

CRYOEM 001 : EM SAMPLE PREPARATION

NCCAT Embedded Training — Master Class series

September 2020

NATIONAL CENTER FOR
CRYOEM ACCESS & TRAINING



New York Structural
Biology Center

SIMONS ELECTRON
MICROSCOPY CENTER



CRYOEM 001 : SINGLE PARTICLE MASTERCLASS

Introduction to cryoEM: SPA

Building a cryoEM toolkit

EM compatible samples

EM support films and grids

Sample preparation

Tools of the trade:

microscopes and detectors

Microscope operations

Data collection strategies

Data assessment & QC

Data processing:

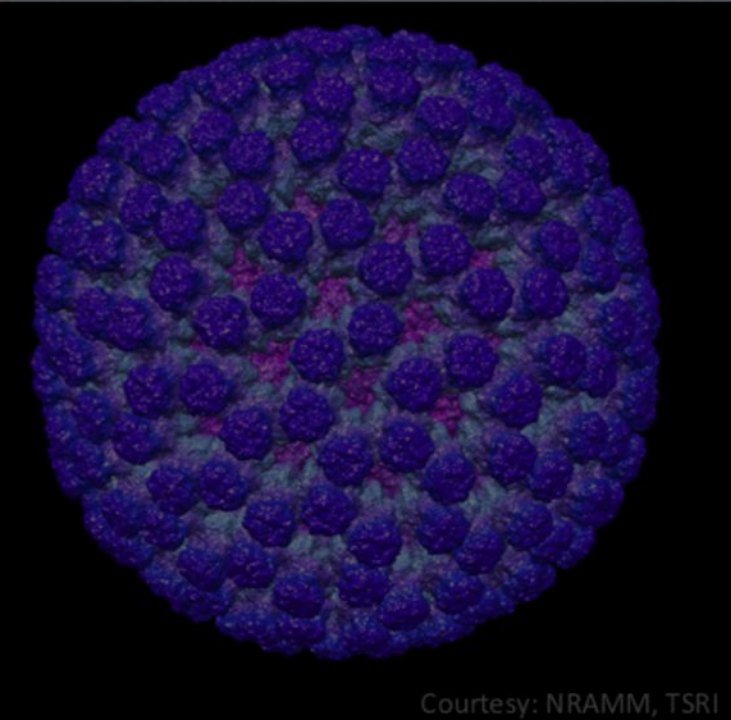
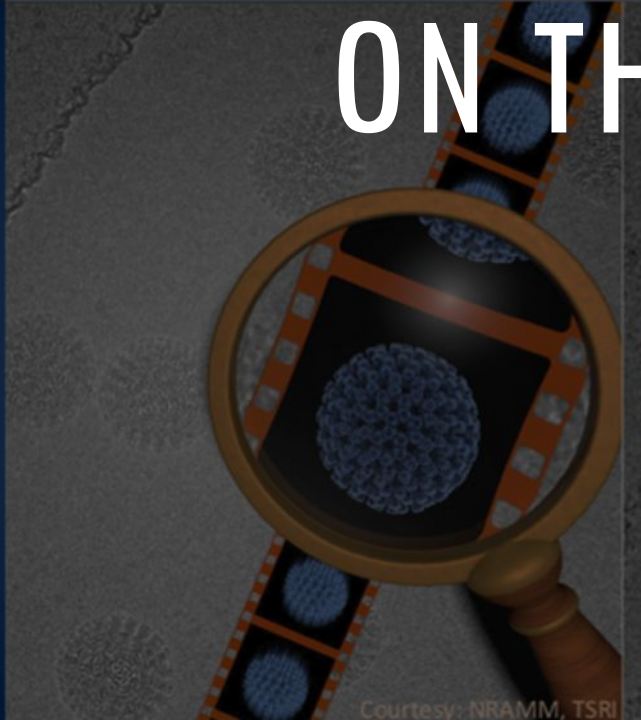
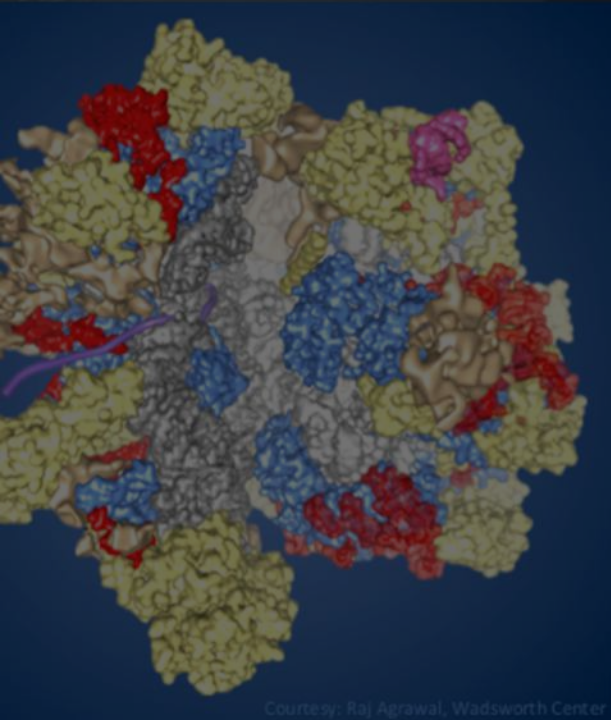
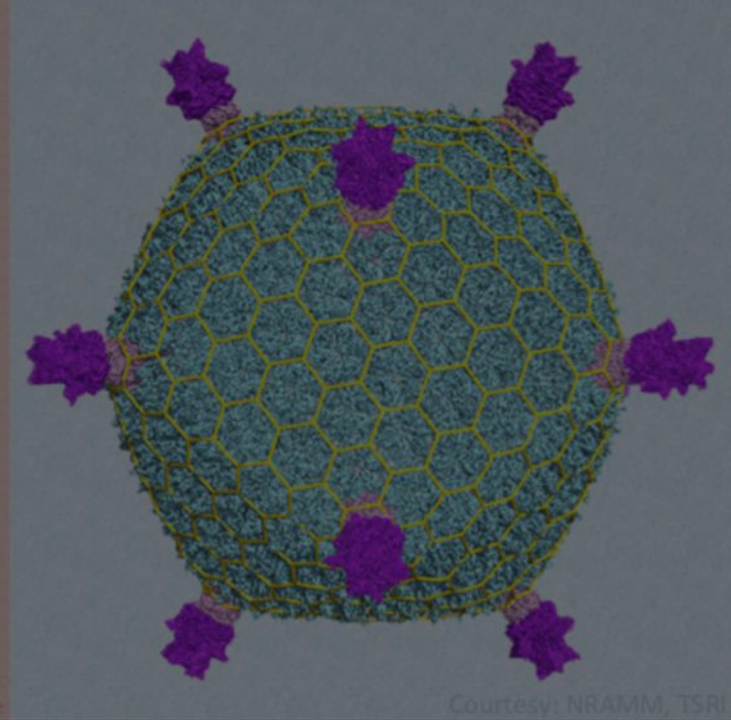
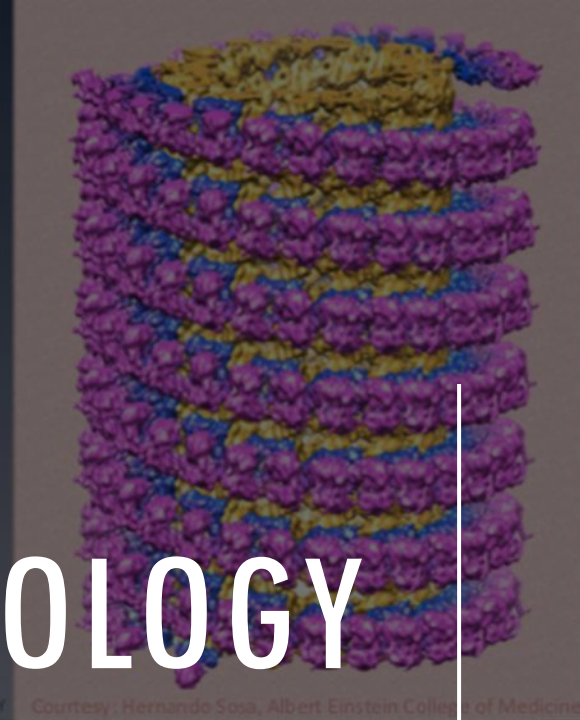
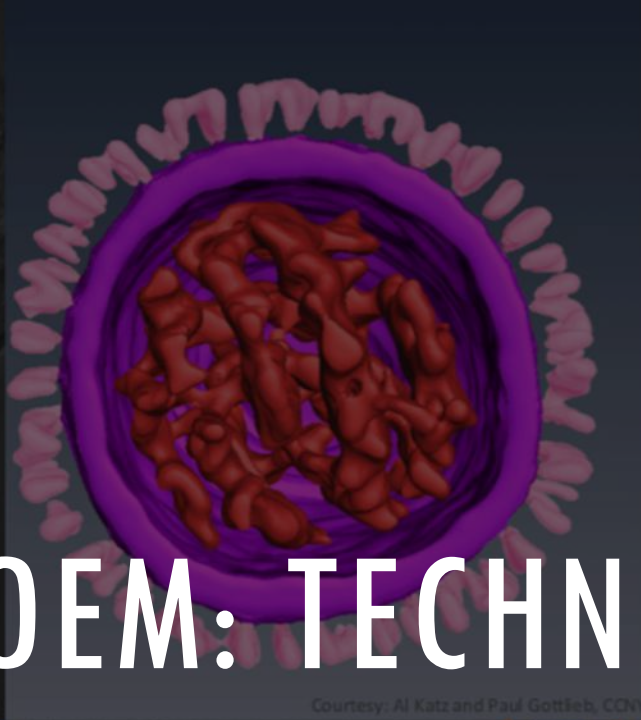
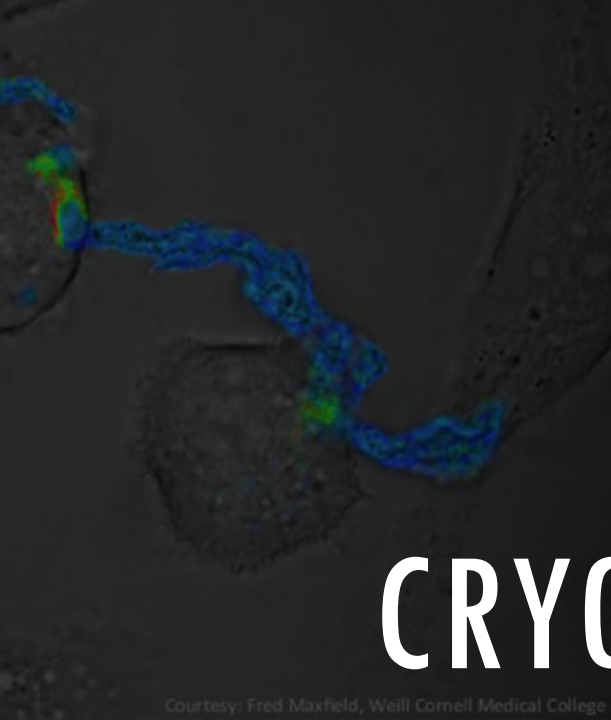
cryoEM IT infrastructure

