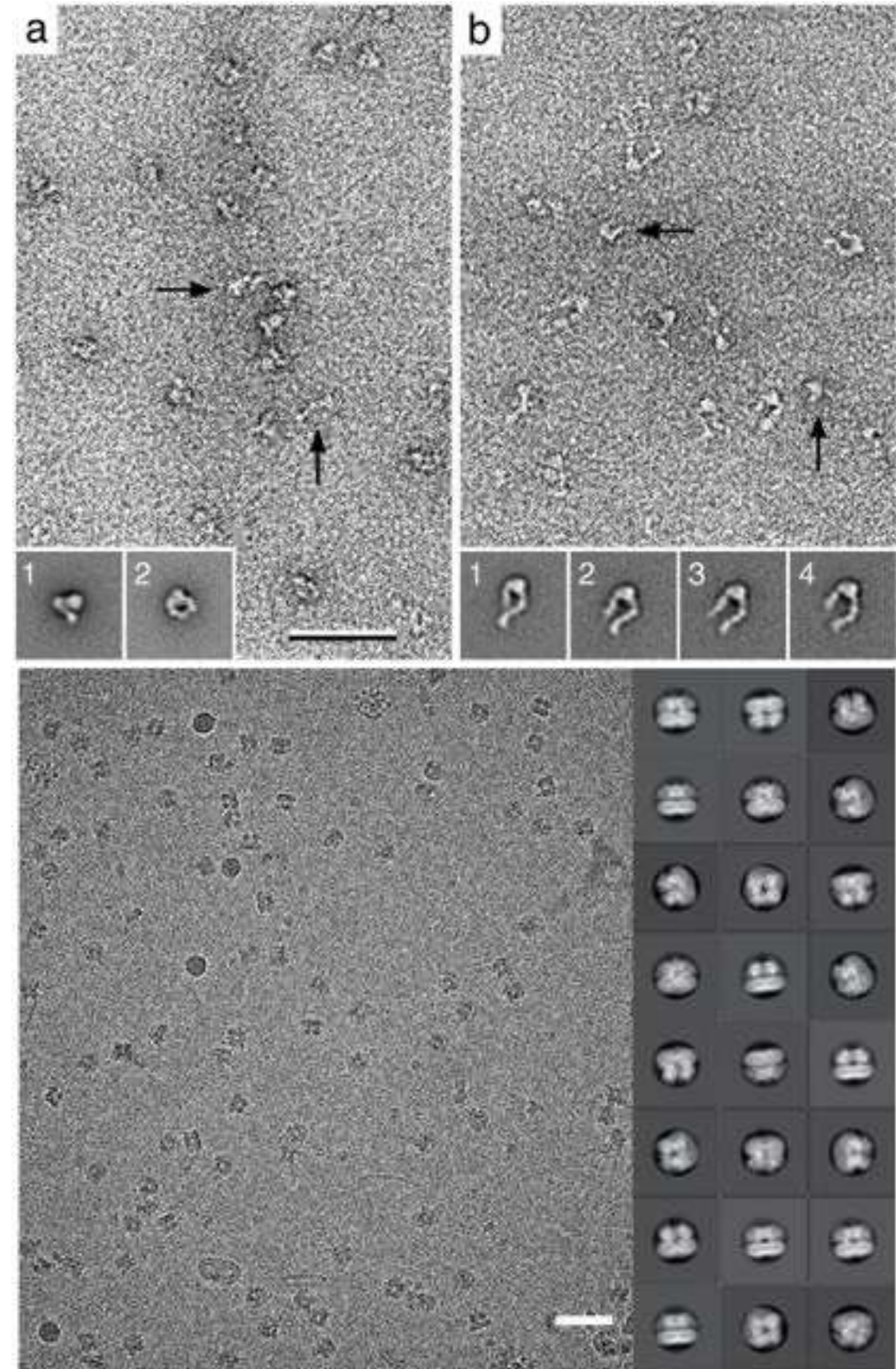
Single-particle electron microscopy

- Cryo-EM images are 2D projections of 3D objects
- Each image therefore can fully describe a single view of the object
- 3D reconstructions are generated from combining many views together



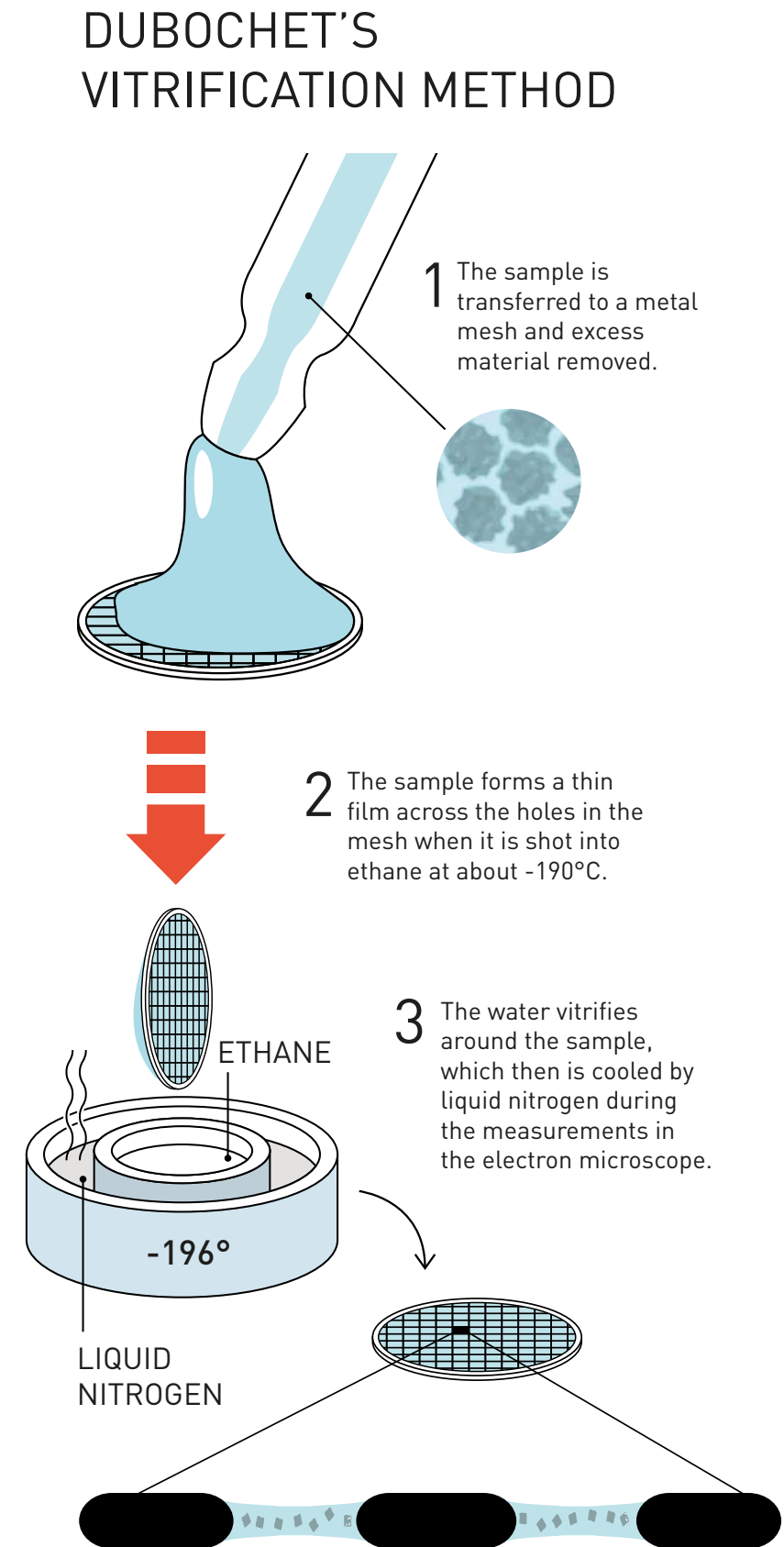
Specimen preparation techniques

- Negative staining - embed the protein in a thin layer of heavy metal
 - Heavy metal scatters electron strongly
 - Visualize areas without stain (i.e. areas with protein)
 - Resolve the envelope of a protein complex
 - Limited to ~ 20 Å (grain size of the staining metal)
- Cryogenic vitrification - embed the protein in a thin layer of vitreous ice
 - Protein scatters more strongly than vitreous ice
 - Resolve complete protein structure (when ordered)
 - Resolution is limited by protein conformational stability



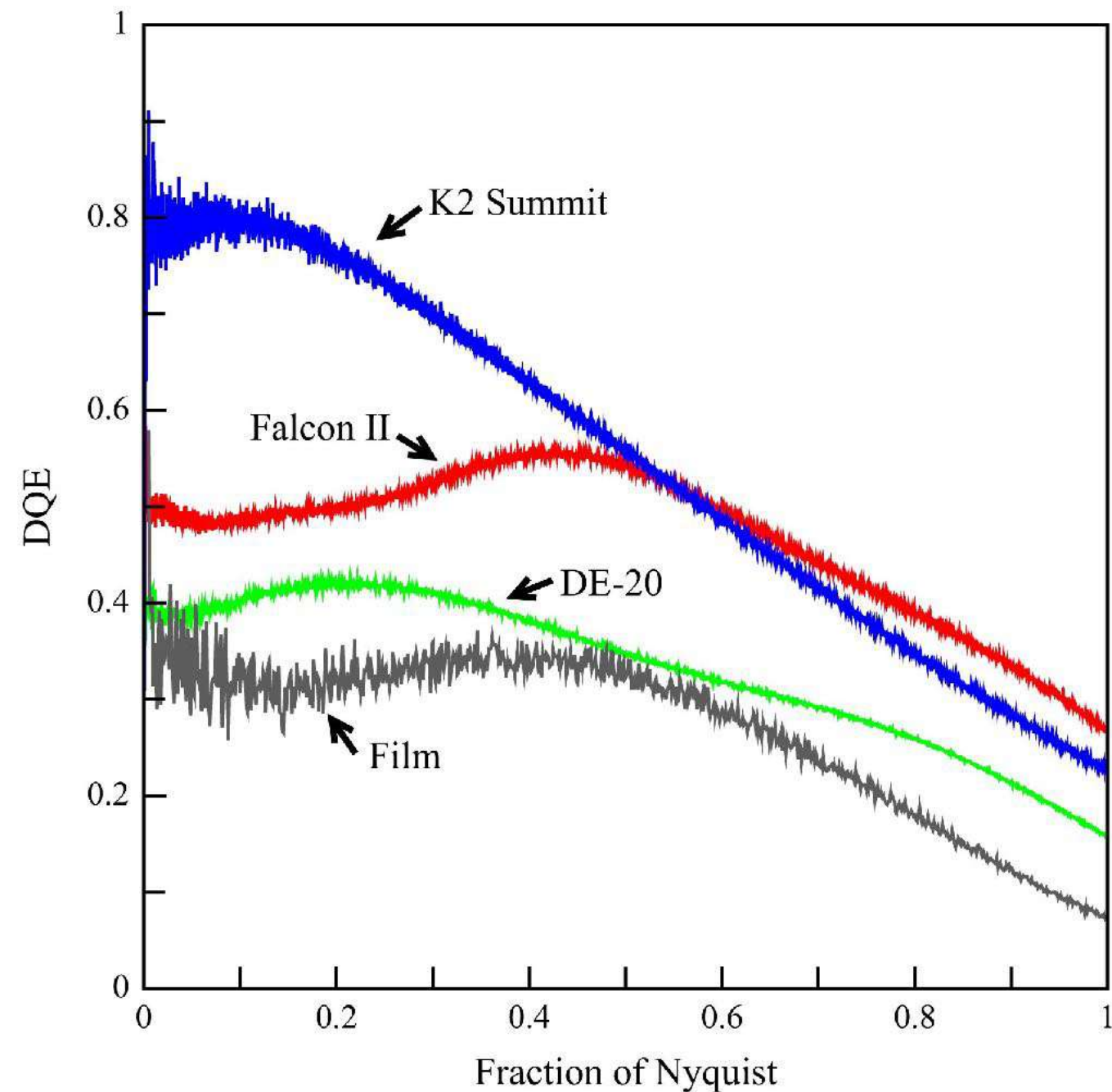
Development of high-resolution cryo-EM

- **Vitrification** of viruses and proteins (early 1980's)
 - Enabled the first cryo-EM images to be recorded of individual protein complexes
 - Proteins no longer needed to be crystallized in order to be visualized
 - Proteins were much less sensitive to beam-induced radiation damage
 - Resolution was drastically improved

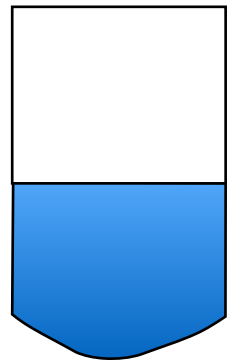


Direct electron detectors

- Traditionally, EM images were recorded on film negatives and then digitized with a scanner
- In 2013, **direct electron detectors** were commercialized
 - Massively increased signal-to-noise ratio
 - Enabled data collection to be automated, increasing the number of images that can be obtained
- DEDs sparked the current era of rapid cryo-EM structure determination



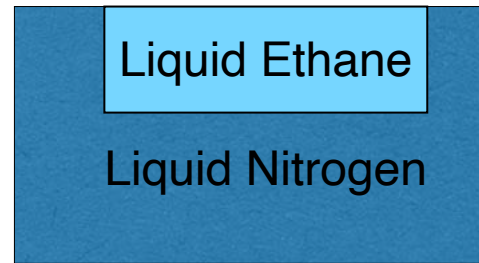
Cryo-EM single particle analysis



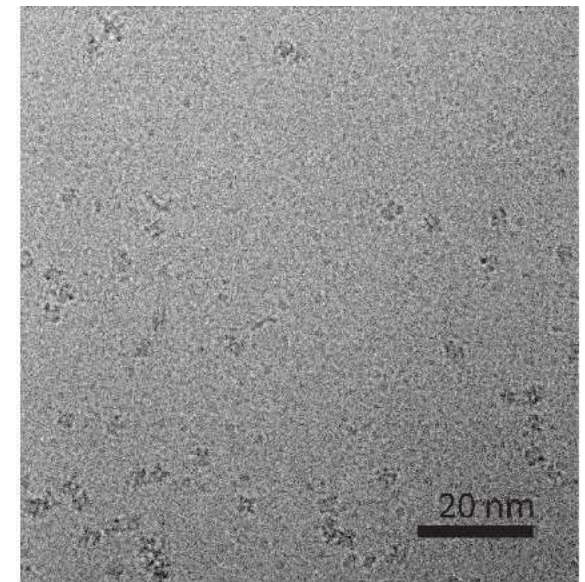
Purify sample



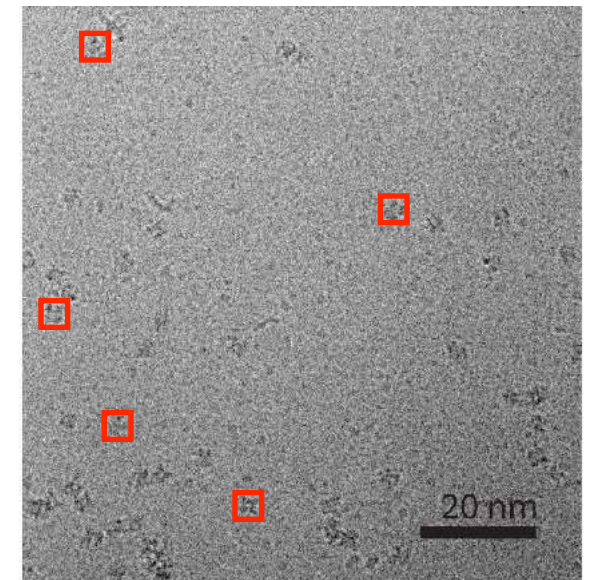
Holey carbon
copper EM grid



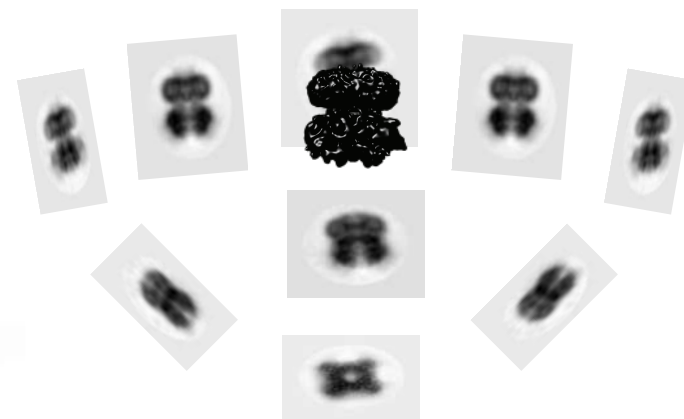
Freeze sample



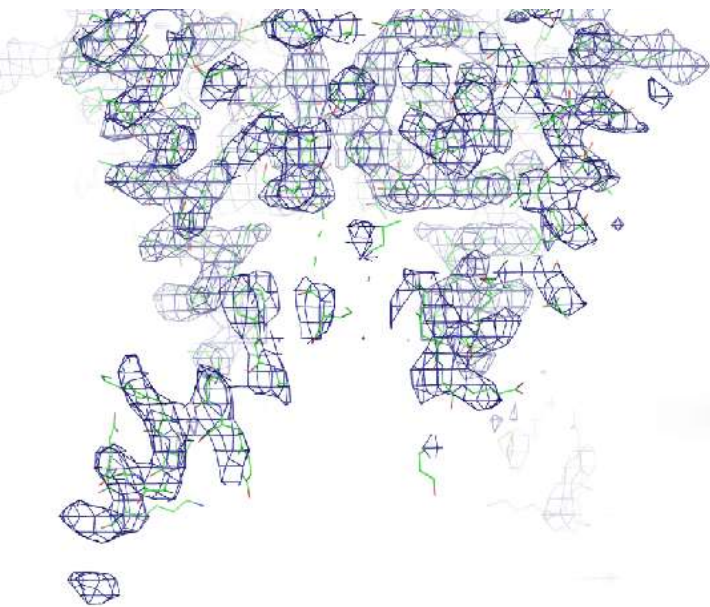
Collect images



Select particles



Reconstruct 3D volume
by projection matching



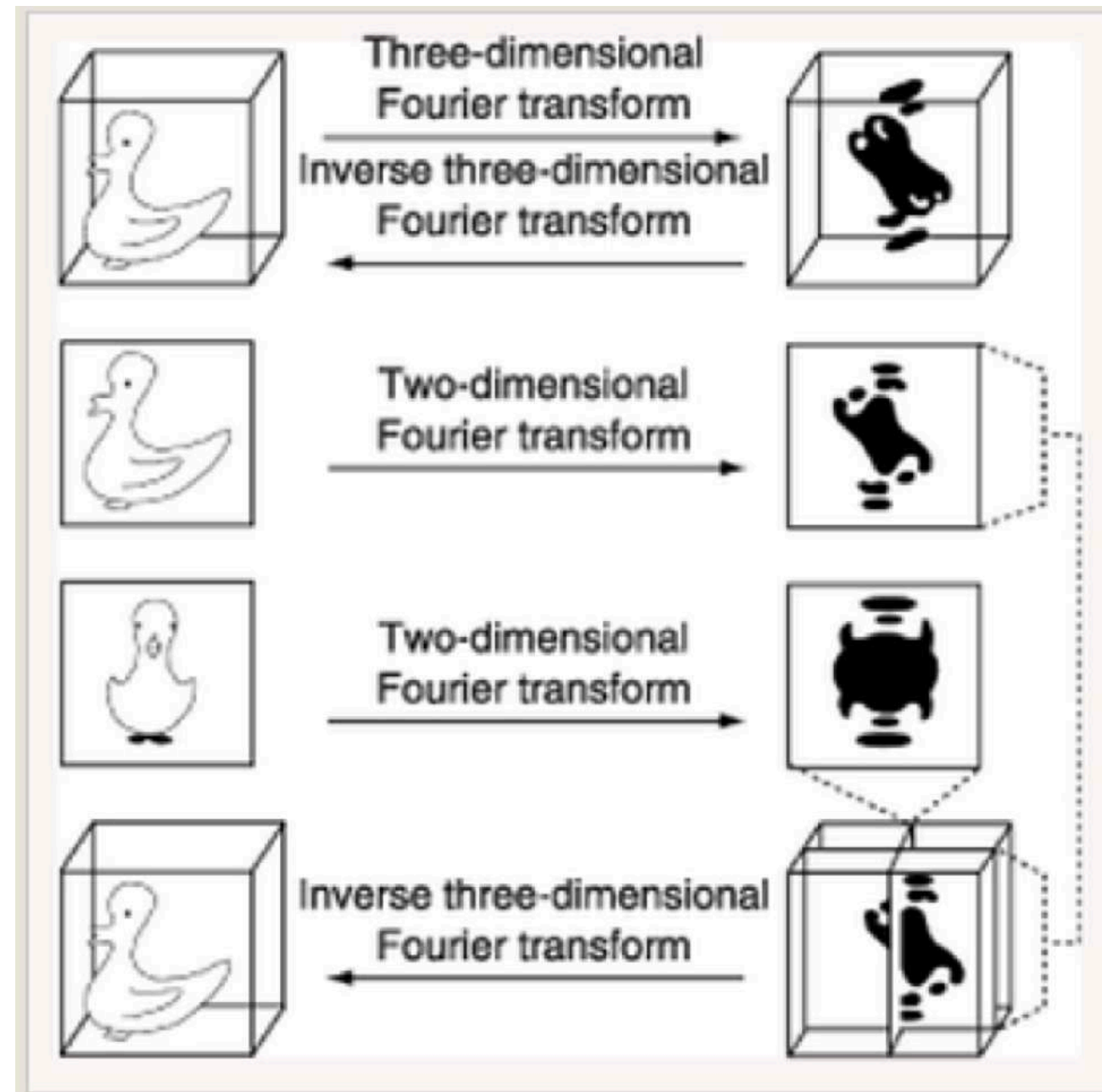
Build and refine
atomic model

Advantages of cryo-EM

- Crystallization is not necessary
 - Growing well-ordered protein crystals can require years of work
 - Large protein complexes are particularly difficult to crystallize
- Immediate feedback regarding sample quality
 - Particles can be directly viewed in the images
 - Reconstructions can be calculated on the fly
- Compositional and conformational heterogeneities can be overcome by particle classification
 - Proteins rarely adopt a single conformation in physiological conditions
 - Classification can resolve protein dynamics