On-the-fly feedback

3D Reconstruction

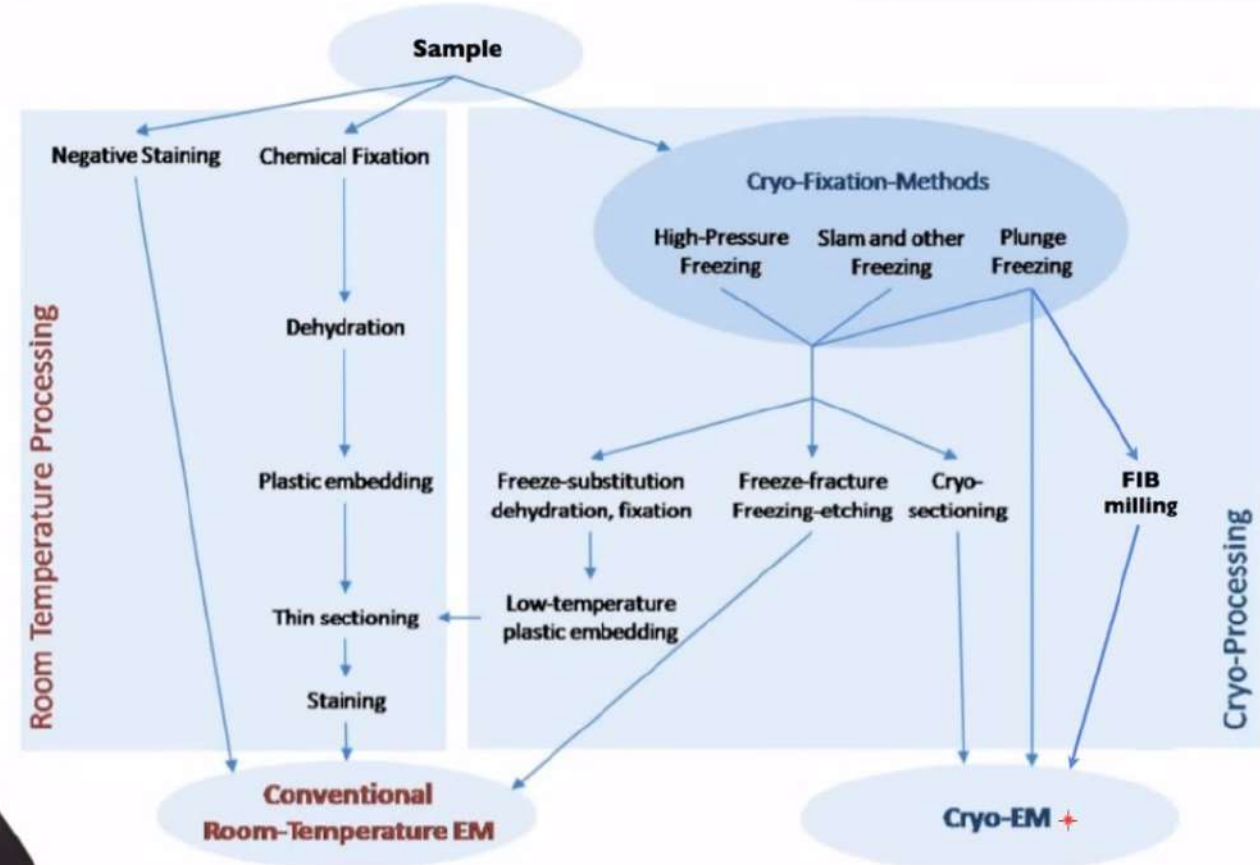
Visualization and validation

CRYOEM: TECHNOLOGY ON THE RISE



RT & CRYO SAMPLE PREP METHODS

adapted from
Pilhofer et al.,
MCB 2010



HOW ARE SAMPLES PREPARED?