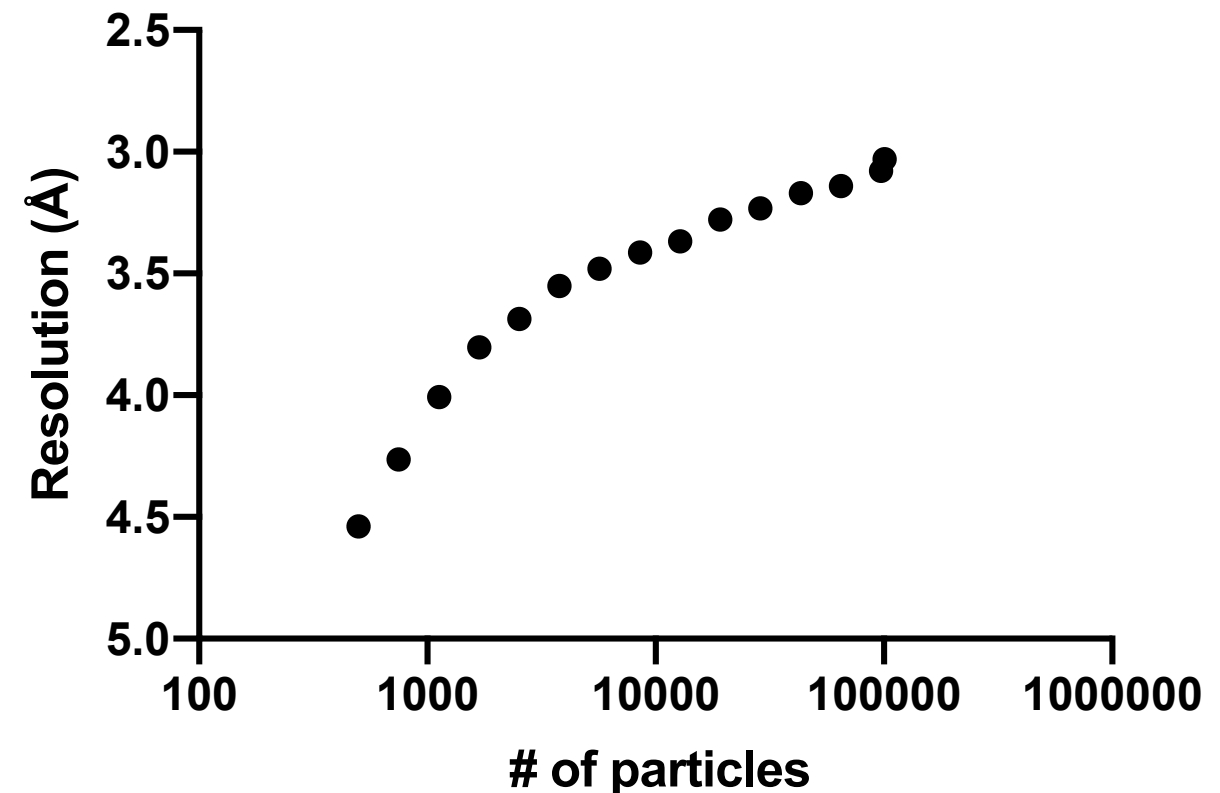
Cryo-EM image reconstruction

- How to obtain 3D dimensional information from 2D images?
- We rely on the Central section theorem which states that a 2D Fourier transform of the 2D projection of a 3D density is equal to the central section of the 3D Fourier transform of the density perpendicular to the direction of the projection
- By imaging many different projection of the sample, we can sample the entire 3D Fourier transform of the density
- Using an inverse Fourier transform, we can then visualize the 3D volume of the object



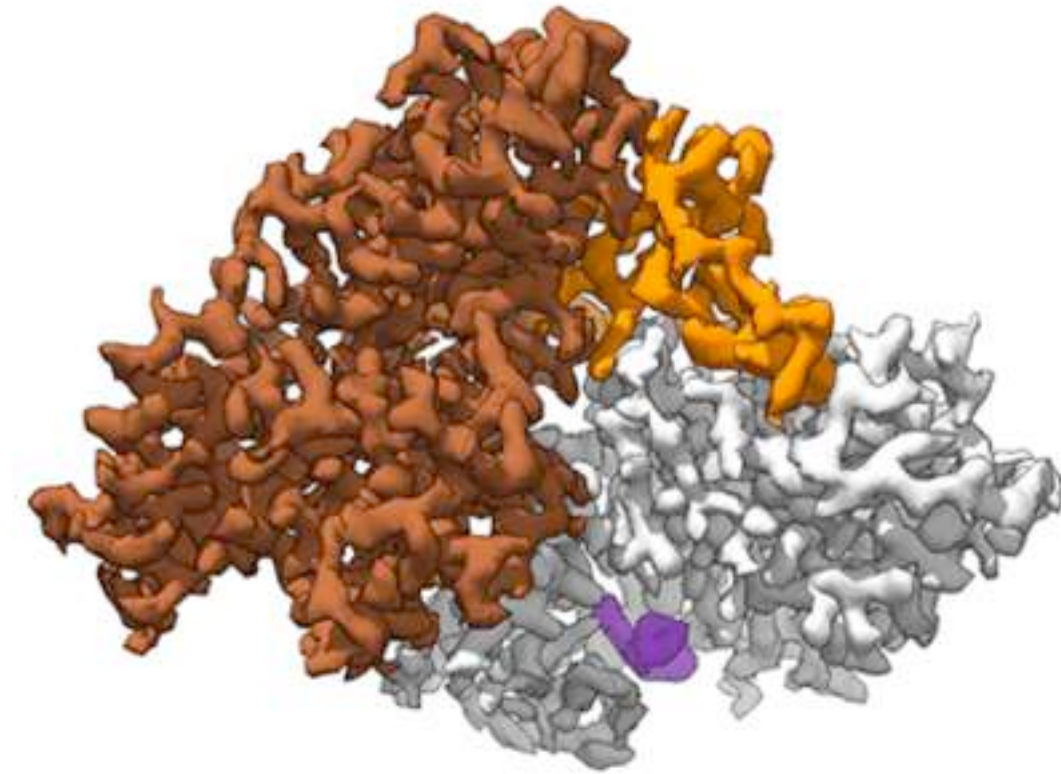
Why millions of particles?

- Biological specimen are extremely sensitive to radiation damage - electron beam can break bonds
- To minimize the effects of radiation damage, a very low number of electrons is used to image the specimen
- Signal-to-noise ratio of cryo-EM data is low, especially at high resolution
- Averaging techniques can alleviate the low signal-to-noise issue because the signal is invariant from particle to particle while the noise is random and will vary between particles



How to evaluate a cryo-EM map

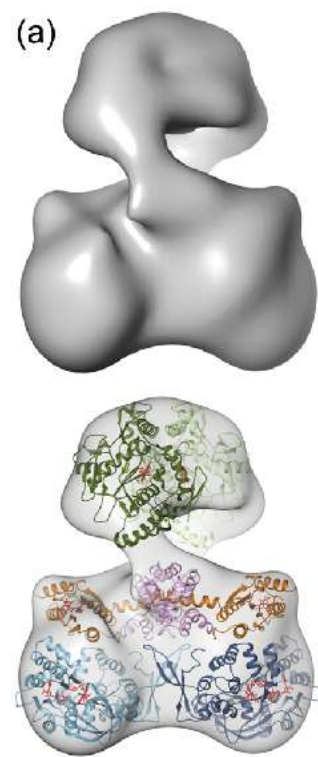
- How was the map validated?
 - What evidence supports the hypothesis that the map faithfully describes the structure of the sample?
 - What methods can you use to provide additional evidence?
- Map resolution and interpretation
 - What is the resolution of the map?
 - How uniform is the resolution?
 - Do the features of the map (i.e. secondary structure, side chains) correspond to the resolution?
 - Was the model properly positioned in the structure and how was it validated?
- Sample heterogeneity
 - Does the sample contain a mixture of different proteins and/or conformations?
 - What can you learn from the heterogeneity of the sample?



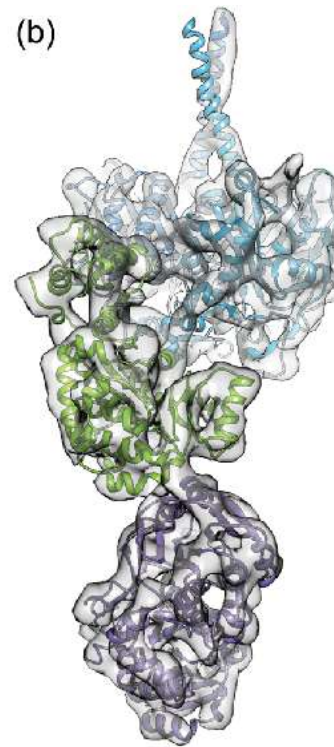
EMD-12042
2.5 Å

Resolution - what does it mean?

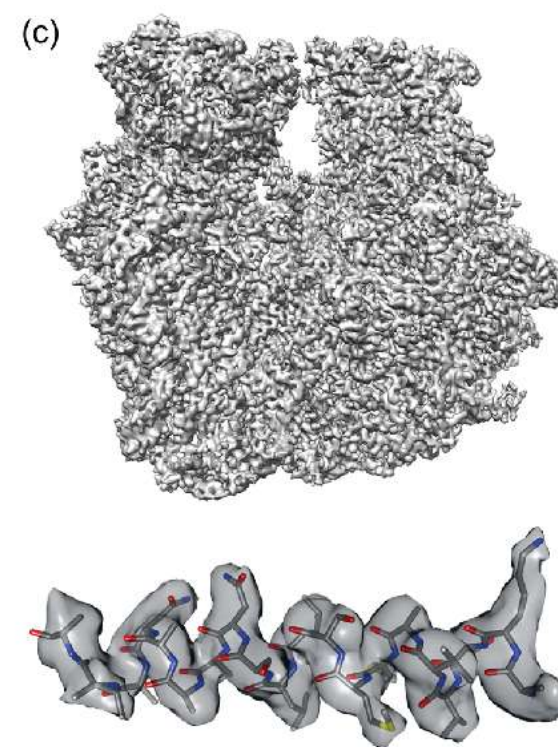
- Resolution corresponds to the distance between two objects that can be reliably separated in the map
- In EM, resolution is estimated by the Fourier shell correlation (FSC)
- FSC is determined by comparing the cross correlation between two independently calculated maps in Fourier space resolution shells
- Practically, this is done by separating a data set (10,000 - 1,000,000 particle images) into two halves and determining independent reconstructions of each half
- Different resolutions have different types of features that can be visualized in the density map



10-30 Å
Overall architecture.
Low resolution modeling
of available structures
following constraints.



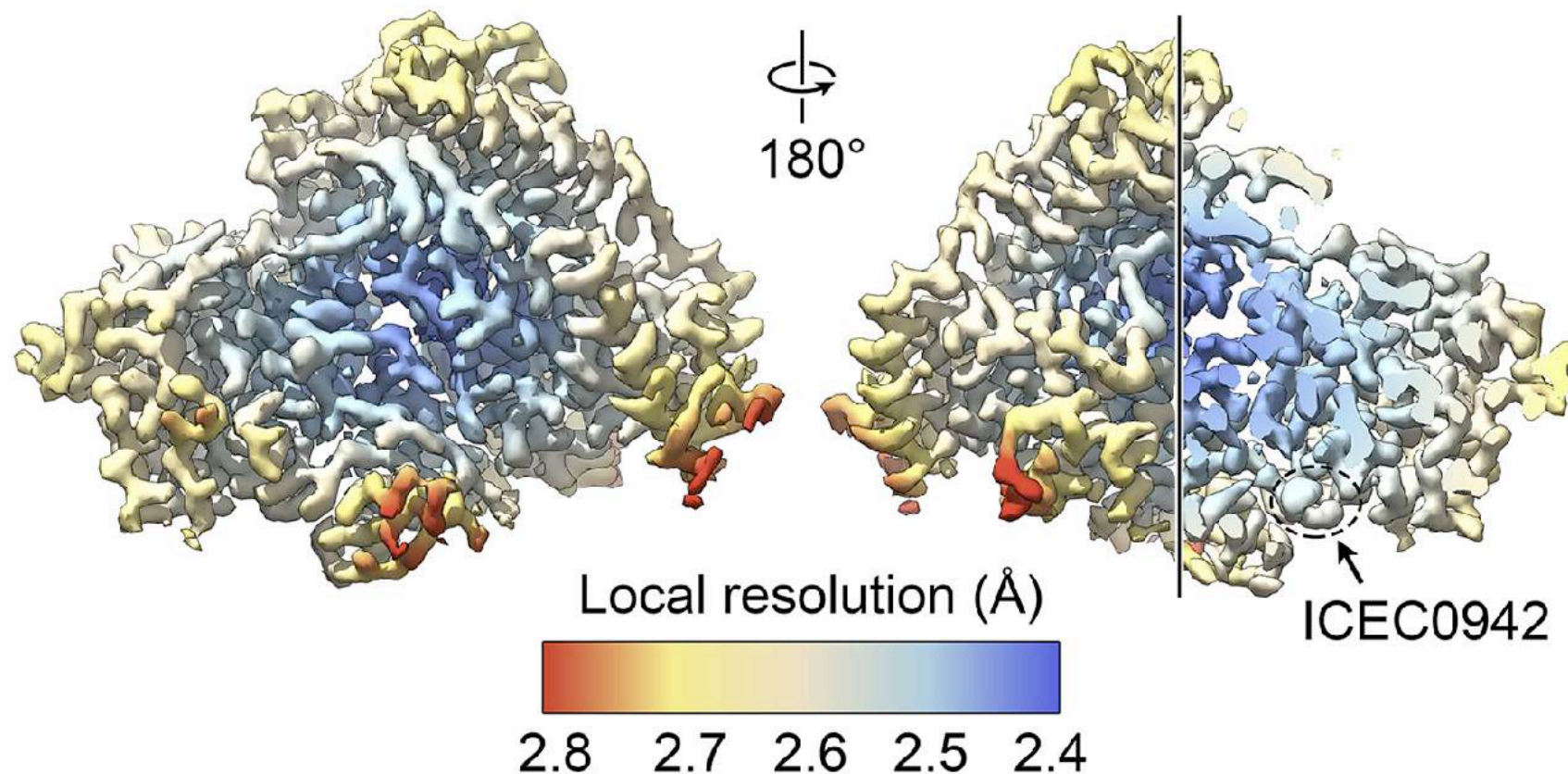
5-10 Å
Secondary structure.
Accurate rigid body
docking or flexible fitting
of available structures.



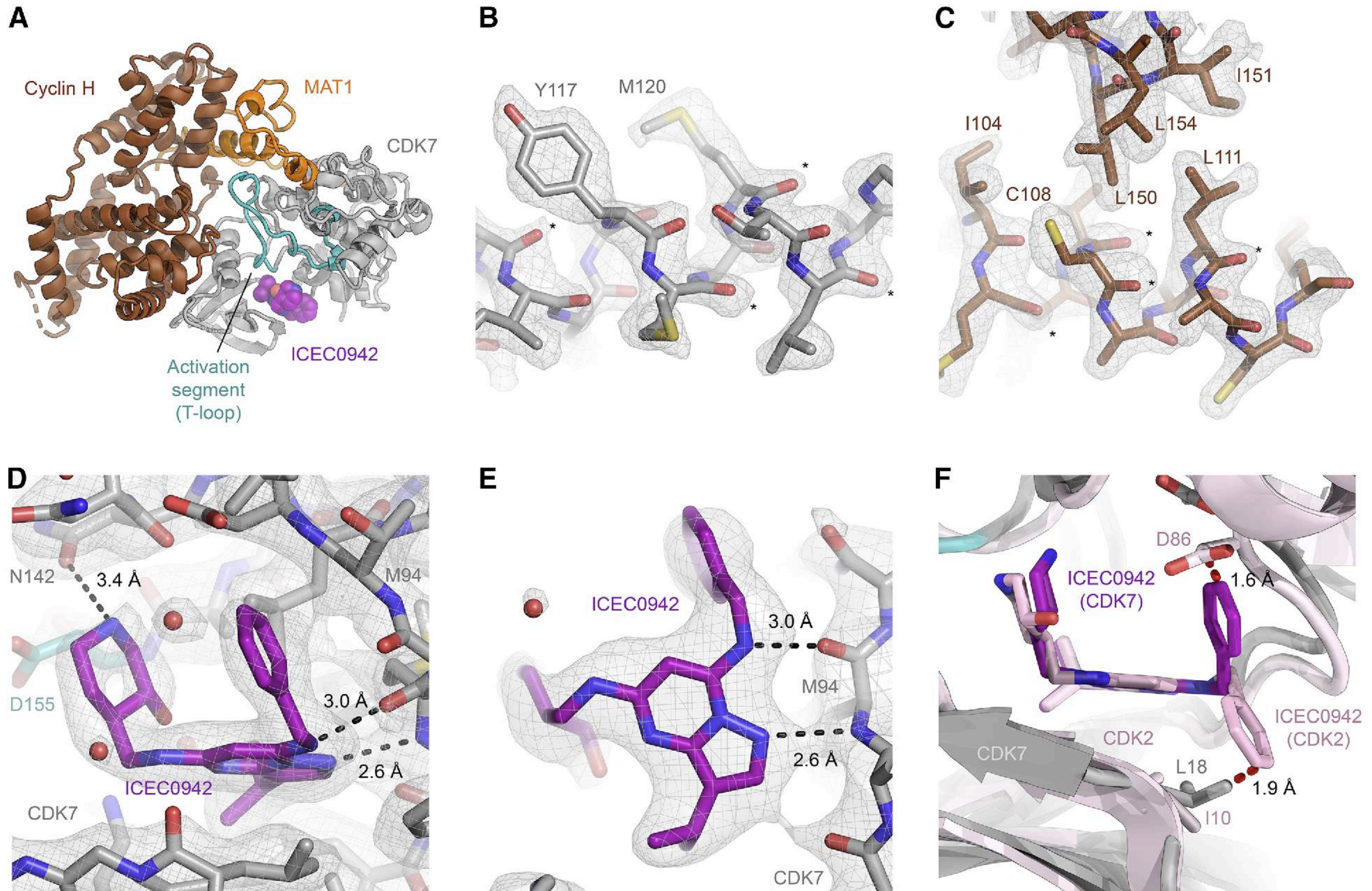
< 4 Å
Near atomic resolution.
Backbone tracing,
side chain modeling
and refinement.

Resolution - what does it mean?

- Resolution corresponds to the distance between two objects that can be reliably separated in the map
- In EM, resolution is estimated by the Fourier shell correlation (FSC)
- FSC is determined by comparing the cross correlation between two independently calculated maps in Fourier space resolution shells
- Practically, this is done by separating a data set (10,000 - 1,000,000 particle images) into two halves and determining independent reconstructions of each half
- Different resolutions have different types of features that can be visualized in the density map

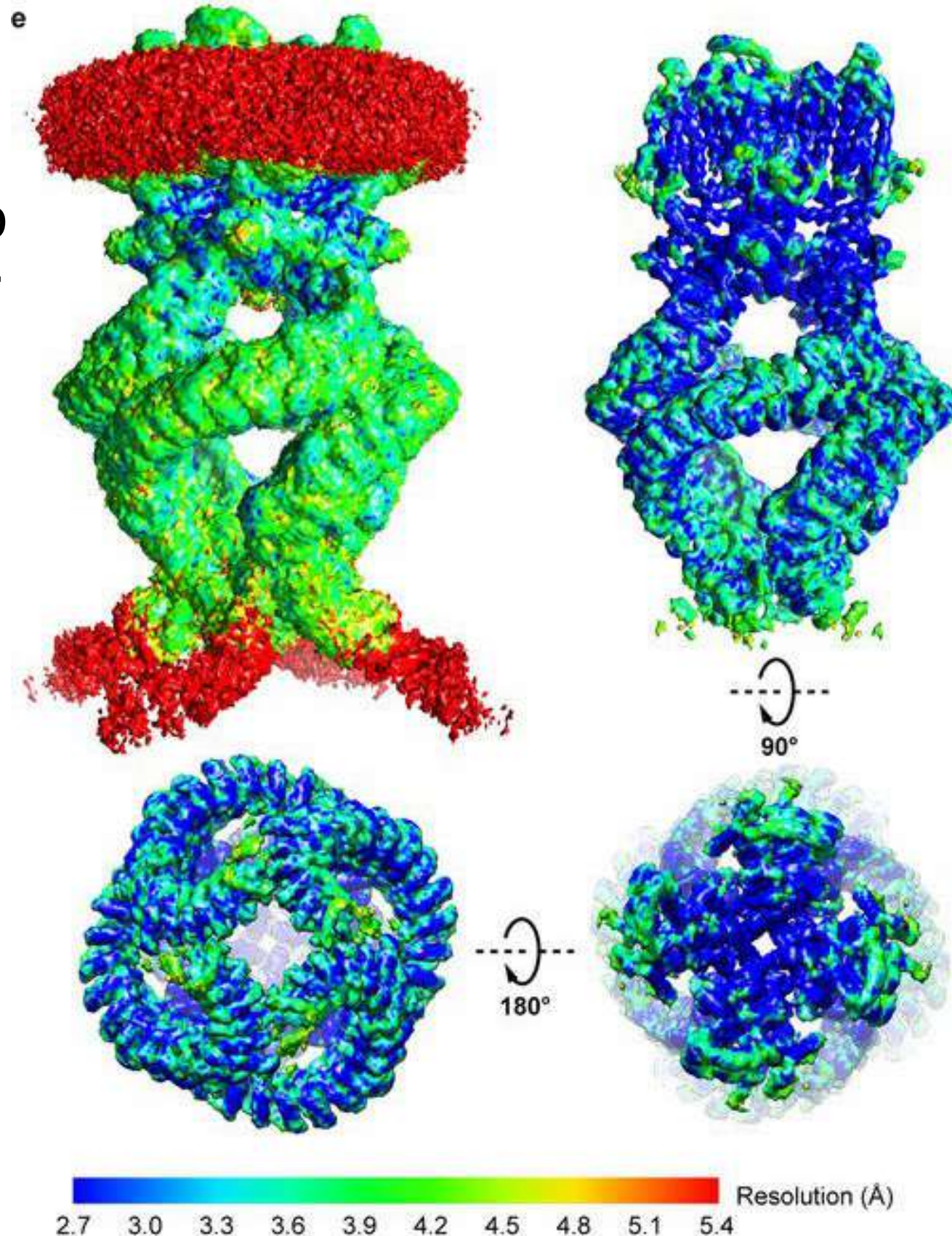


Resolution - what does it mean?