Sample

proteins/
macromolecular
complexes

Biochemically
homogeneous

Biochemically
heterogeneous

cells/tissues/
organisms

Structurally
homogeneous

Structurally
heterogeneous

Sample preparation

2D/3D
crystallization

Helical
assembly

Single particle
isolation

Serial sectioning

Cryo embedding

EM technique

Electron
crystallography

Helical
reconstruction

Single particle
analysis

Electron
tomography

FIB-SEM/cryoET

Resolution range

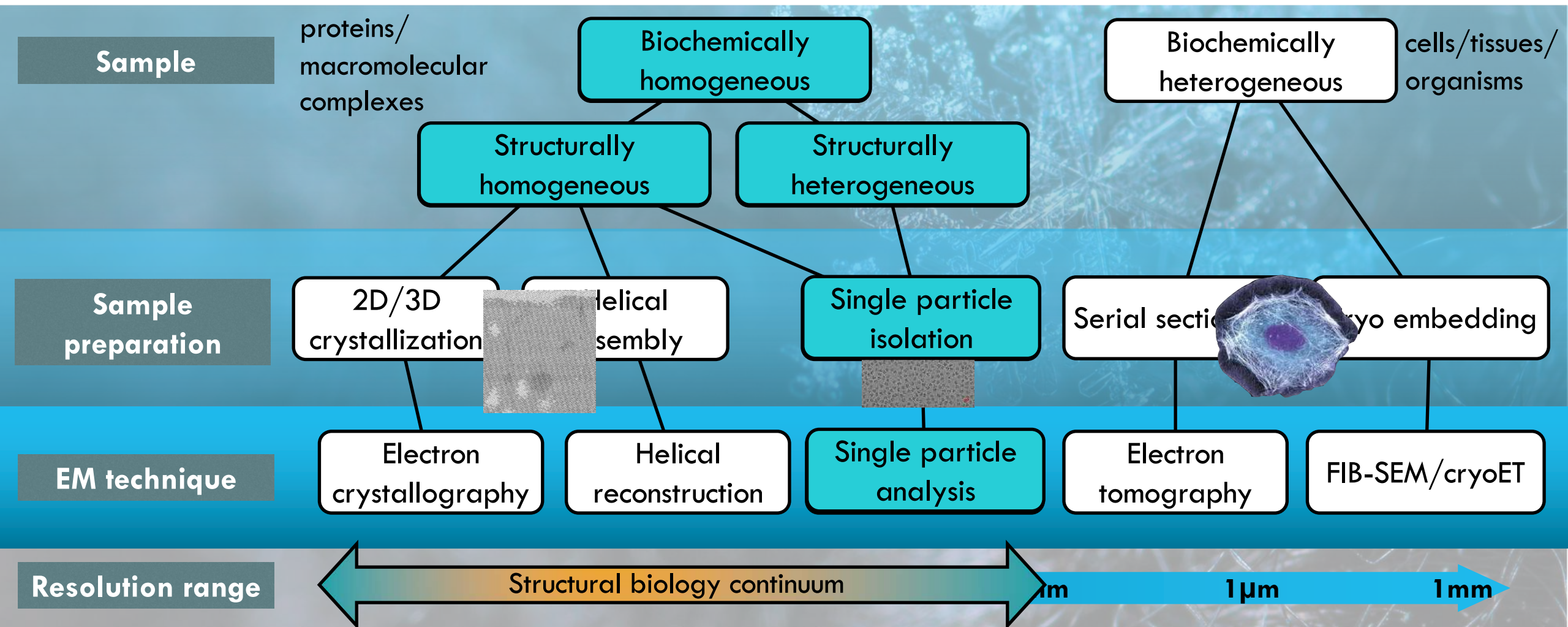
1Å

1nm

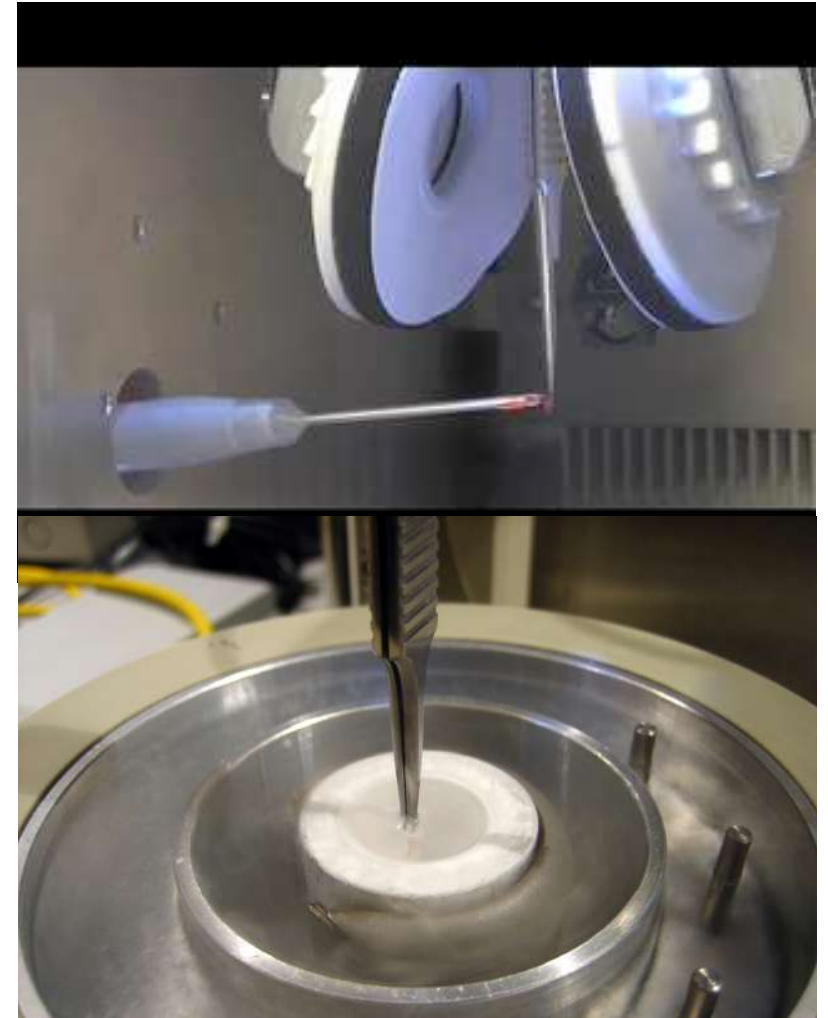
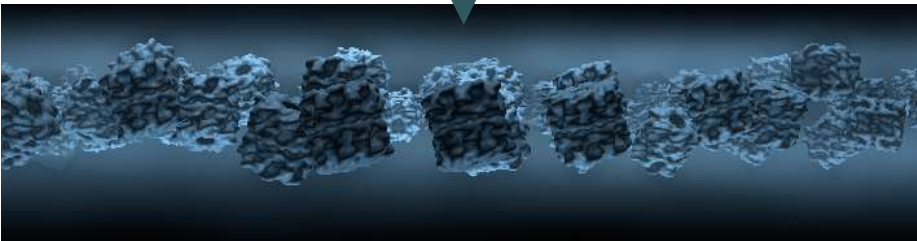
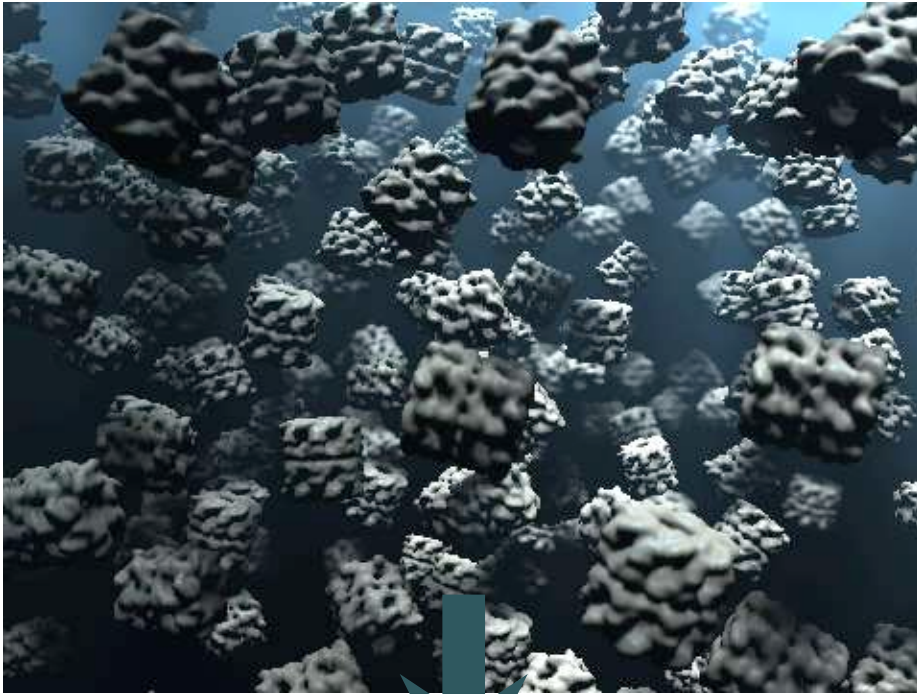
1µm

1mm

HOW ARE SAMPLES PREPARED?

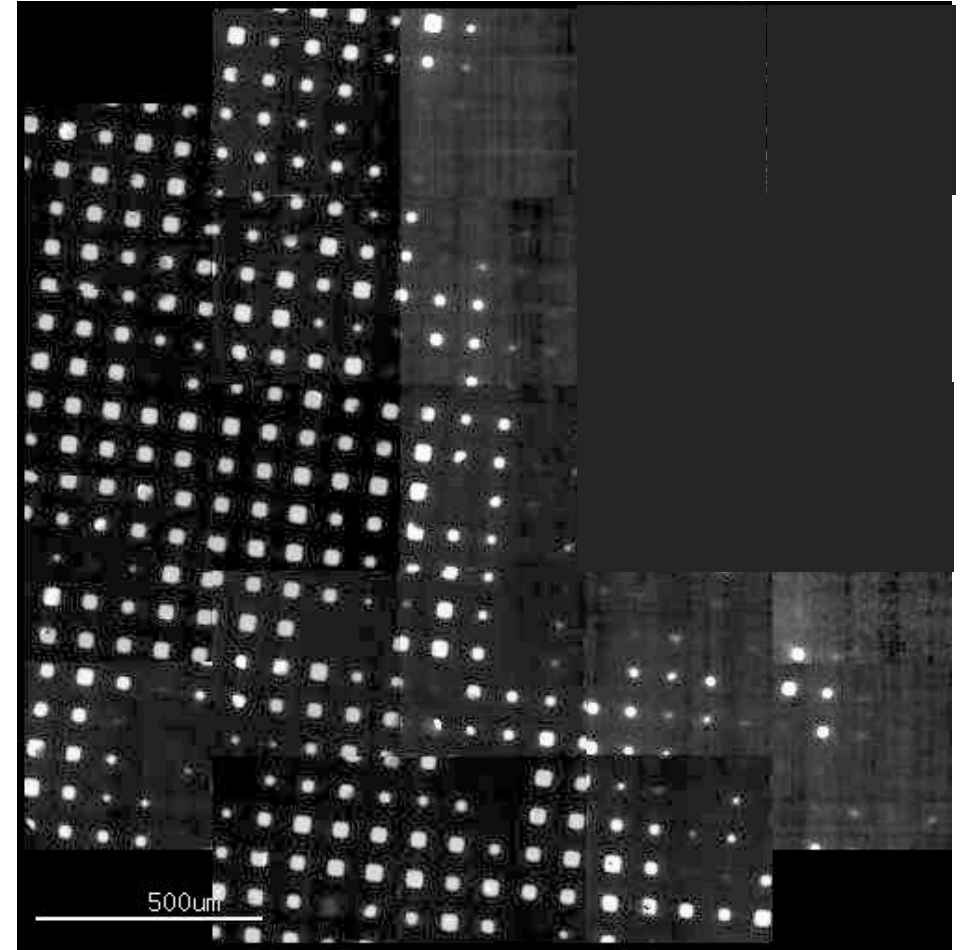
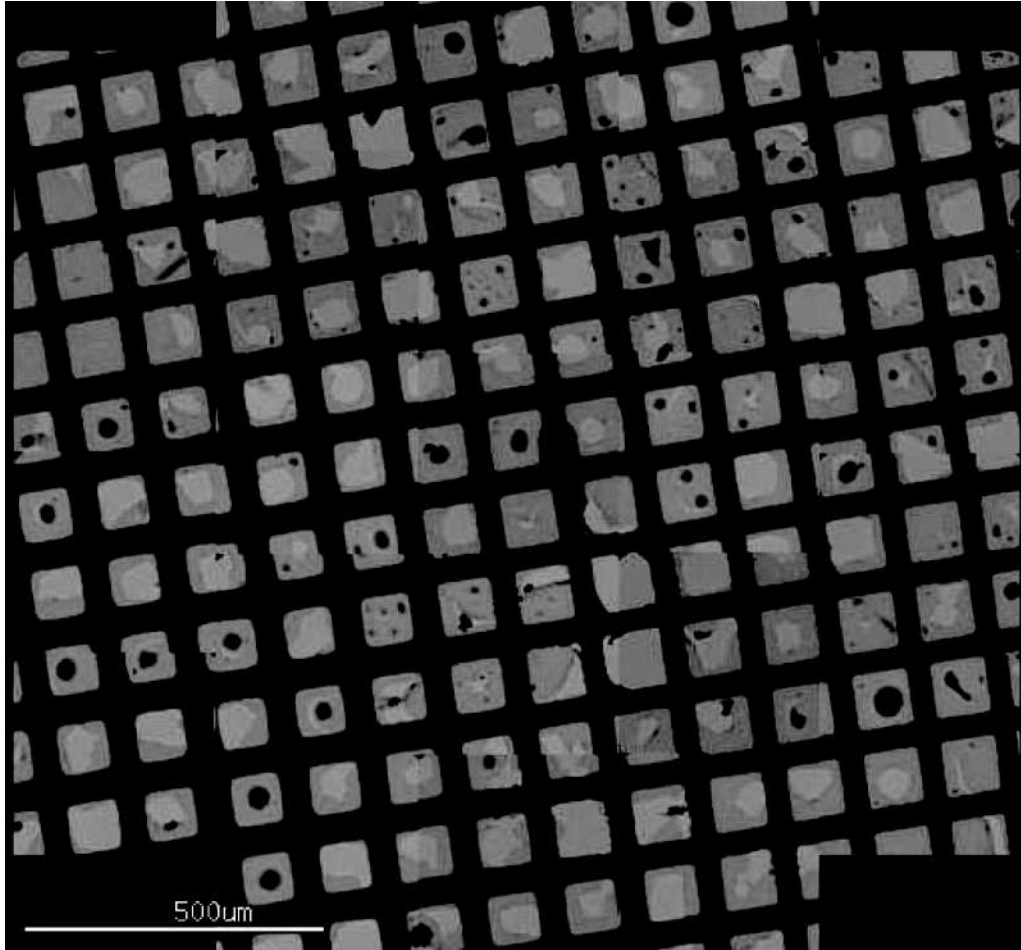