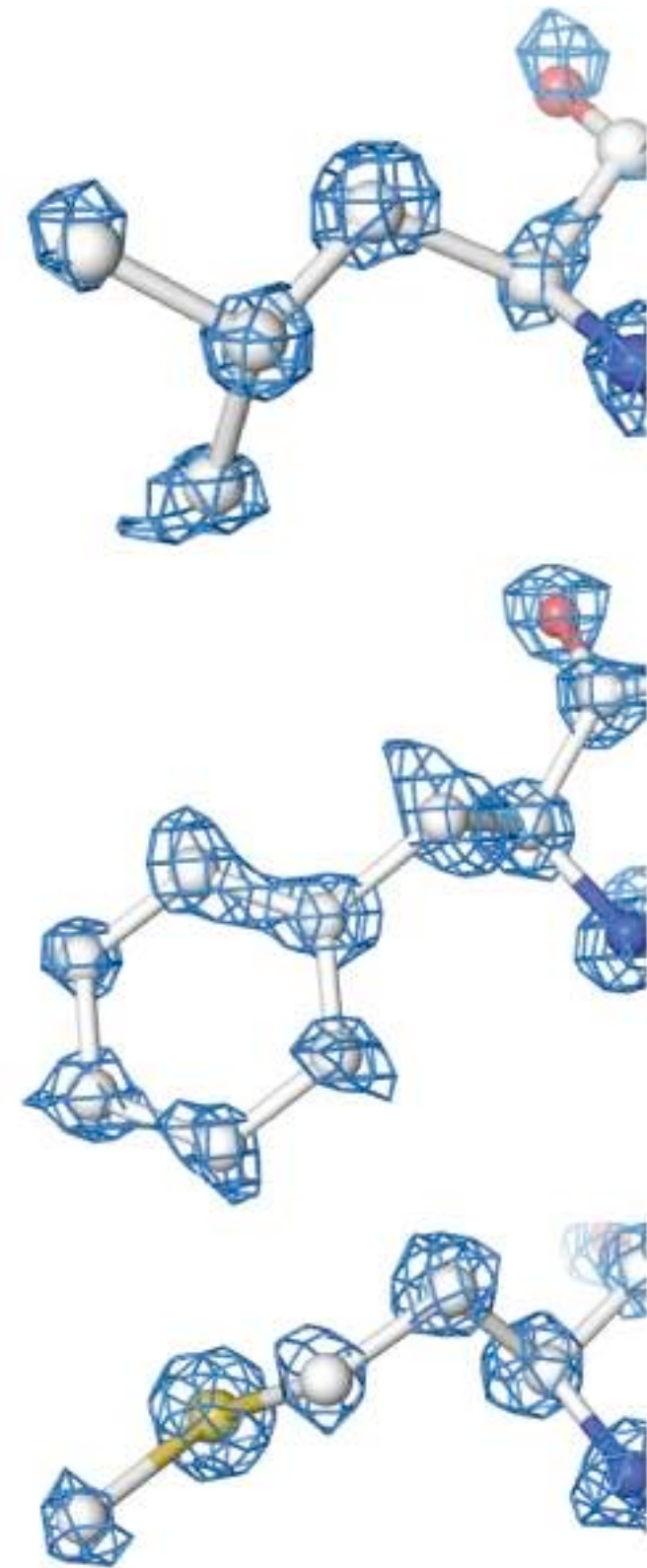
Resolution - what does it mean?

- Overall resolution does not equal local details
- Quality can vary greatly within a map and care should be taken to not over interpret poorly ordered domains
- Different resolutions have different types of features that can be visualized in the density map
 - 7 Å - alpha-helices
 - 4.5 Å - beta-strands
 - 4 Å - large side chains
 - > 3 Å water molecules
- Local resolution calculations can be performed to estimate local resolution
- Density slices can also be extremely informative for evaluating local map quality

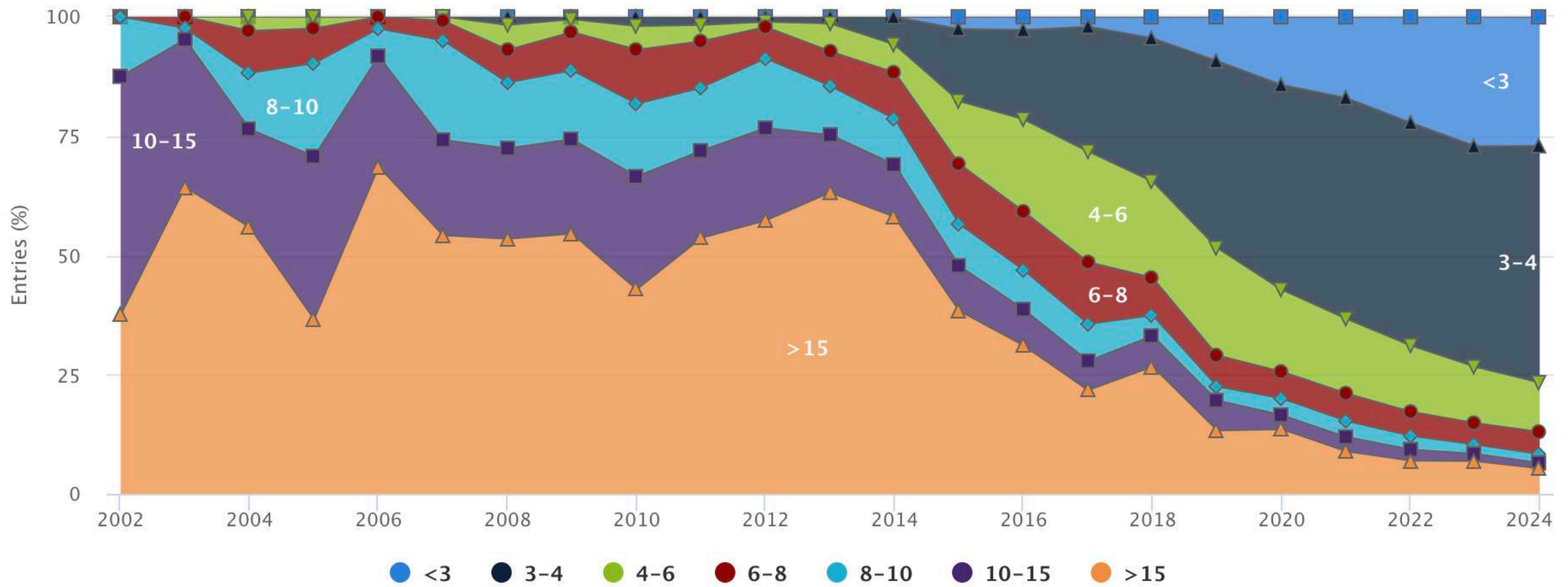


Resolution - what does it mean?

- Overall resolution does not equal local details
- Quality can vary greatly within a map and care should be taken to not over interpret poorly ordered domains
- Different resolutions have different types of features that can be visualized in the density map
 - 7 Å - alpha-helices
 - 4.5 Å - beta-strands
 - 4 Å - large side chains
 - > 3 Å water molecules
 - 1.2 Å allows resolving individual atoms

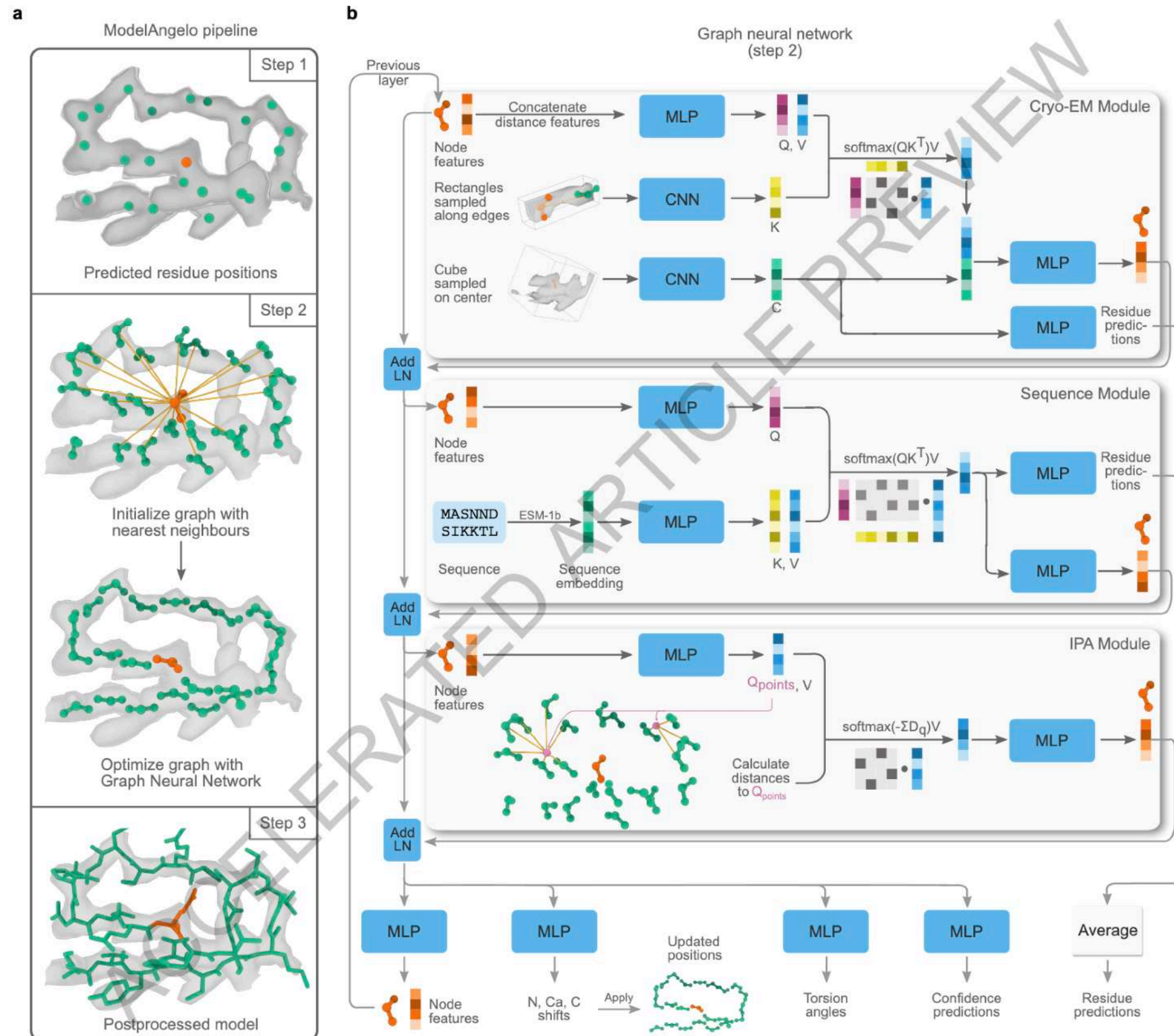


Resolution - what does it mean?



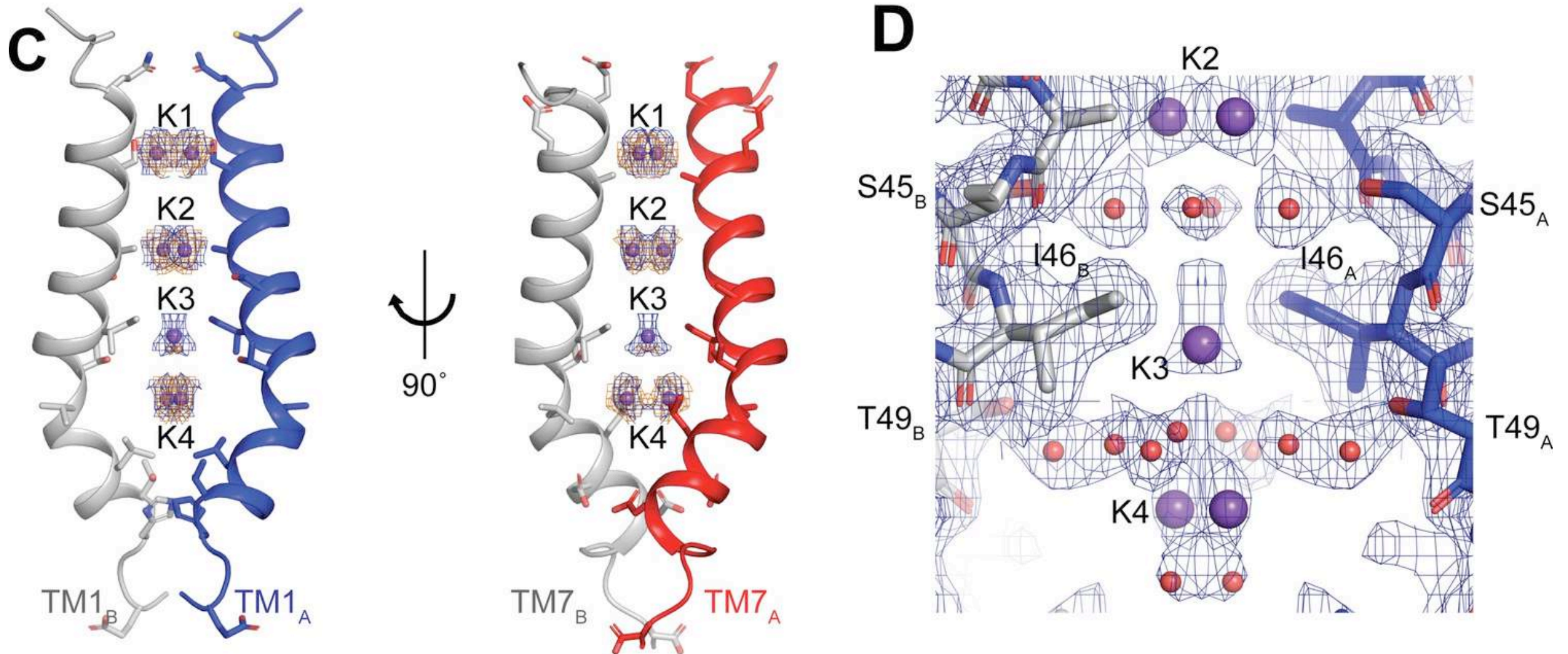
Map to Model

- Model building has been rapidly accelerated
- Hand building of high-resolution structures has been replaced with automated approaches
- Modelangelo is one of them that can rapidly build into maps at better than 3.5 Å with high accuracy



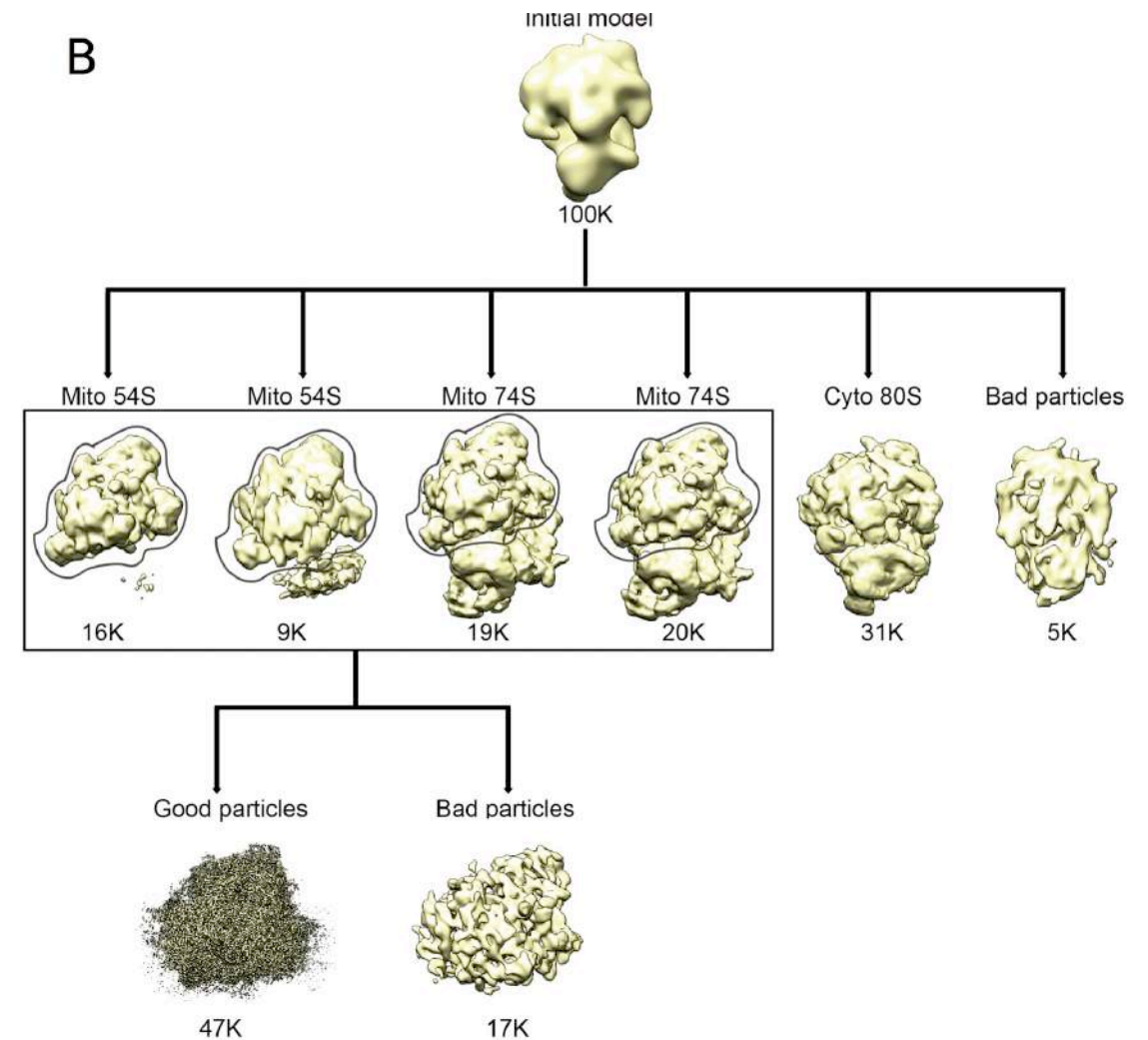
Is all density in a map is the same?

- Identifying non-protein densities in cryo-EM is quite challenging as there are few tools that allow accurate discrimination between atoms in density maps.
- In some circumstances one can replace an atom with a similarly structured atom to distinguish ion binding sites



Sample heterogeneity

- There are multiple sources of heterogeneity in sample preparation
 - Compositional heterogeneity - mixture of different components or mixtures with varying subunit stoichiometry
 - Structural heterogeneity - domains of the specimen can adopt multiple conformations
 - In some cases, both types of heterogeneity exist within a single sample
- These will degrade the resolution of reconstructions if not sorted computationally, but can provide insights into function of the specimen

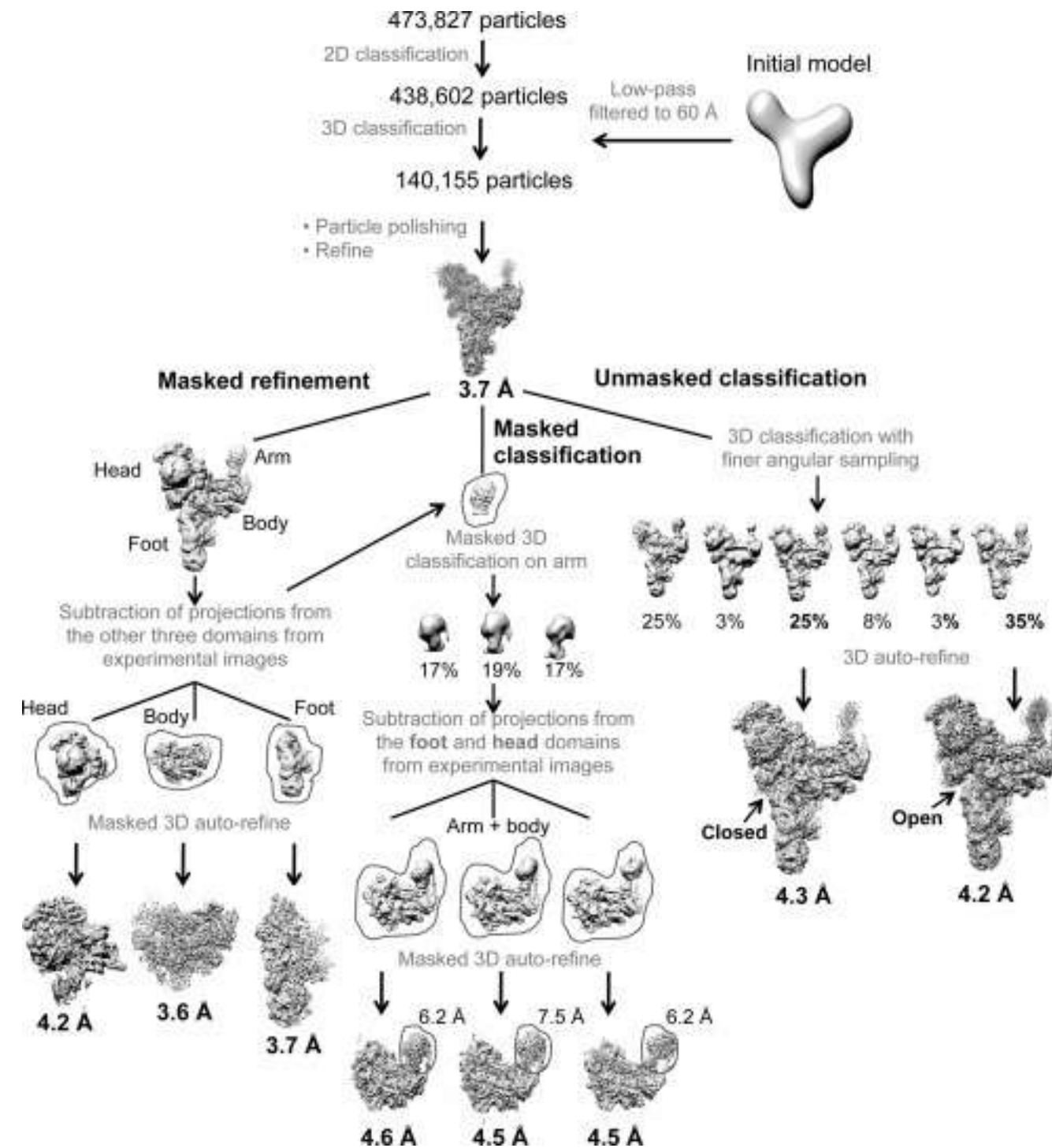


Overcoming heterogeneity - biochemistry

- Optimizing biochemistry can often help to alleviate heterogeneity and is generally the best place start to improve sample quality
 - Improvements in sample purification can reduce compositional heterogeneity by obtaining a more uniform starting sample
 - Structural heterogeneity can be minimized by altering purification conditions (i.e. presence of activating or inhibiting ligands, different pH or salt conditions)
 - Construct alterations can also reduce sample heterogeneity by removing flexible domains
- In some cases chemical cross-linking can helpful to reduce flexibility
 - Testing cross-linking reagents with different lengths and varying the concentration can be helpful to optimize conditions
 - However, it is essential that the chemically cross-linked structure be validated with a non-cross-structure to demonstrate the the cross-linking does not introduce artifactual protein-protein interactions

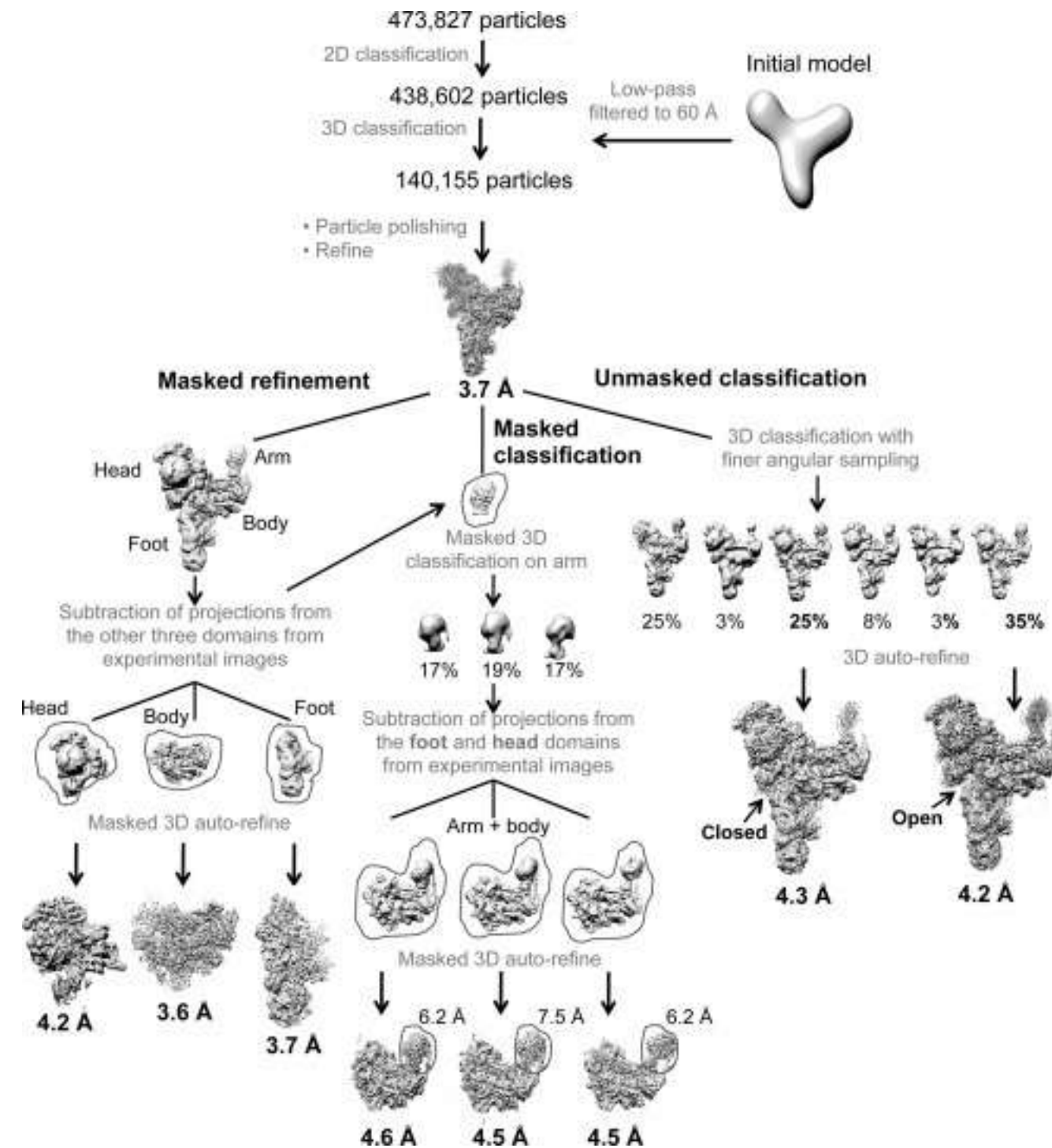
Overcoming heterogeneity - computation

- Heterogeneity may be unavoidable for some samples and must be dealt with computationally after image acquisition
- There are now several different software packages that sort and classify particles, allowing one to create “pure” subsets of the particles images
- The simplest approach is classify based upon the entire molecule, which works well with large conformational differences



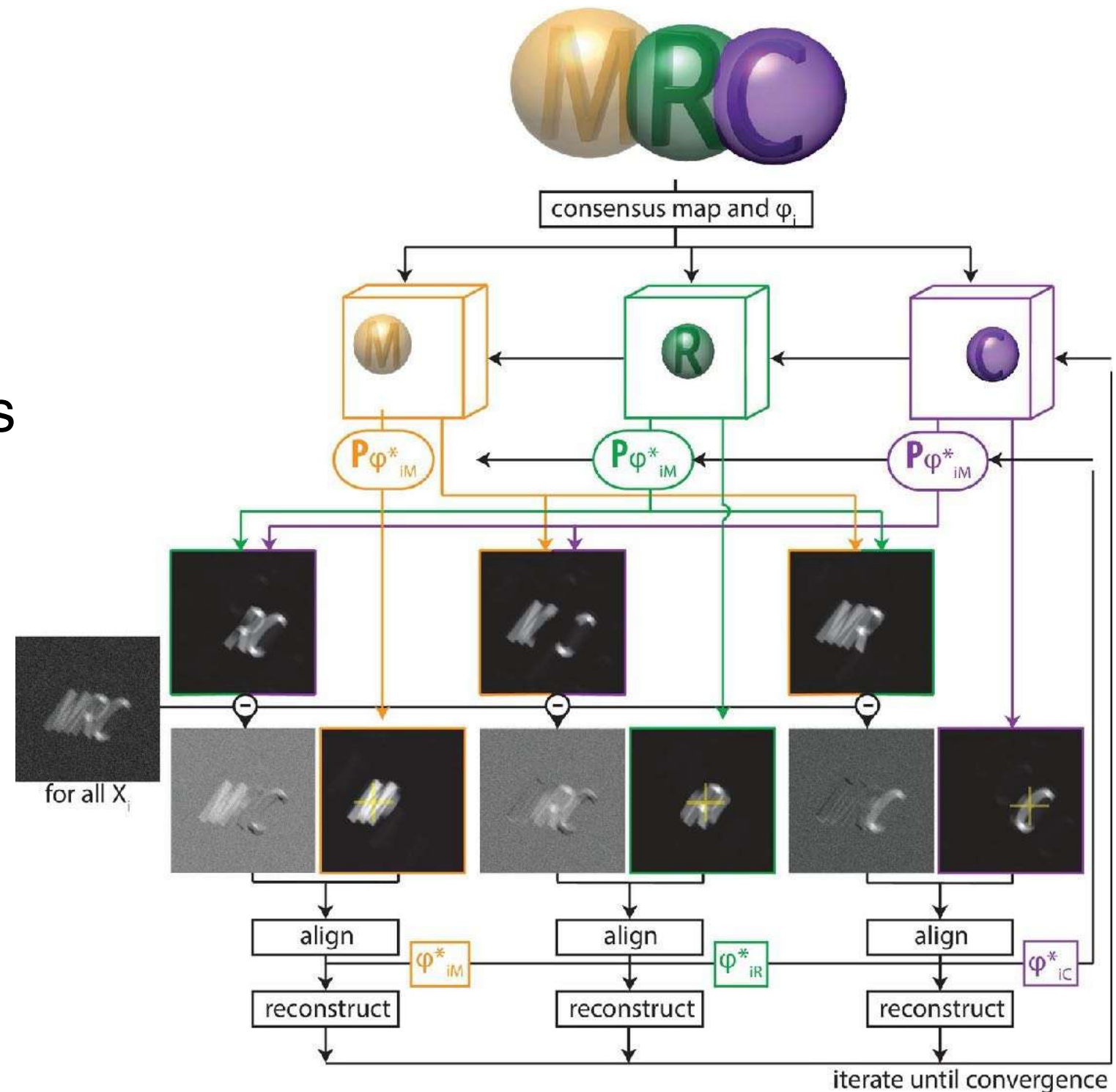
Overcoming heterogeneity - computation

- Classification can be enhanced through the use of masks
- A mask can be placed around the region of interest - allowing independent sorting of different domains
- This multi-classification approach is particularly powerful for samples that have multiple different types of movements
- Another modification to classification is the use of background subtraction prior to classification to reduce the signal of constant domains during classification



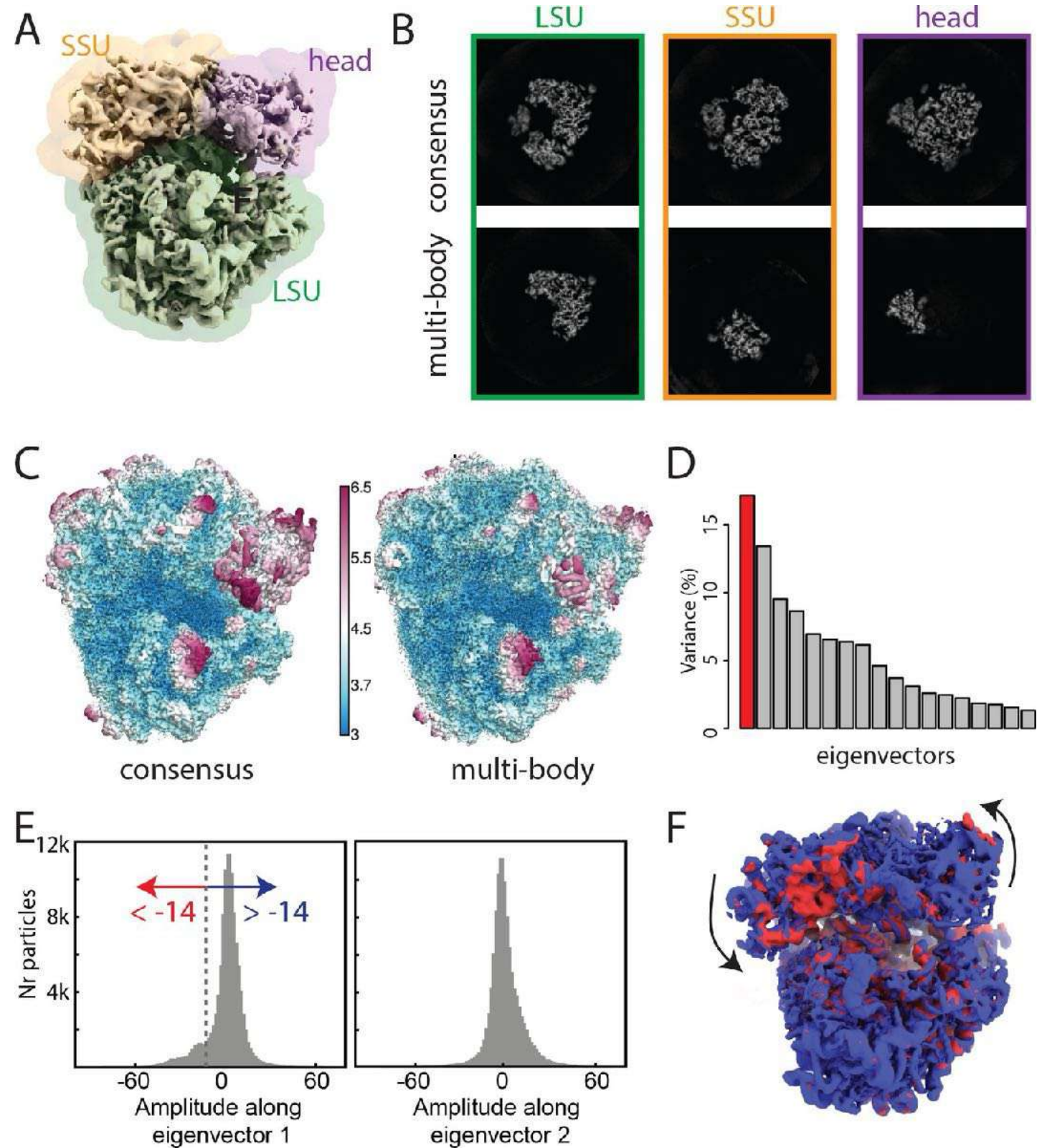
Overcoming heterogeneity - computation

- Relion has an automated procedure to apply masks based upon distinct flexible domains
- This approach is known as multi-body refinement
- It also determines the vectors of movement allowing an understanding of conformational dynamics across protein complexes



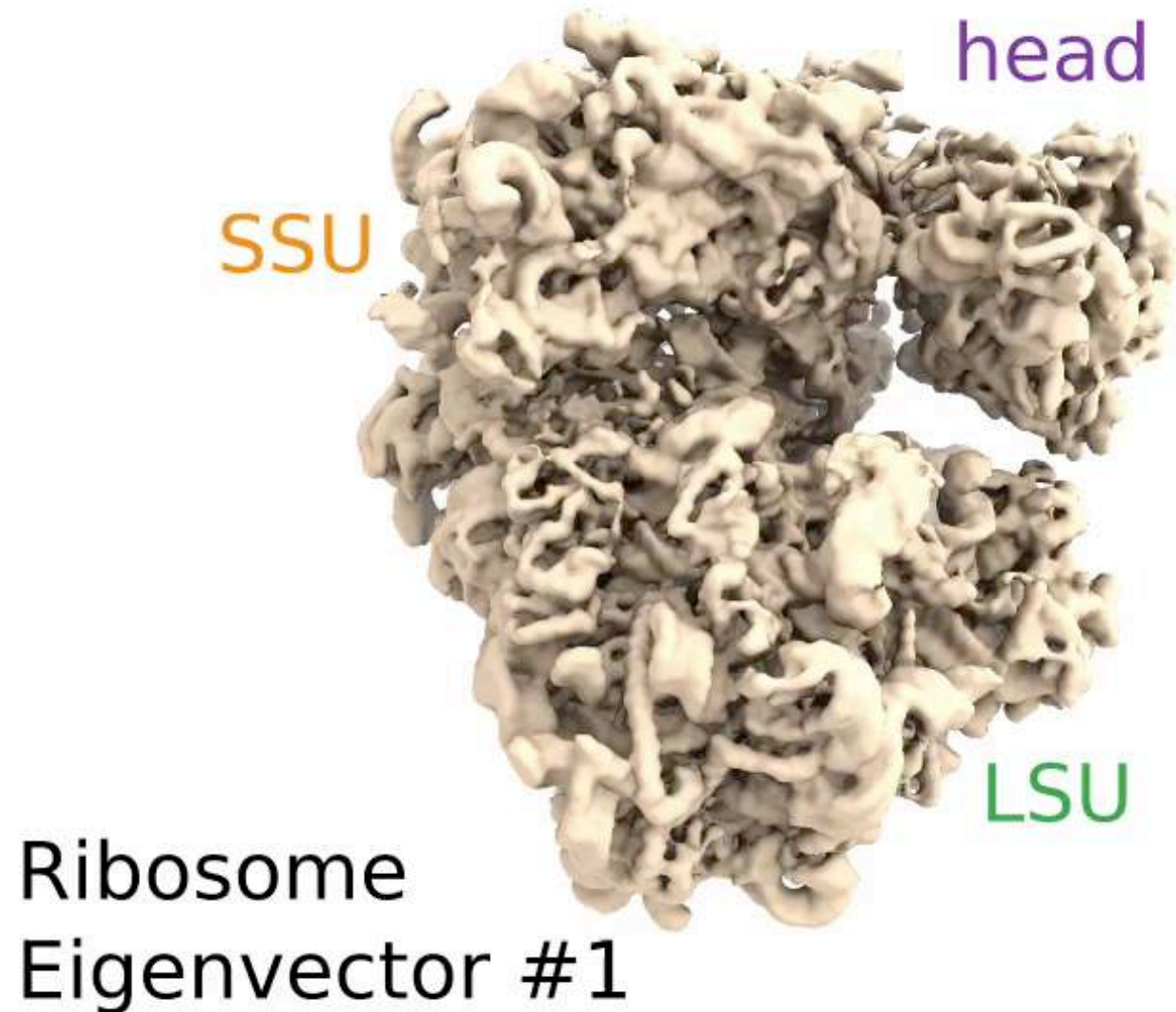
Overcoming heterogeneity - computation

- Relion has an automated procedure to apply masks based upon distinct flexible domains
- This approach is known as multi-body refinement
- It also determines the vectors of movement allowing an understanding of conformational dynamics across protein complexes



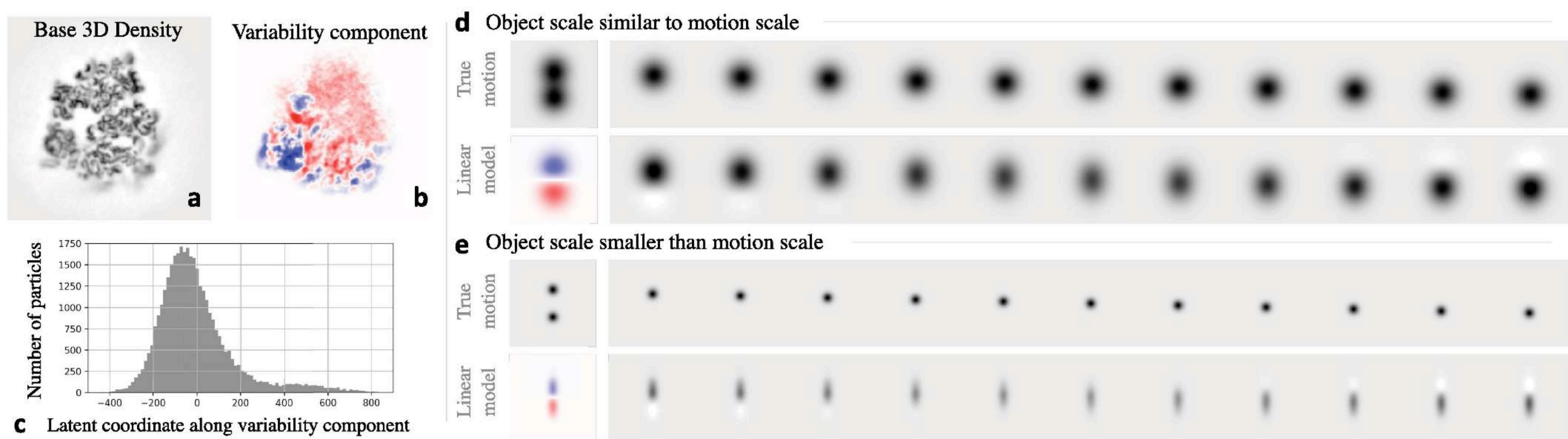
Overcoming heterogeneity - computation

- Relion has an automated procedure to apply masks based upon distinct flexible domains
- This approach is known as multi-body refinement
- It also determines the vectors of movement allowing an understanding of conformational dynamics across protein complexes
- Requires knowledge of the major domains of movement