PLUNGE FREEZING

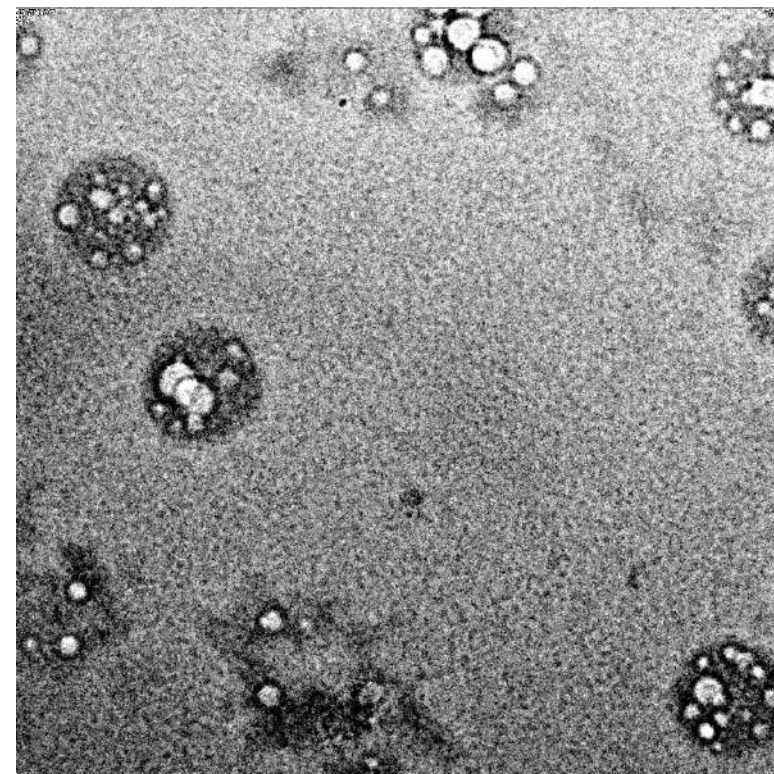
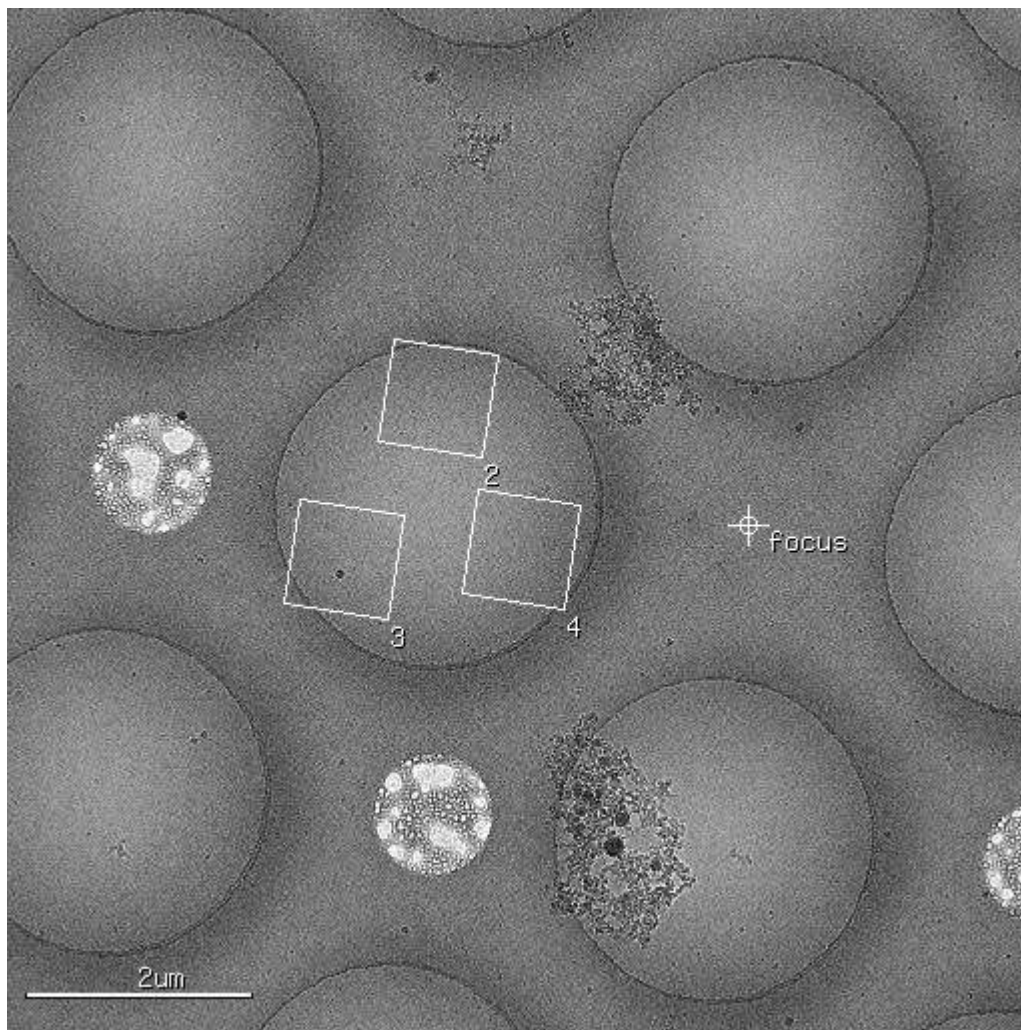




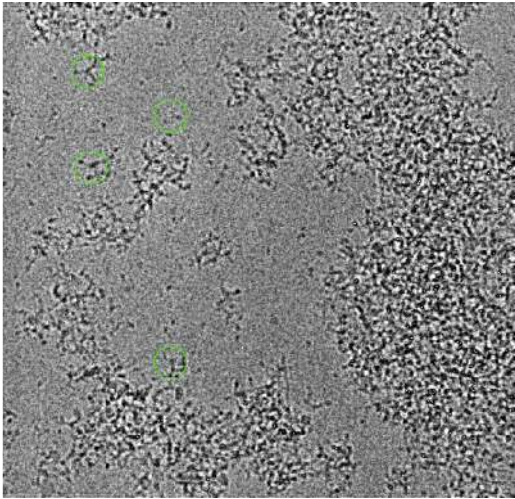
WHAT DO GRIDS LOOK LIKE?