Overcoming heterogeneity - computation

- Cryosparc has an alternative approach that they call 3D variability analysis
- Generates frames of a movie that reveal conformational dynamics along N principal components
- Requires classified particles (to some extent)

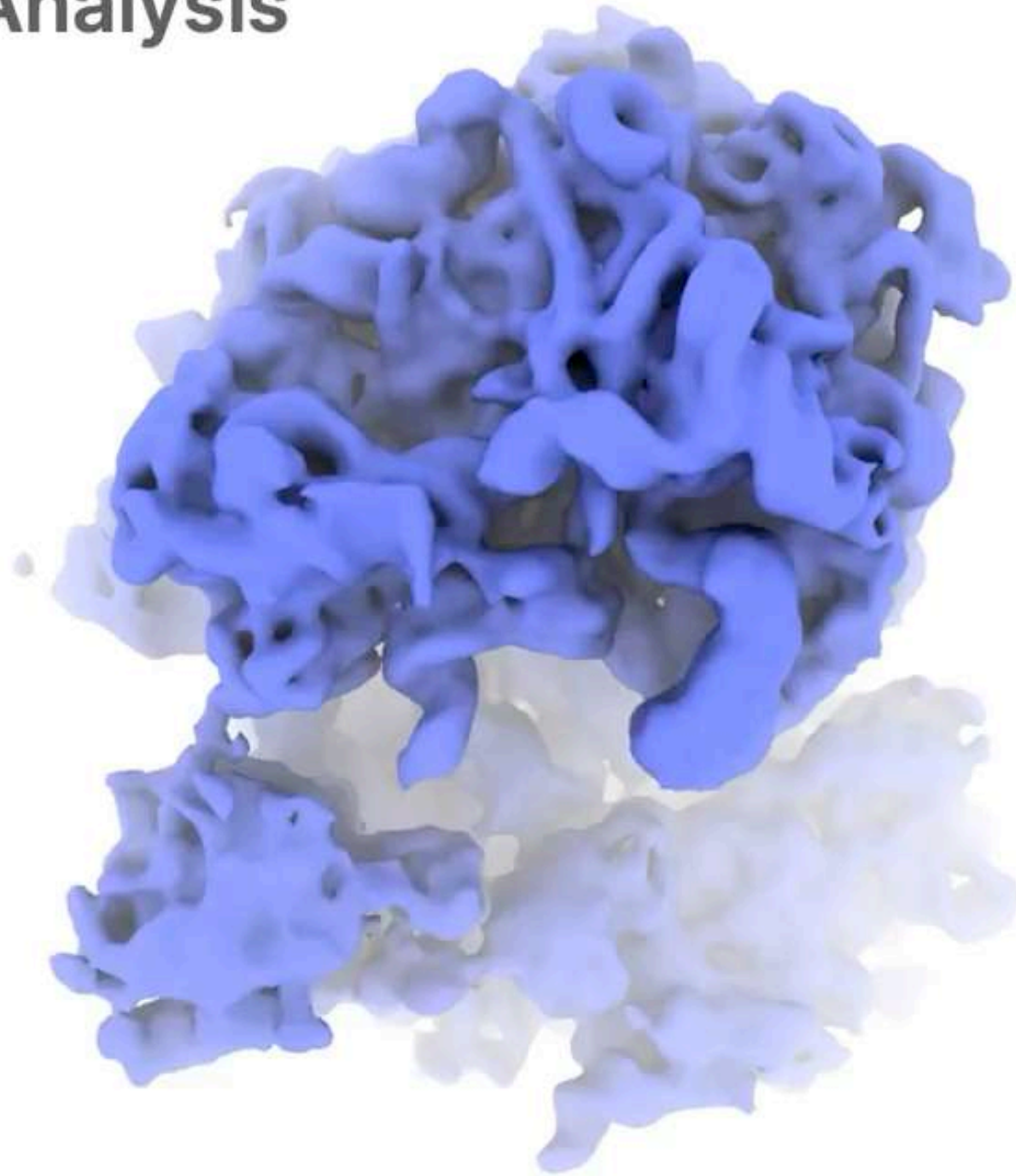


Overcoming heterogeneity - computation

3D Variability Analysis

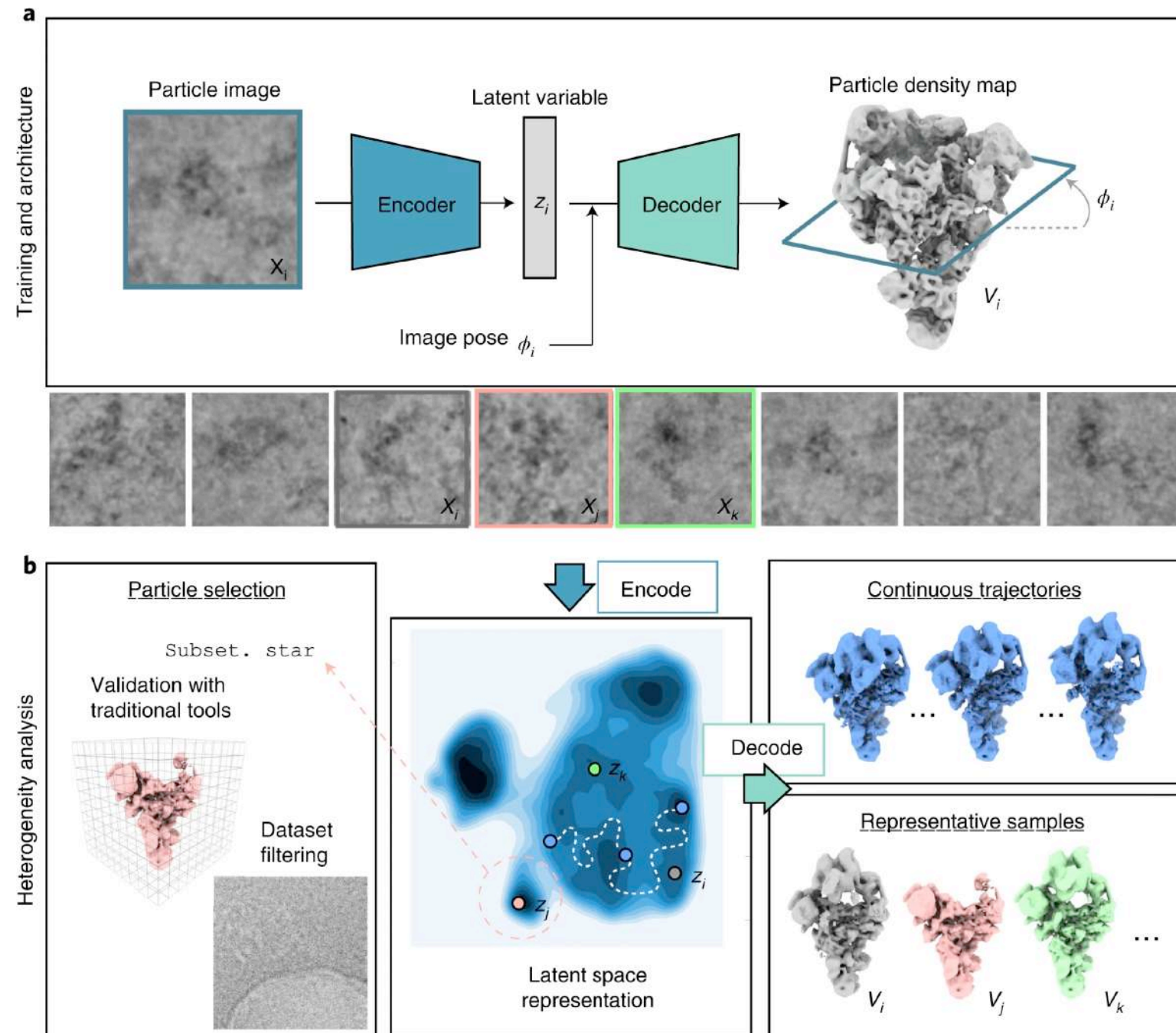
Pf80S Ribosome
(EMPIAR-10028)

Component 1



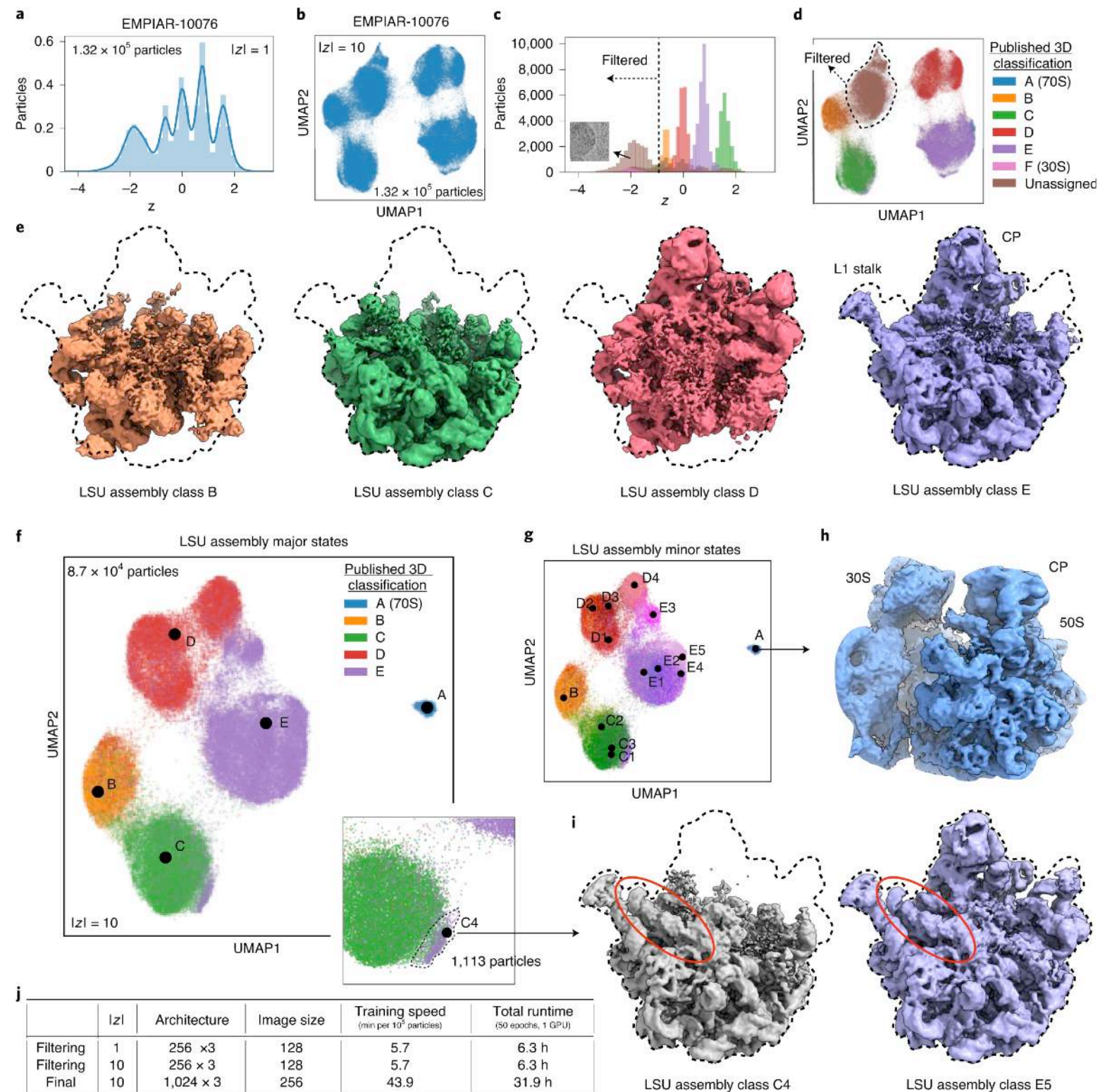
Overcoming heterogeneity - computation

- CryoDRGN - deep learning approach to visualize conformational dynamics
- Does not require classification, uses particles directly and sorts into a latent space representation
- Allows analysis of highly complex data sets



Overcoming heterogeneity - computation

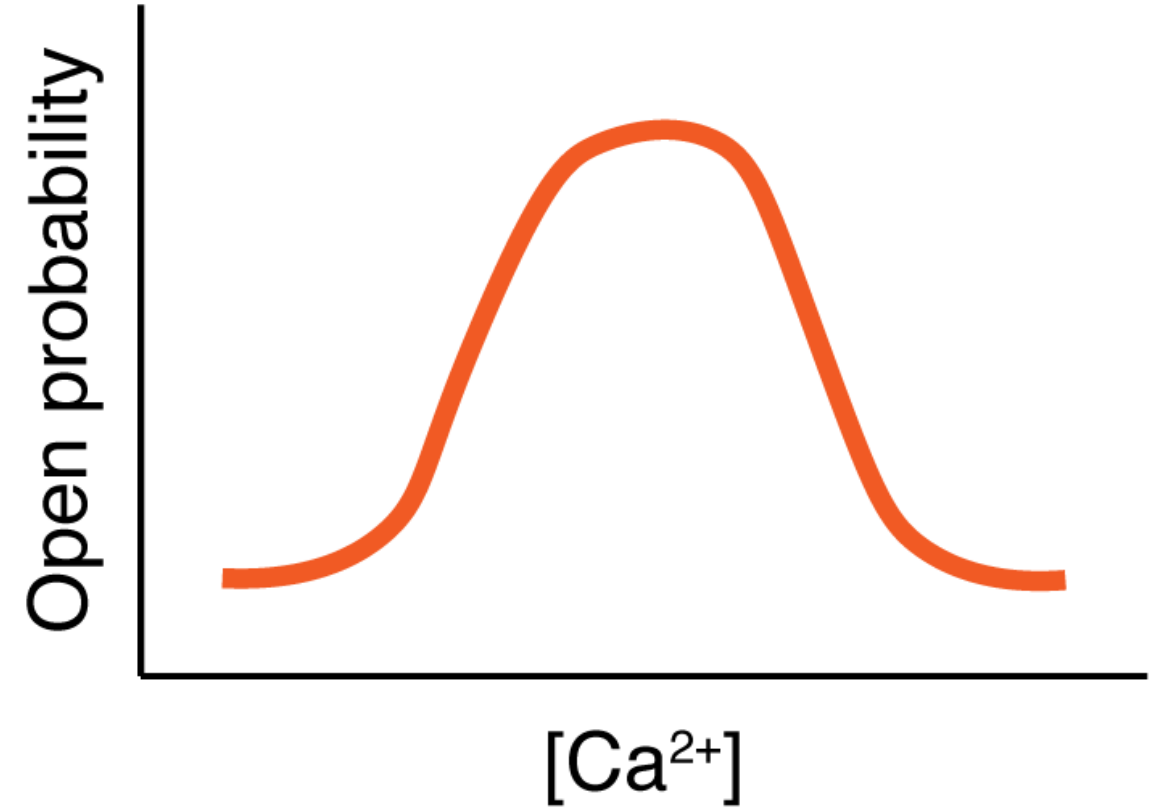
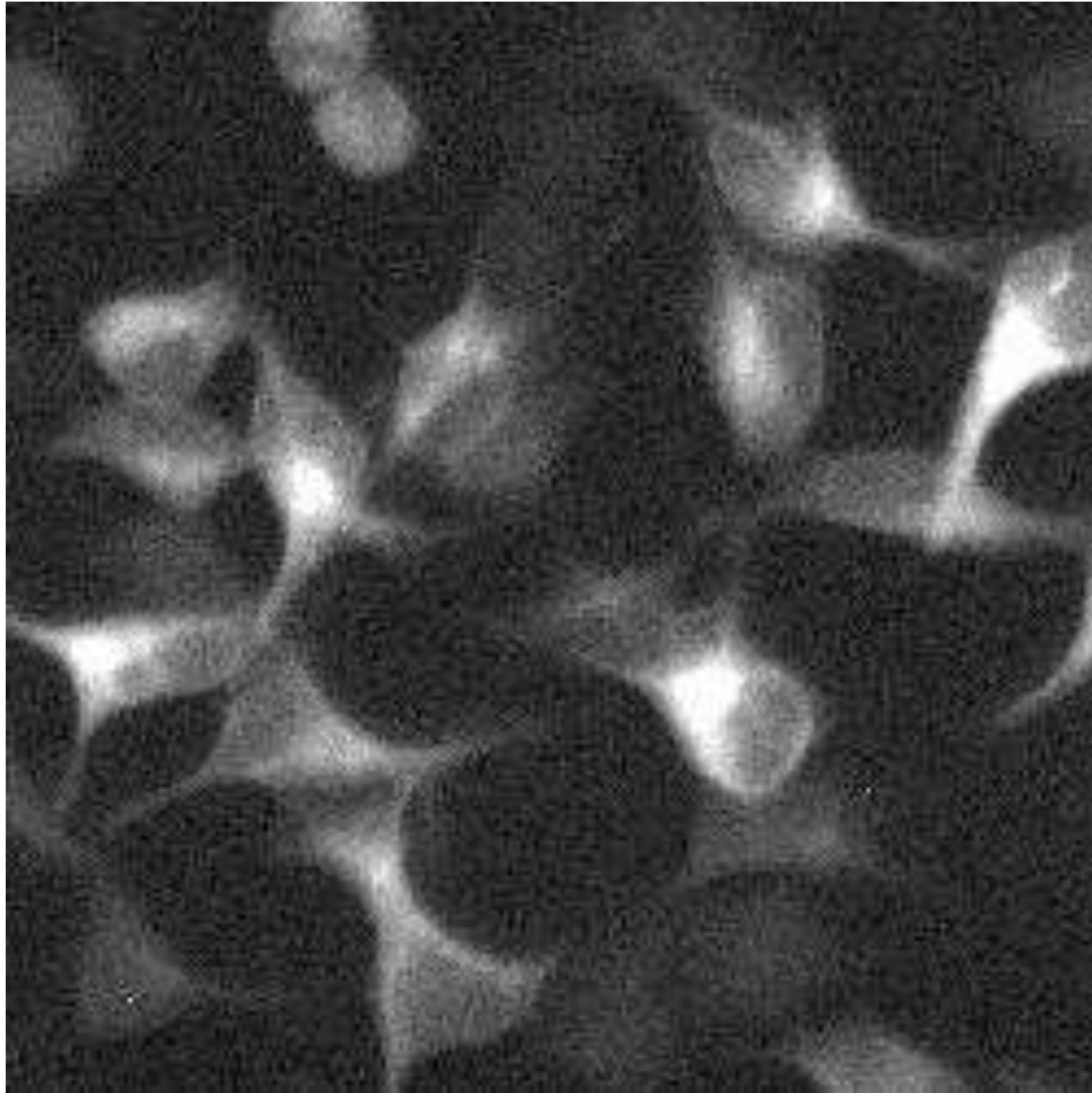
- CryoDRGN - deep learning approach to visualize conformational dynamics
- Does not require classification, uses particles directly and sorts into a latent space representation
- Allows analysis of highly complex data sets



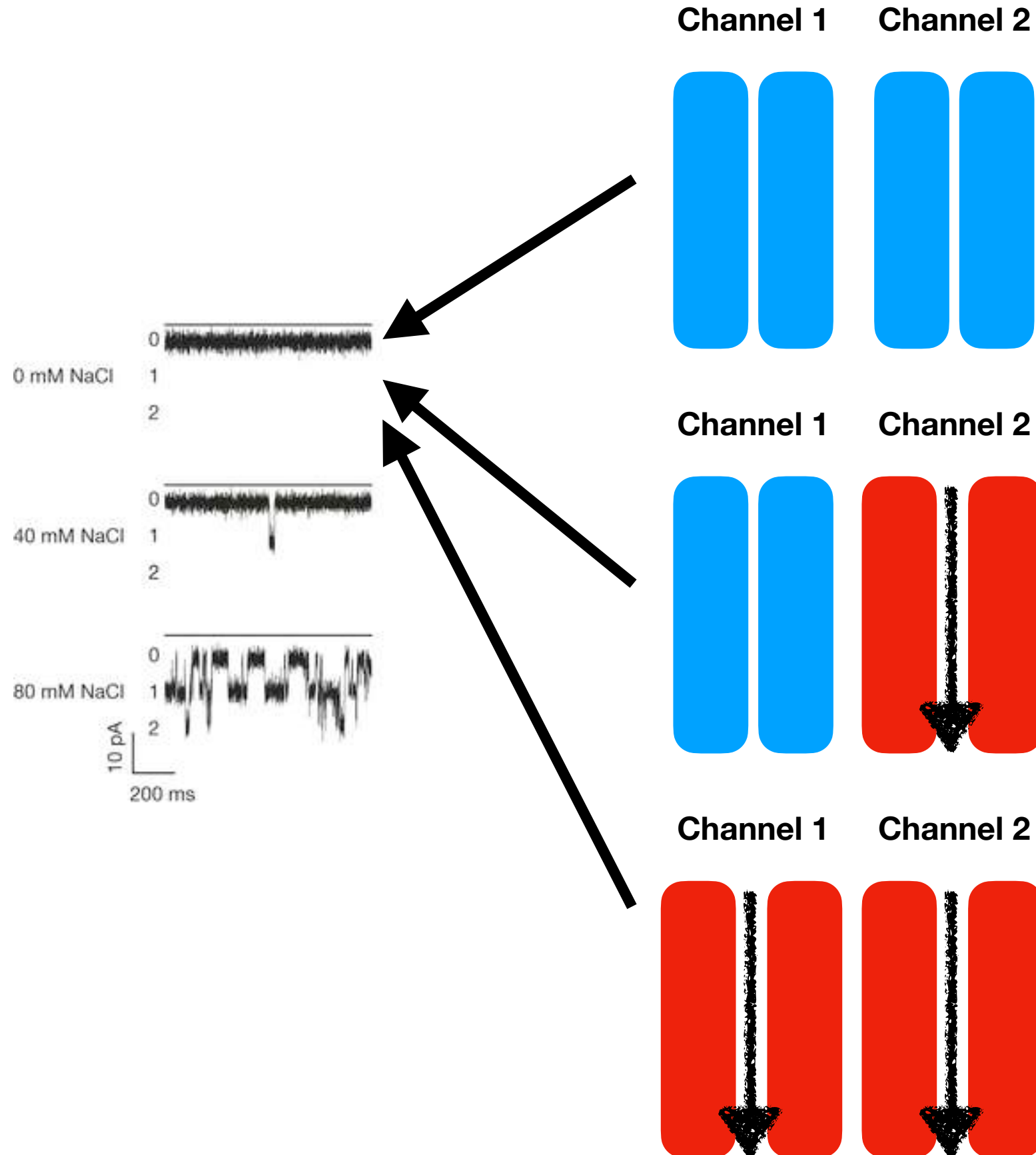
Benefits of heterogeneity?

- How can you use heterogeneity to better understand the biology of your samples?
- Does your heterogeneity correlate with functional changes?
- Always test to ensure that your representative density map is actually representative of your sample, and not merely some small portion of the particles that generate a high-resolution structure?
 - If the map does result from a very small fraction of particles, try to understand why?
 - Can you test activity to see if that makes sense biologically?

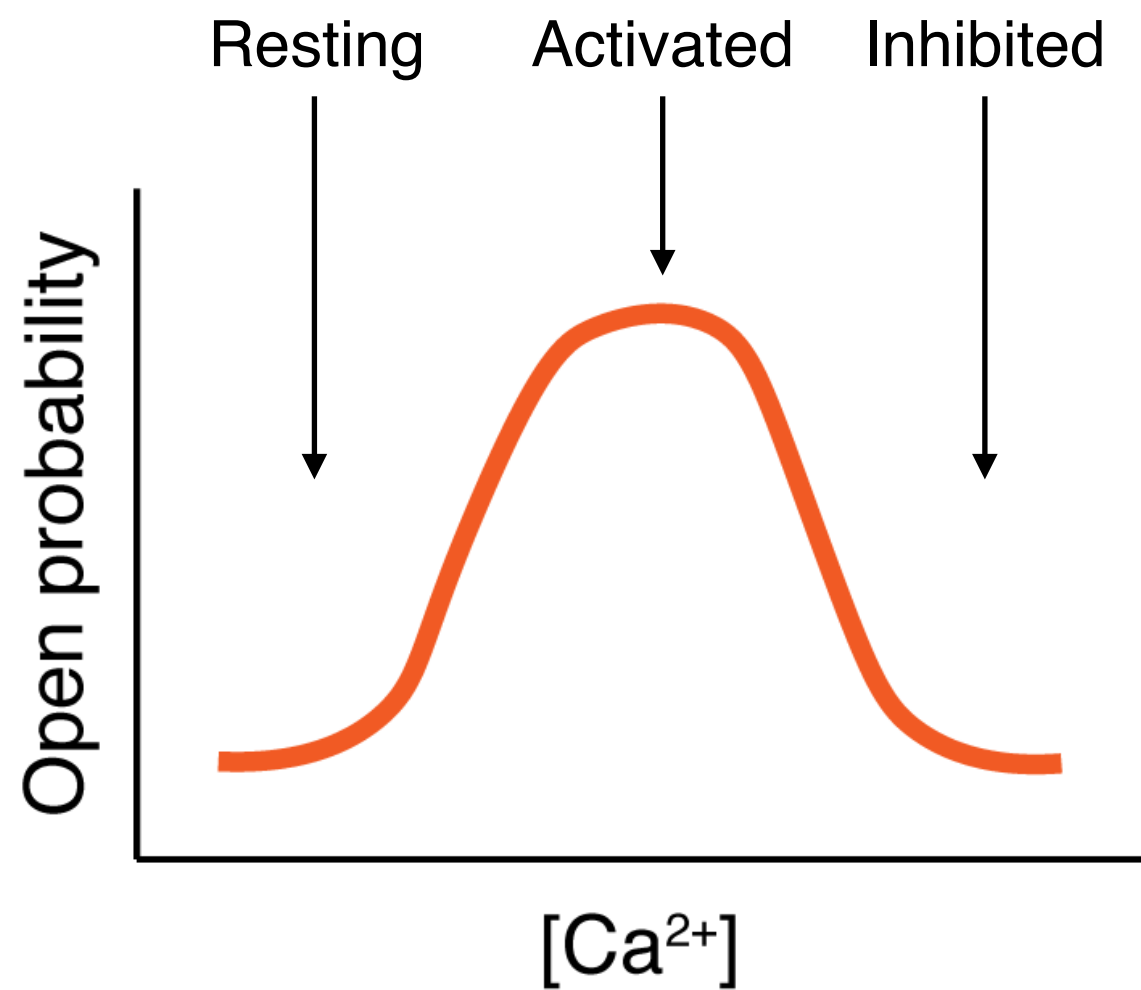
Cytosolic Ca^{2+} signaling



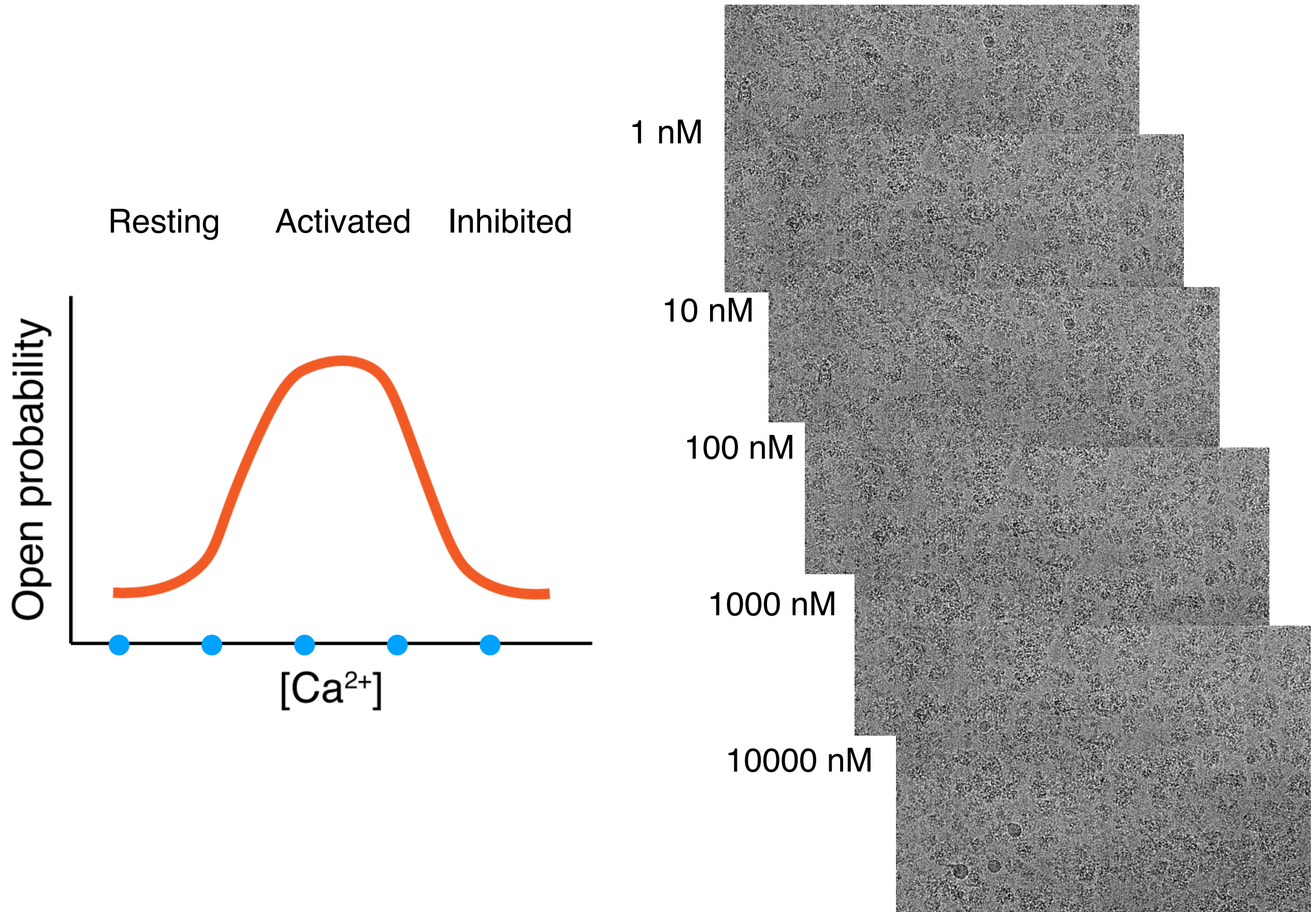
Ion channel gating



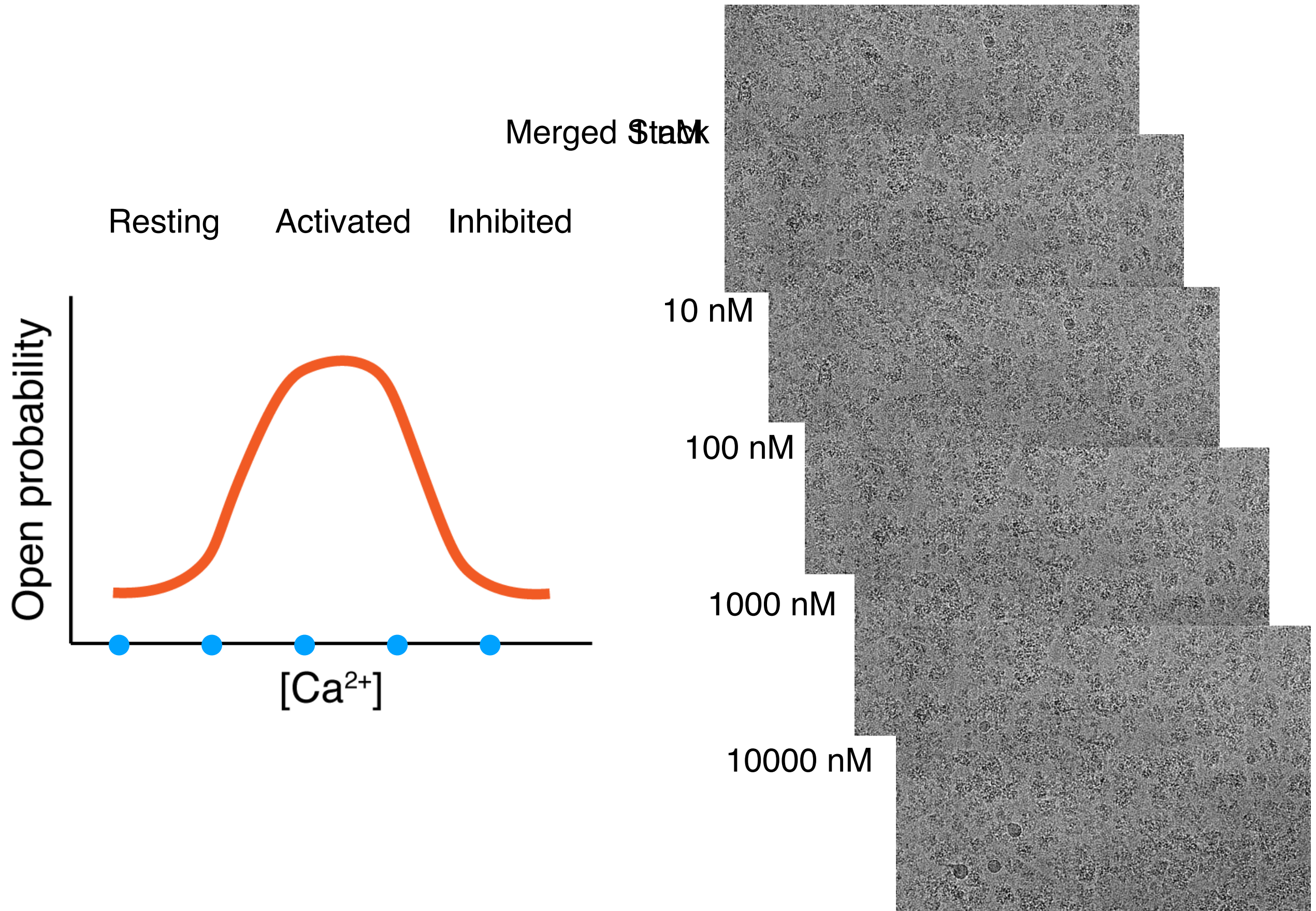
Visualizing ligand-dependent activation and inhibition