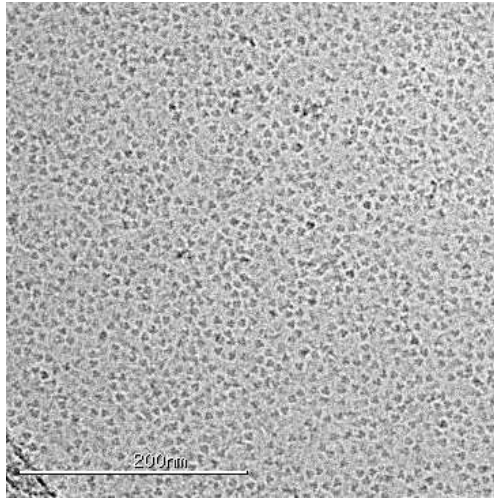
LOW DOSE IMAGING



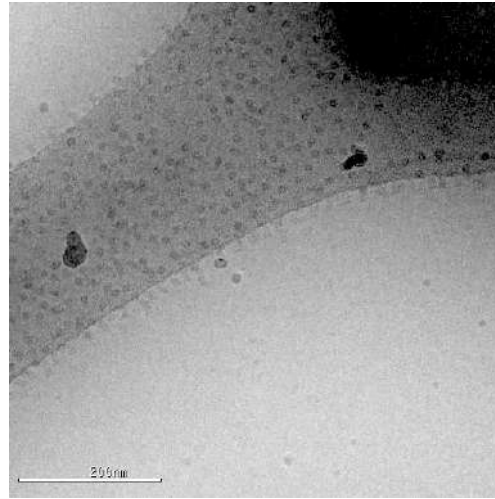
WHAT ISSUES ARISE?



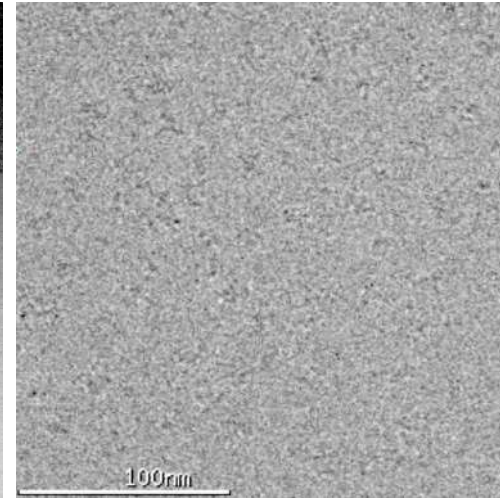
Aggregating in ice



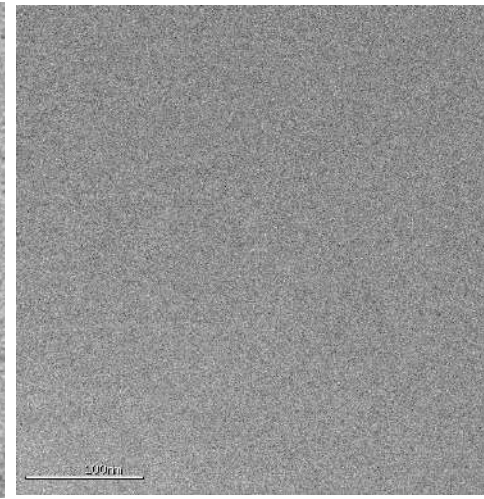
Preferred orientation



Particles not going into holes



Rejecting 90% of particles

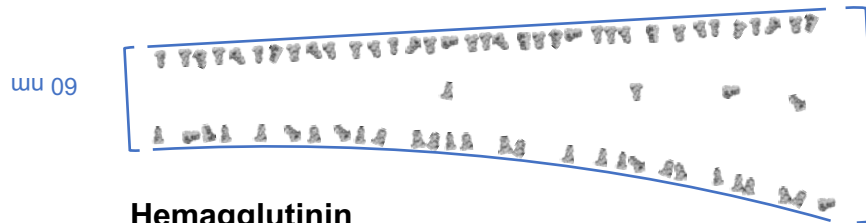


Particles disappearing in ice

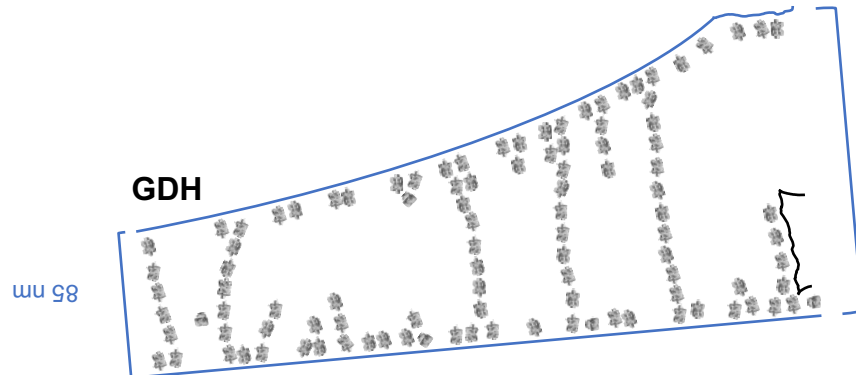
WHAT ISSUES ARISE?