Visualizing ligand-dependent activation and inhibition

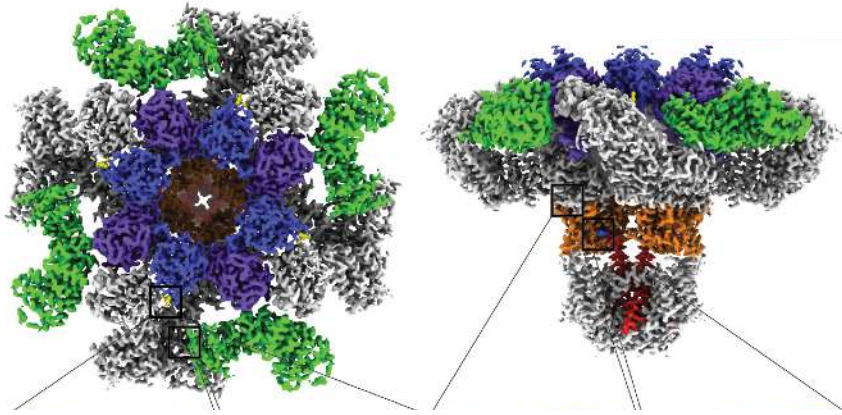


Visualizing ligand-dependent activation and inhibition

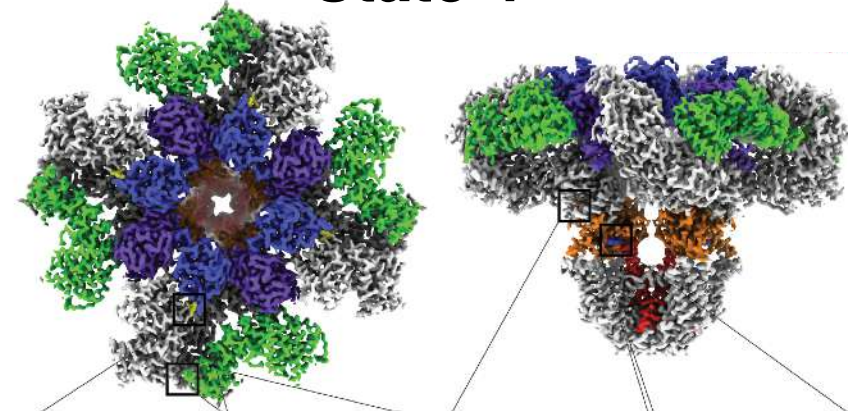


Interpreting the classes from the titration

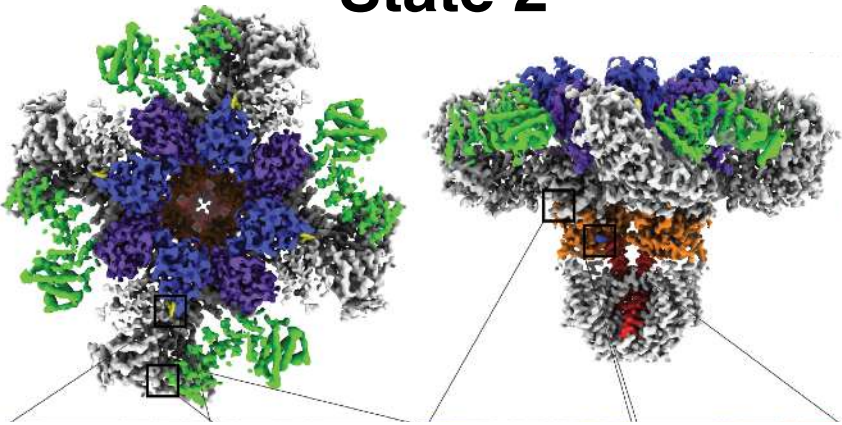
State 1



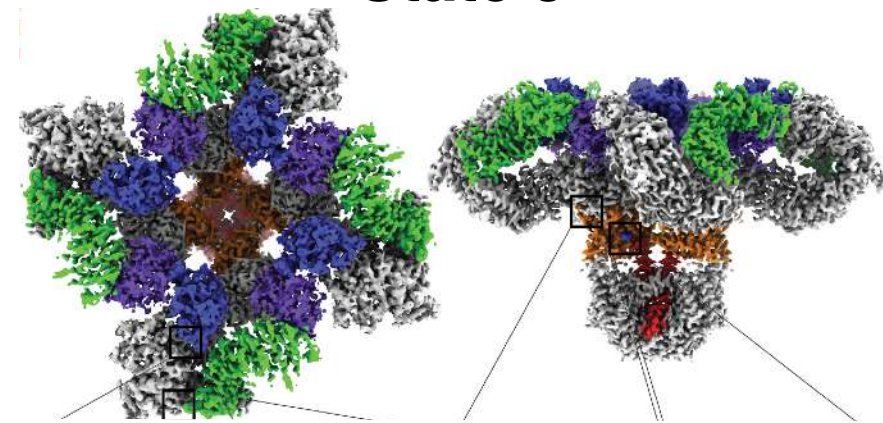
State 4



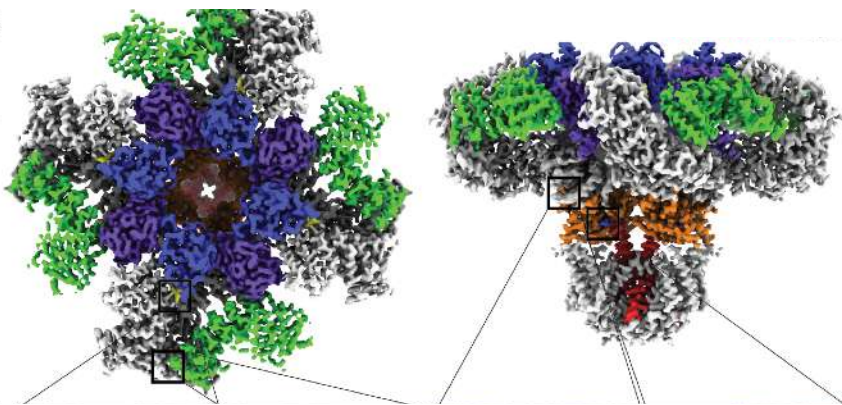
State 2



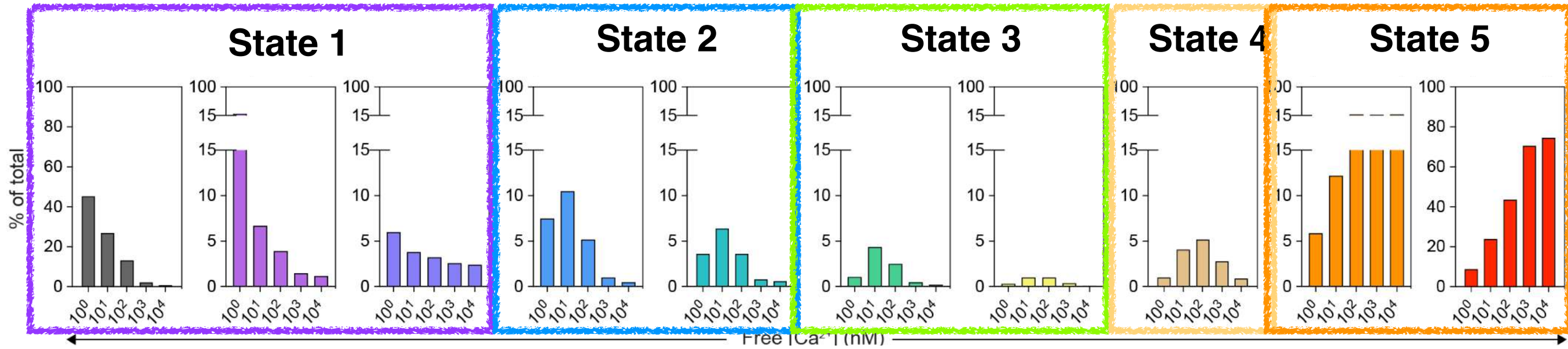
State 5



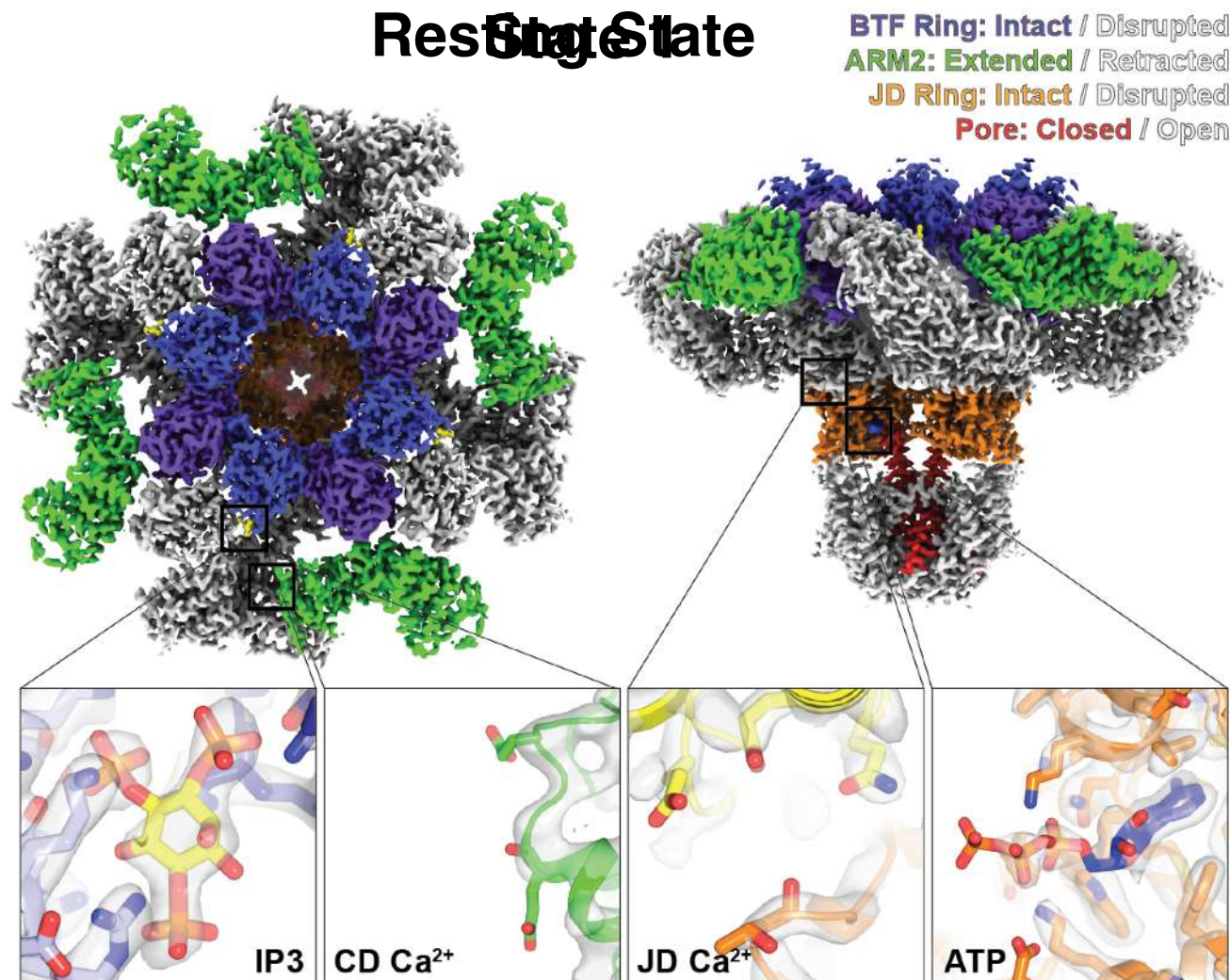
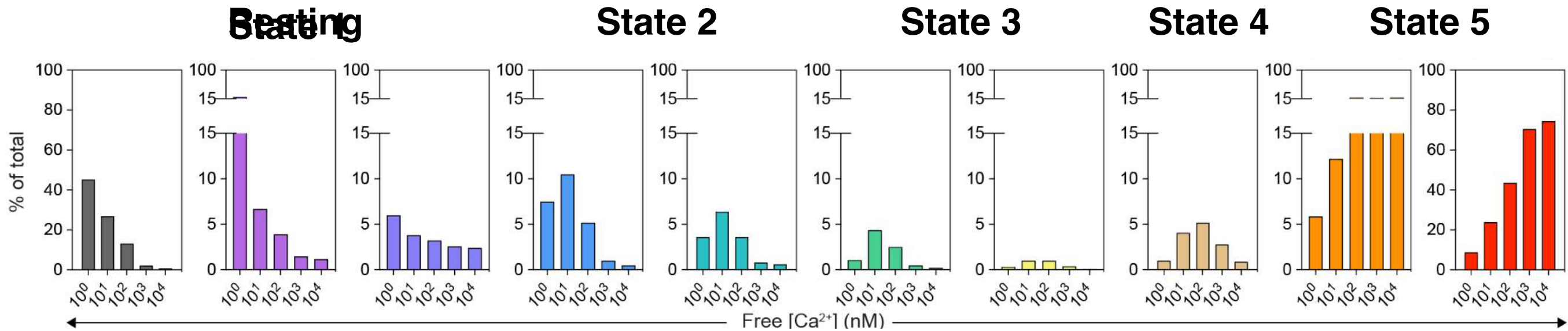
State 3



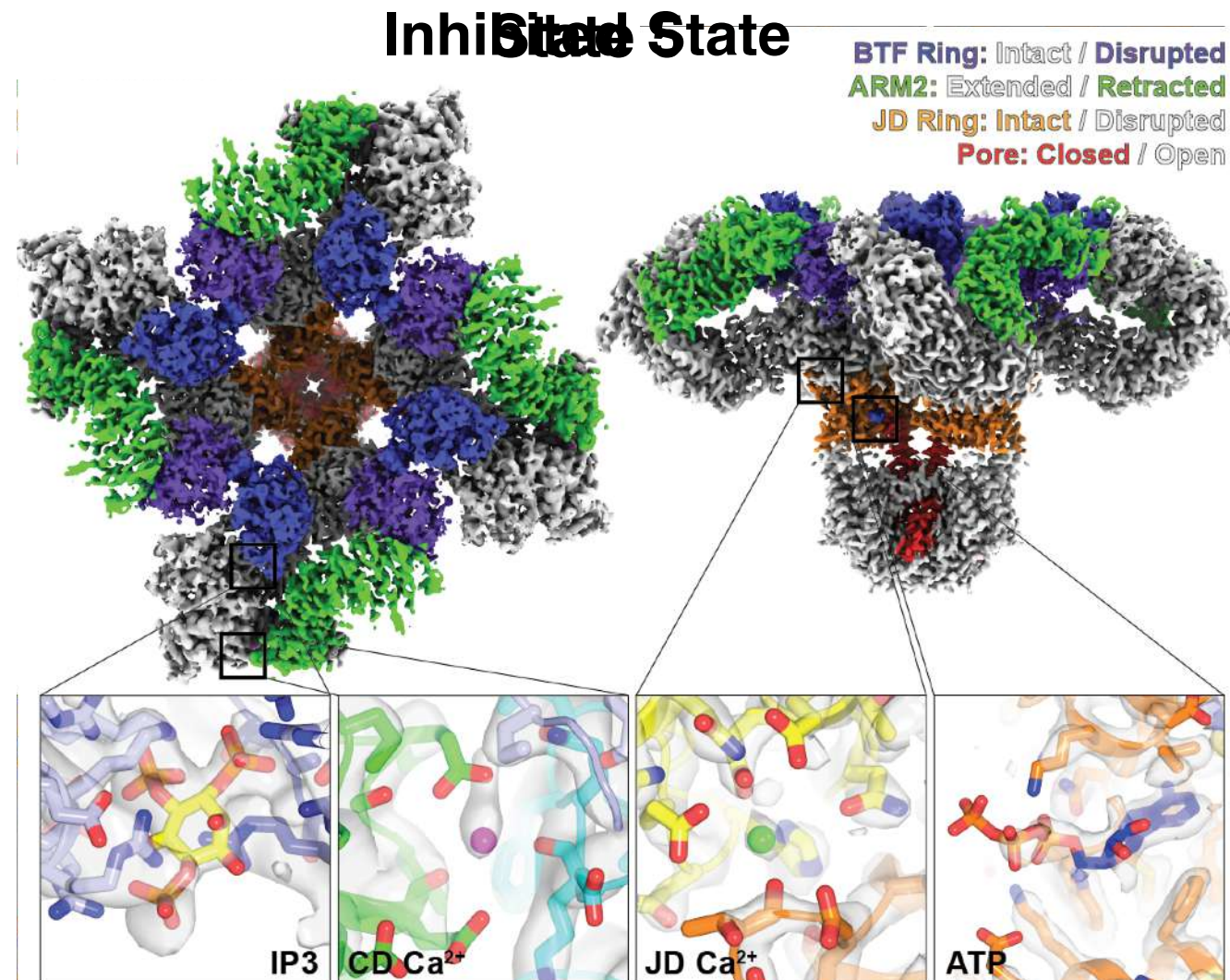
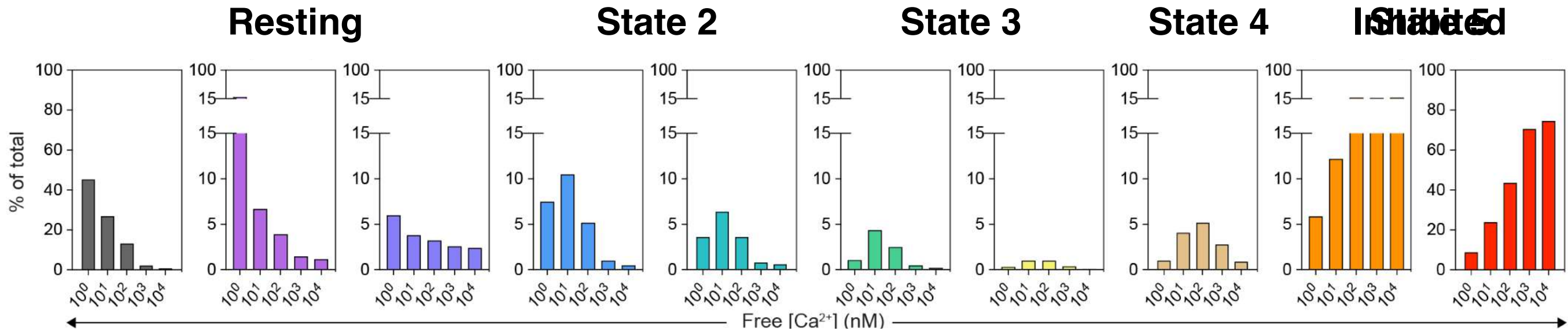
Ca²⁺ dependence of conformational state



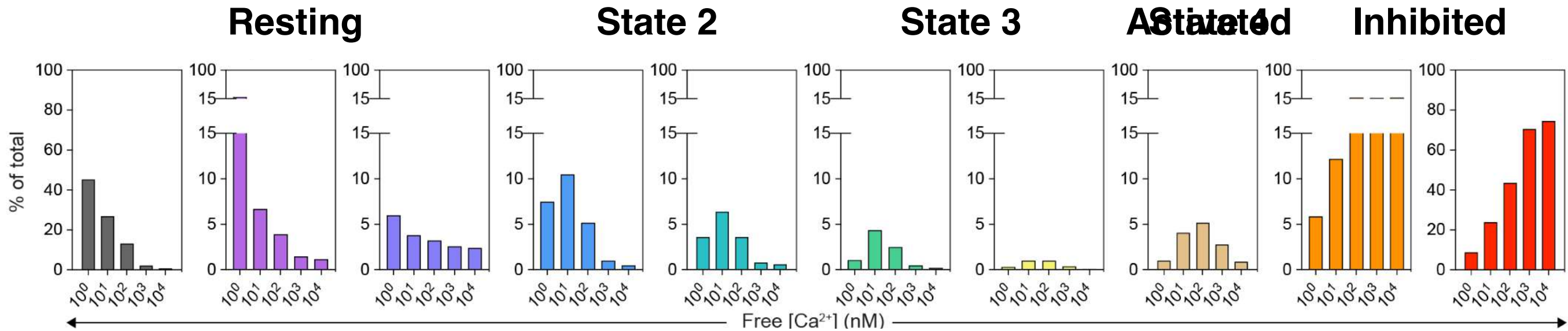
Ca²⁺ dependence of conformational state



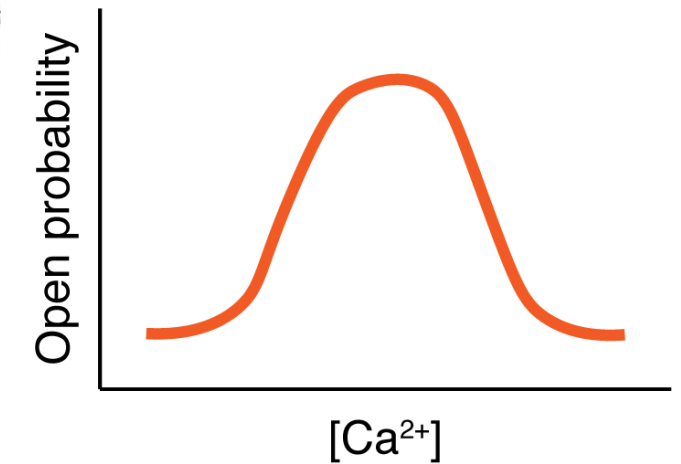
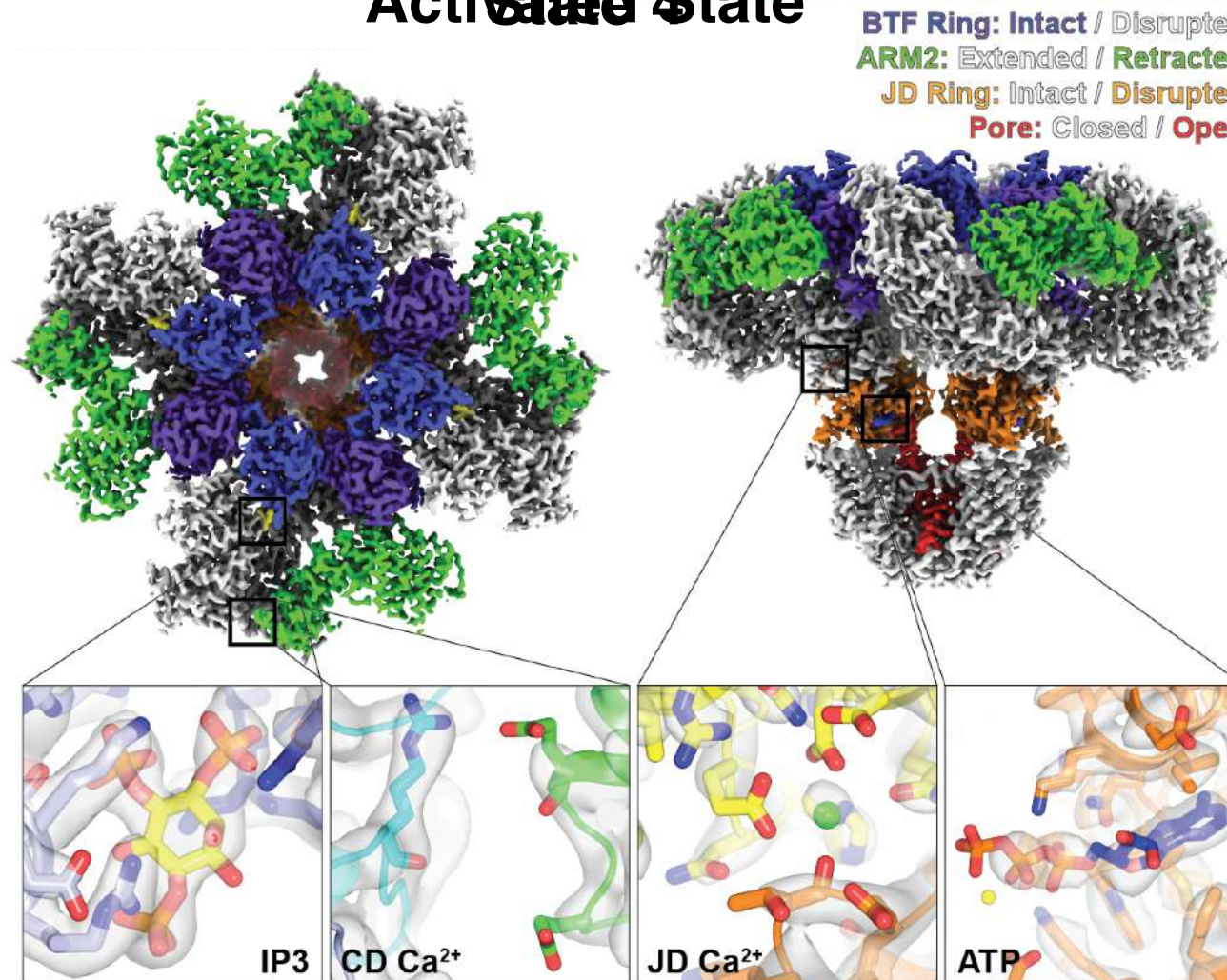
Ca²⁺ dependence of conformational state



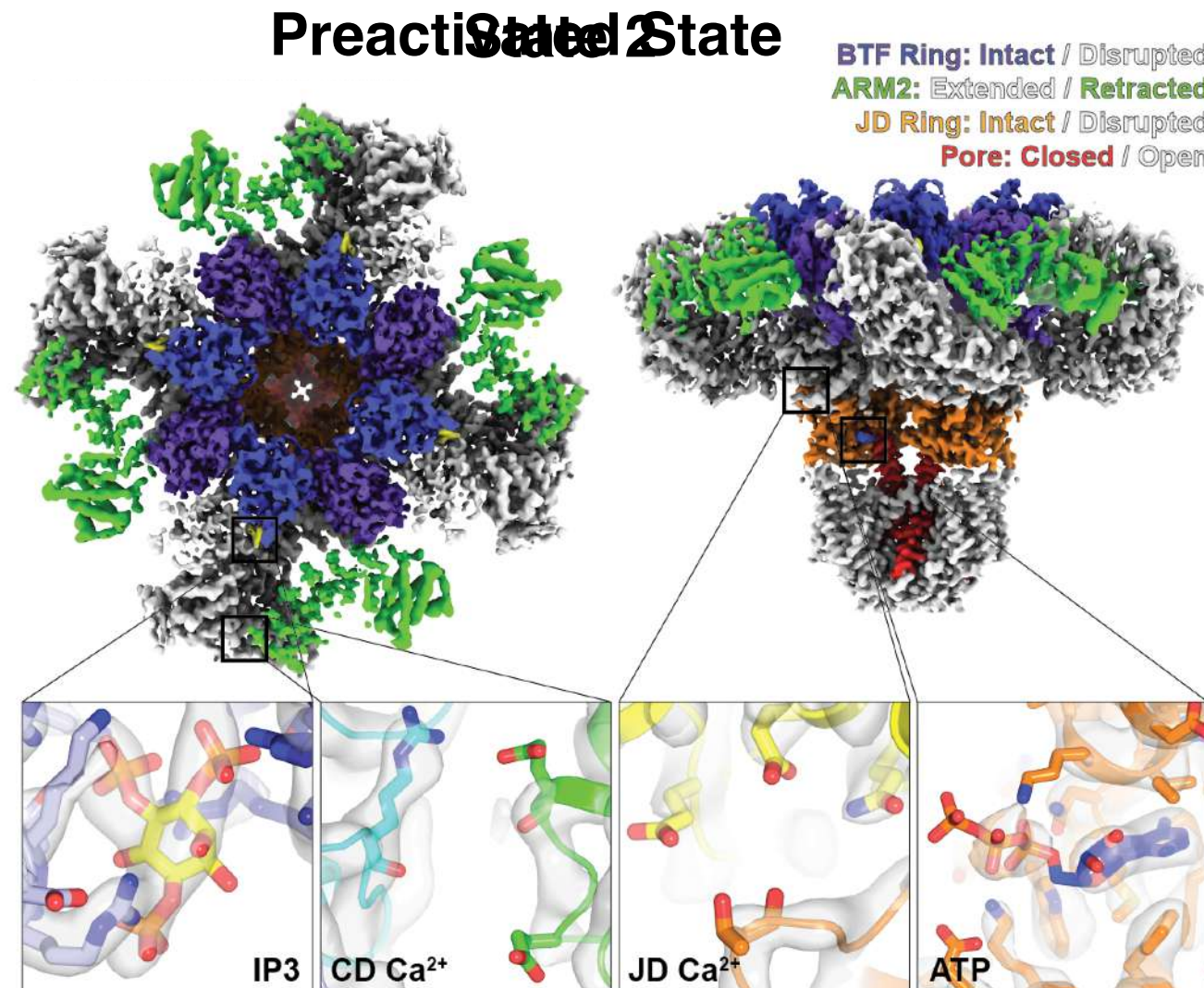
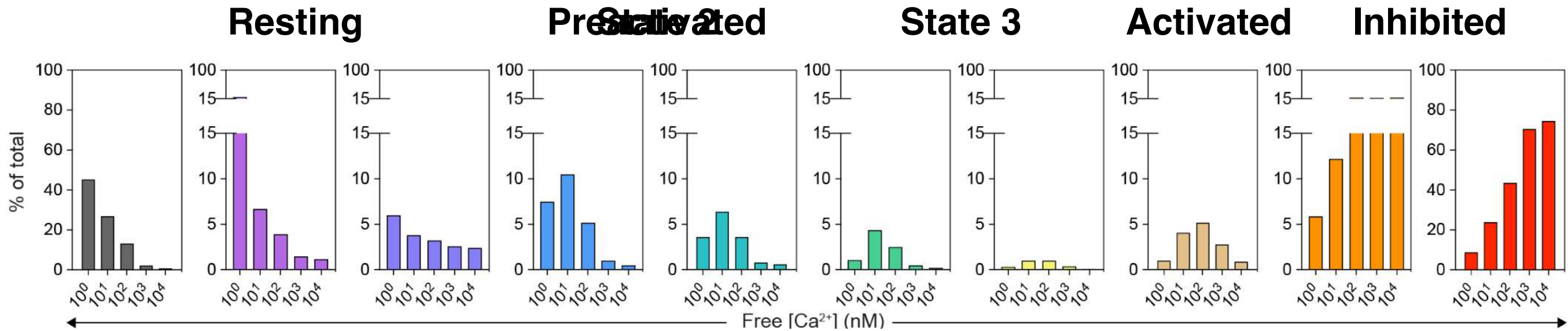
Ca²⁺ dependence of conformational state