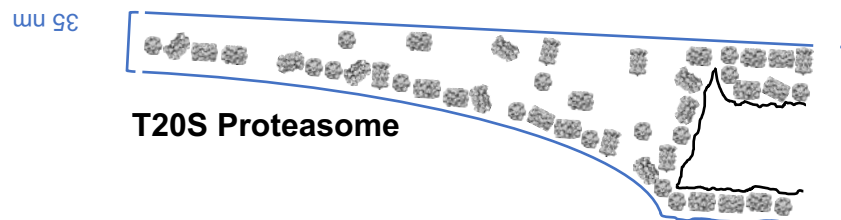
Hemagglutinin



Hemagglutinin

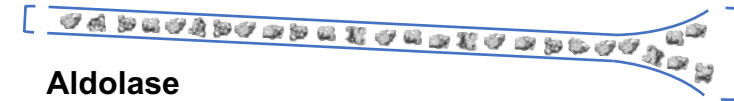


GDH



T20S Proteasome

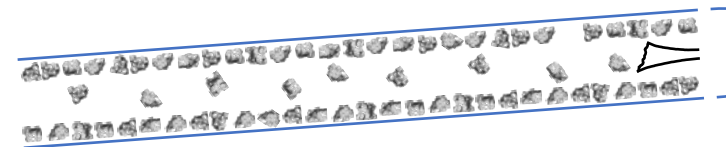
110 nm
ice



Aldolase

45 nm
ice

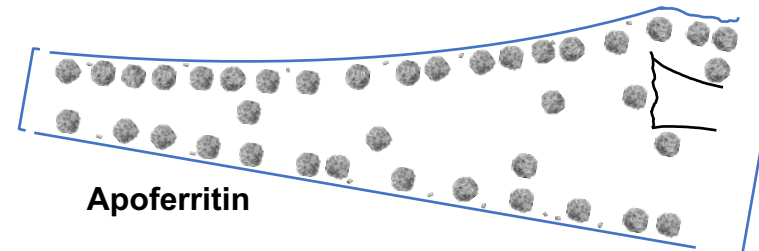
125 nm
ice



Aldolase

50 nm
ice

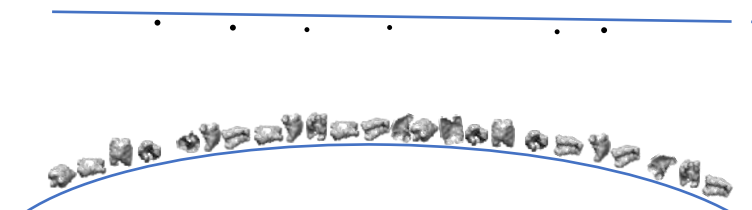
180 nm
ice



Apoferritin

135 nm
ice

115 nm
ice



DNAB Helices

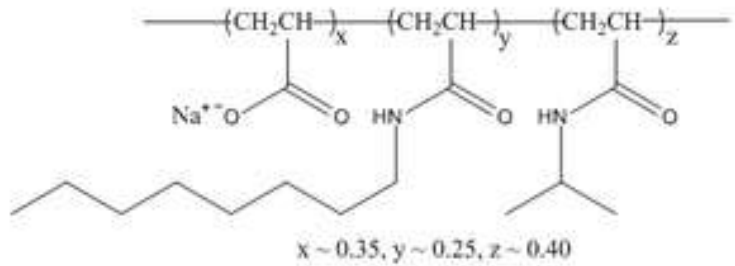
110 nm
ice

Noble AJ, et al.
Routine single
particle CryoEM
sample and grid
characterization
by tomography.
Elife. 2018;7.



Alex Noble

REAGENTS FOR IMPROVING VITRIFICATION OF CRYO-EM GRIDS USED IN SINGLE PARTICLE ANALYSIS.



Molecular Formula:
(C_{6.2}H_{10.3}O_{1.35}N_{0.65}Na_{0.35})₃₅

Molecular Weight:
approx. 8 kDa

CAS#: 1423685-21-5

Amphipol A8-35

A short amphipathic polymer that is specifically designed for membrane protein stabilization. The surfactant possesses a very high affinity for the transmembrane surfaces and allows to solubilize membrane proteins in a detergent-free aqueous solution

REAGENTS FOR IMPROVING VITRIFICATION OF CRYO-EM GRIDS USED IN SINGLE PARTICLE ANALYSIS.

Surfactants and Cryoprotectants	Amount	Conc.	CMC	Class
Fluorinated Octyl Maltoside (FOM)	100 µl	0.41% (w/v)	0.07% (w/v)	non-ionic detergent
Hexadecyl-trimethyl-ammonium Bromide (CTAB)	100 µl	0.34% (w/v)	0.03% (w/v)	cationic detergent
n-Decyl-β-D-Maltoside (DM)	100 µl	0.87% (w/v)	0.09% (w/v)	non-ionic detergent
n-Decyl-α-D-Maltoside (DaM)	100 µl	0.46% (w/v)	0.08% (w/v)	non-ionic detergent
n-Dodecyl-β-D-Maltoside (DDM)	100 µl	0.09% (w/v)	0.01% (w/v)	non-ionic detergent
Sodium Deoxycholate	100 µl	1.66% (w/v)	0.17% (w/v)	anionic detergent
Triton X-100	100 µl	0.15% (w/v)	0.01% (w/v)	non-ionic detergent
Tween 20	100 µl	1% (w/v)	0.01% (w/v)	non-ionic detergent
CHAPSO	100 µl	2.5% (w/v)	0.5% (w/v)	zwitterionic detergent
Amphipol A8-35	100 µl	5% (w/v)		anionic surfactant
Glycerol	1 ml	30% (w/v)		cryoprotectant

- [1] Noble *et al.* (2018) Routine Single Particle CryoEM Sample and Grid Characterization by Tomography. DOI: 10.7554/eLife.34257.
- [2] Thonghin *et al.* (2018) Cryo-electron microscopy of membrane proteins. *Methods* **147**:176.
- [3] Drulyte *et al.* (2018) Approaches to altering particle distributions in cryo-electron microscopy sample preparation. *Acta Cryst. D* **74**:560.
- [4] Glaeser *et al.* (2017) Opinion: hazards faced by macromolecules when confined to thin aqueous films. *Biophys Rep* **3**:1.
- [5] Gatsogiannis *et al.* (2016). Membrane insertion of a Tc toxin in near-atomic detail. *Nat. Struct. Mol. Biol.* **23**:884.
- [6] Efremov *et al.* (2015) Architecture and conformational switch mechanism of the ryanodine receptor. *Nature* **517**:39.

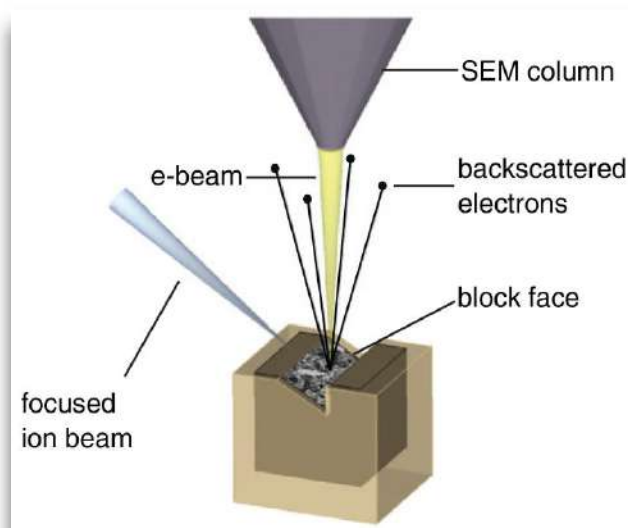
<https://www.mitegen.com/product/cryo-em-vitrification-starter-kit/>