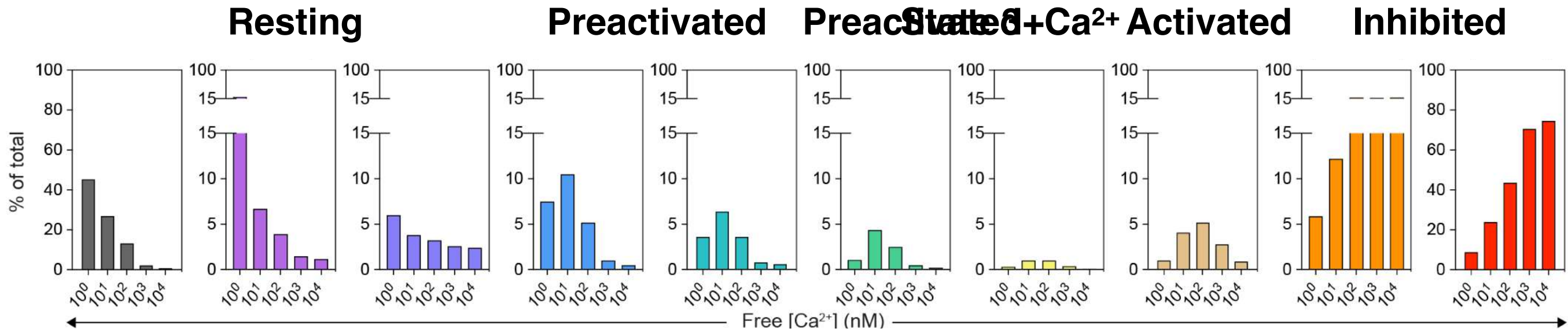
Active State



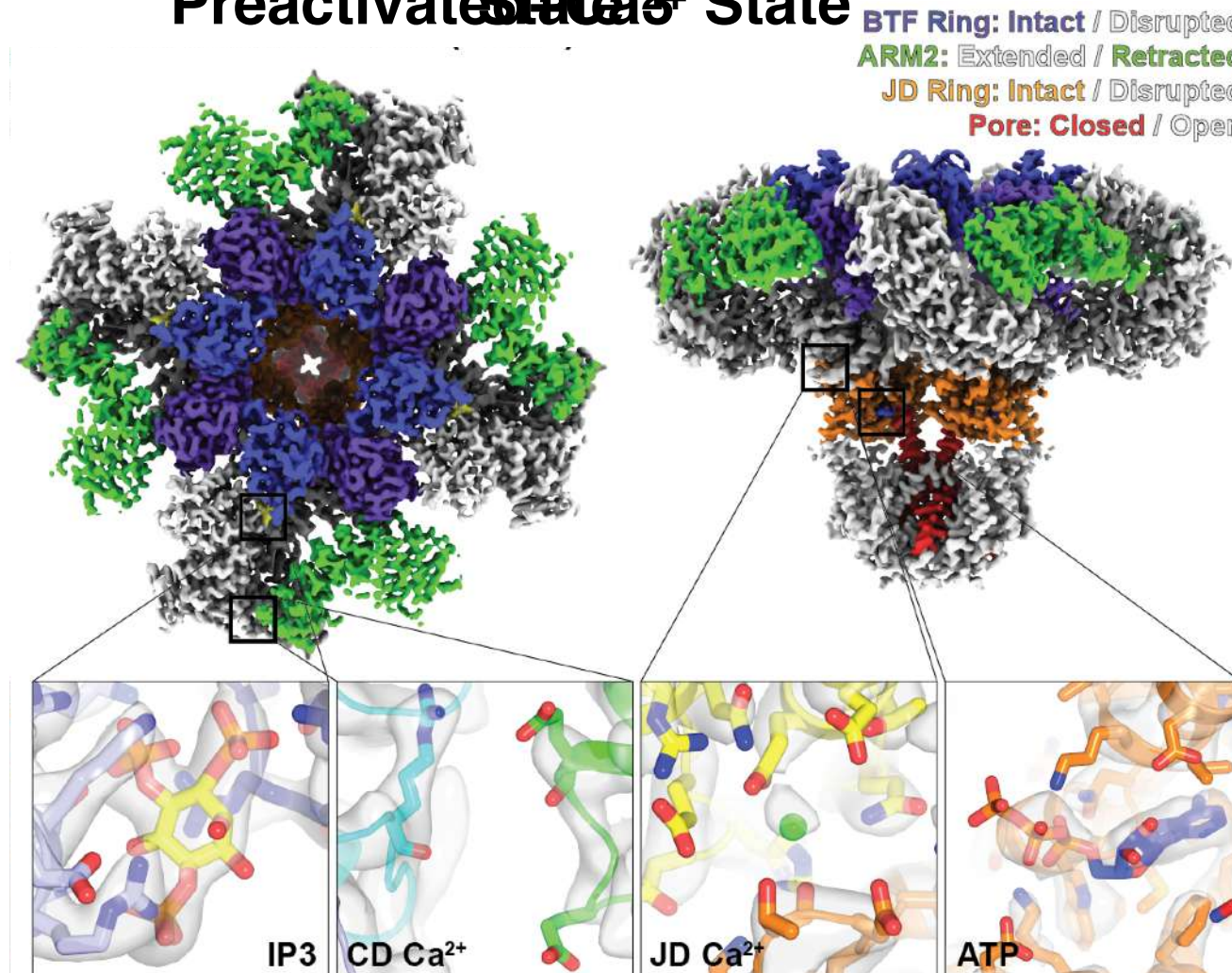
Ca²⁺ dependence of conformational state



Ca²⁺ dependence of conformational state

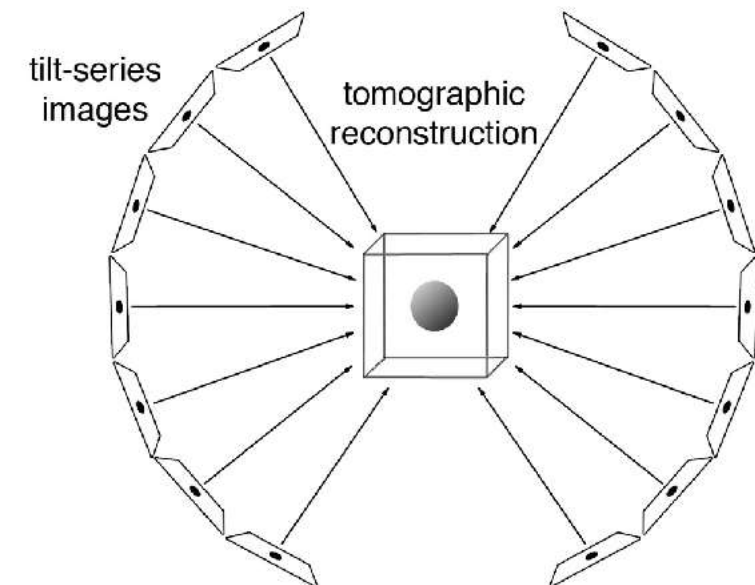
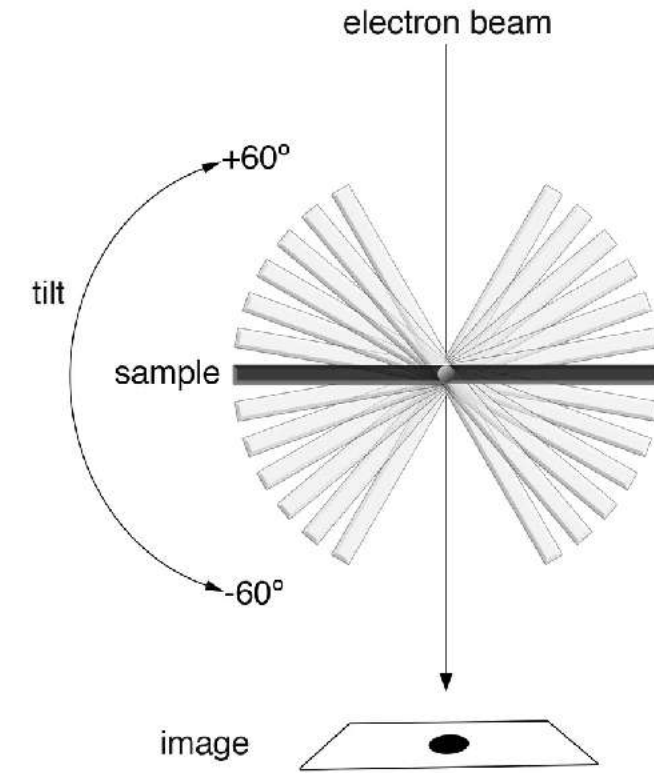


Preactivated State

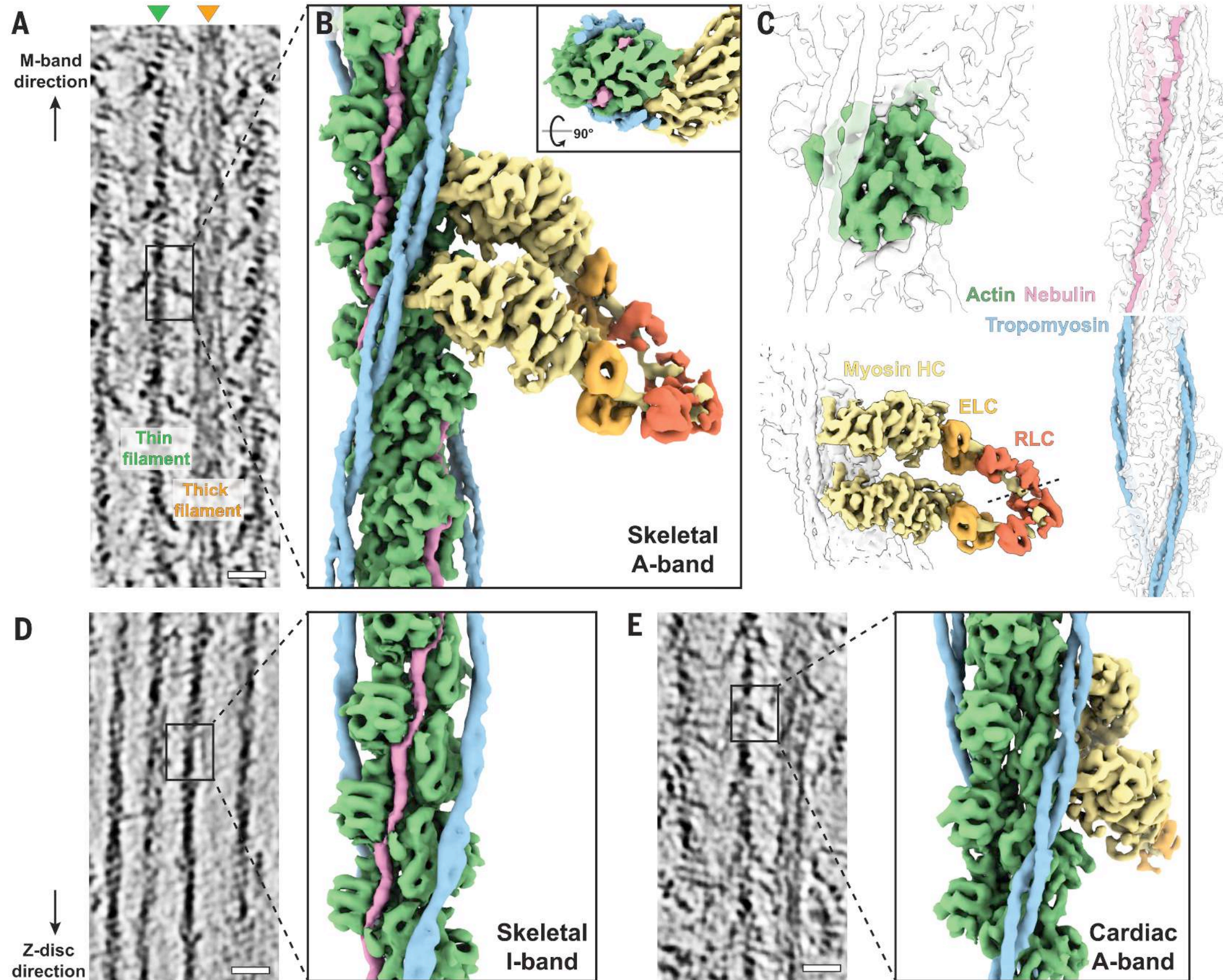


In cellulo heterogeneity?

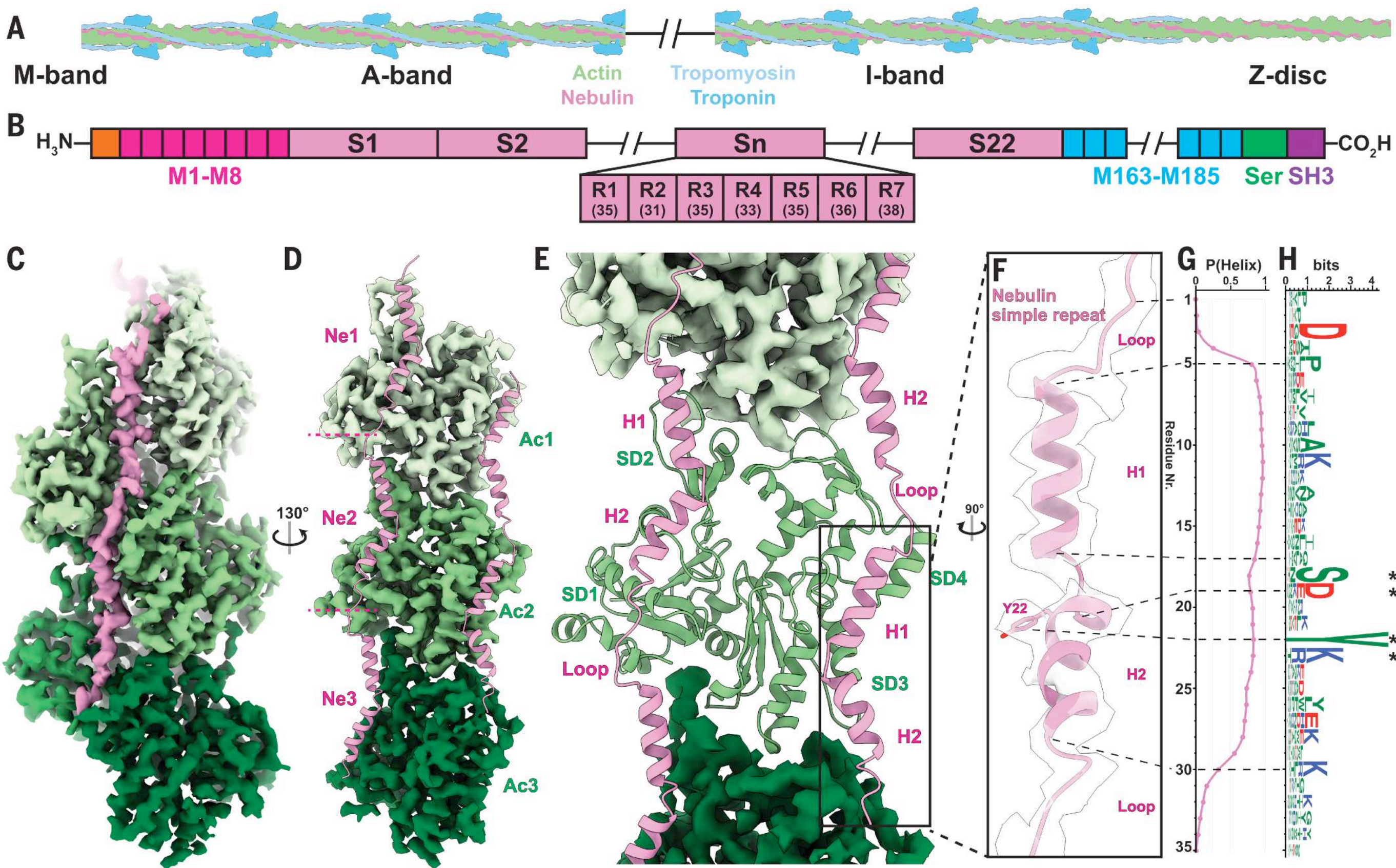
- Cryo-electron tomography
- Many images of the same specimen
- Images are collected at different tilt angles to allow different views of the sample
- Images can then be reconstructed into volumes
- Samples can be purified, crude or recorded from samples in vitrified cells



Sub-tomogram averaging

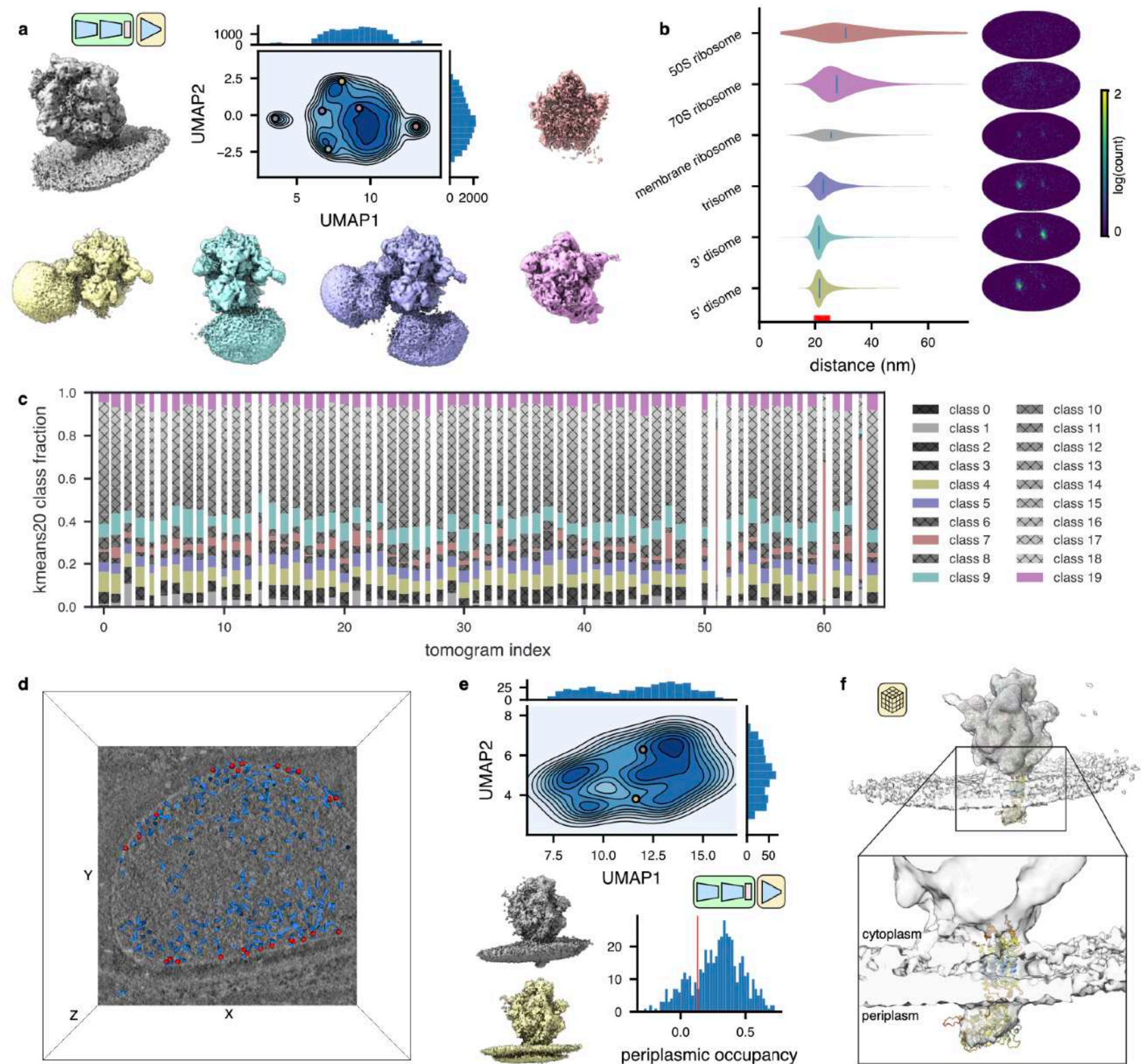


Sub-tomogram averaging



Heterogeneity in sub-tomogram averages

- TomoDRGN is an application of cryoDRGN to tomographic data, allowing one to visualize complex landscapes of complexes in cells



Resources

- Cryo-EM databases
 - Electron microscopy data bank - all published EM structures are deposited and released upon publication
 - Protein data bank - all atomic structures of proteins (crystallography, NMR and EM) are deposited and released upon publication
- Cryo-EM online courses
 - CalTech Getting Started in Cryo-EM <http://cryo-em-course.caltech.edu/videos>
 - LMB Cryo-EM course 2017 <https://www2.mrc-lmb.cam.ac.uk/research/scientific-training/electron-microscopy/>
- NYC resources
 - NYSBC Simons Electron Microscopy winter-spring course - offered annually with distinguished lecturers from New York City and beyond <http://semc.nysbc.org/the-winter-spring-2020-em-course/>