FIB/SEM VS THIN SECTION SAMPLE PREP

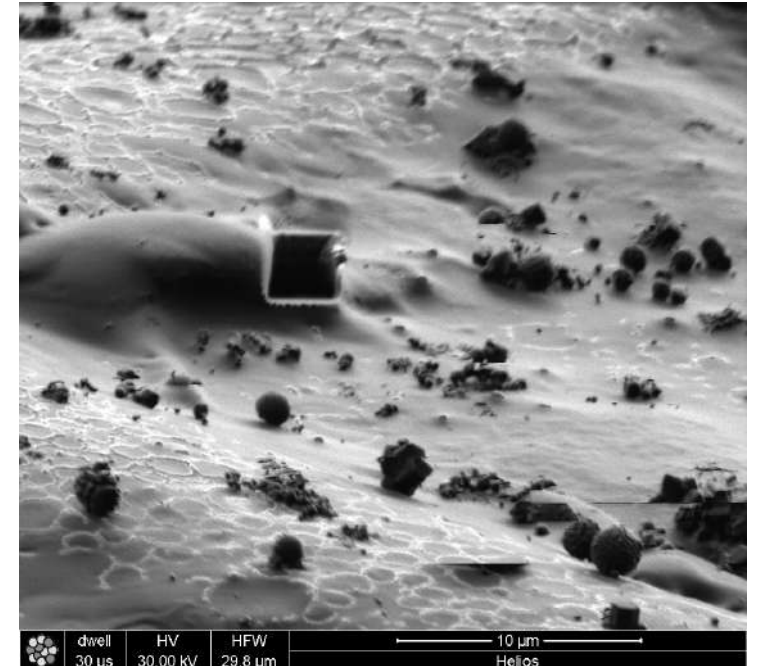
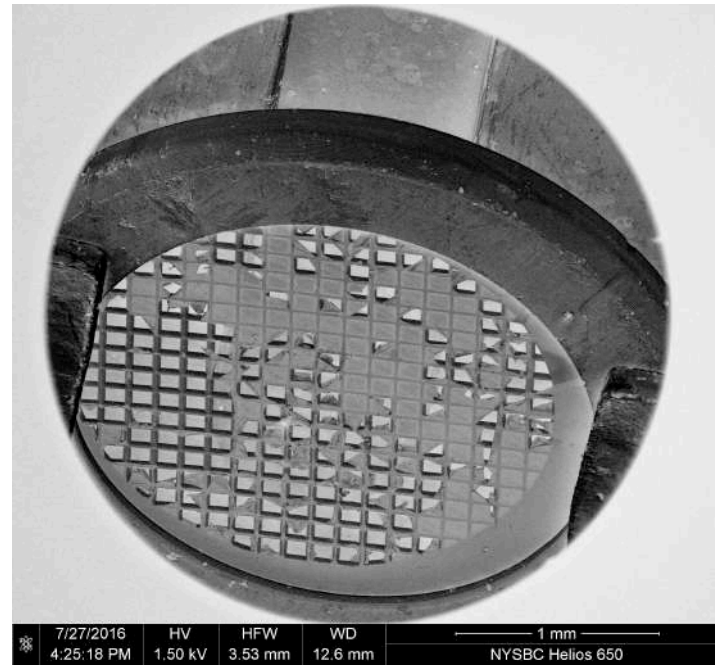
- Chemical fixation
- Staining
 - En bloc, enhanced contrast and electrical conductivity
- Dehydration
- Embedding
- Au/Pd coat
 - Conductivity

Cryofixation: High pressure freezing
Dehydration: Freeze substitution

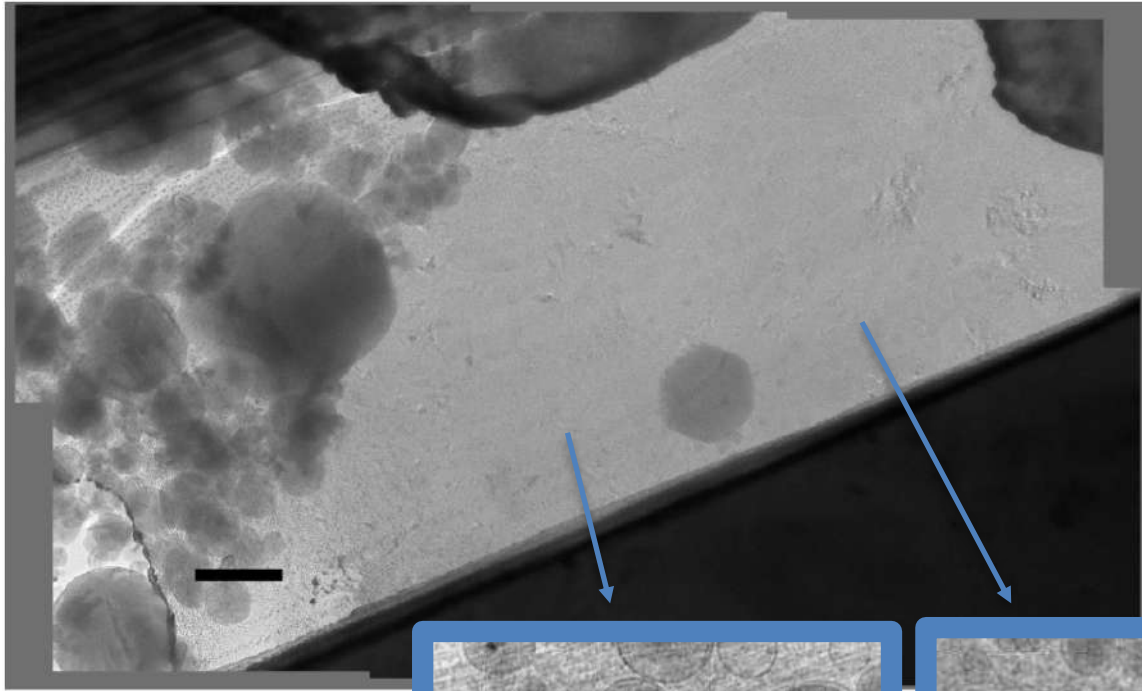
- Chemical fixation
- Dehydration
- Embedding
- Sectioning
- Staining



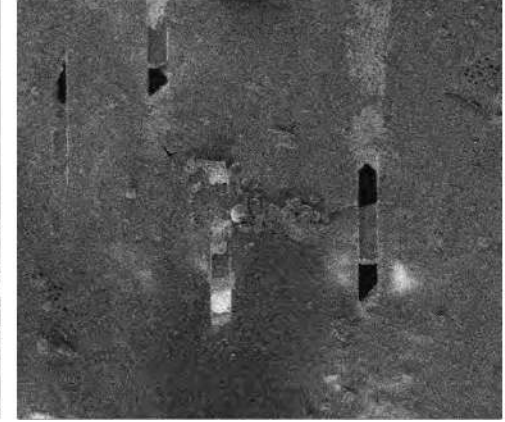
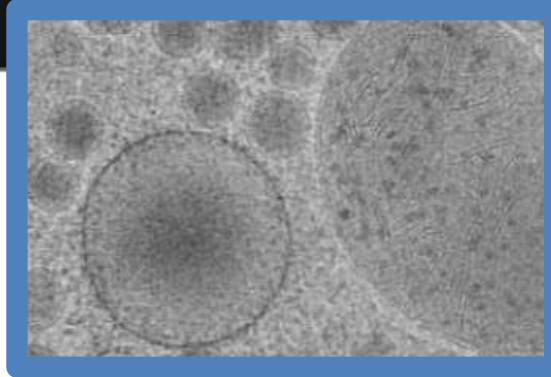
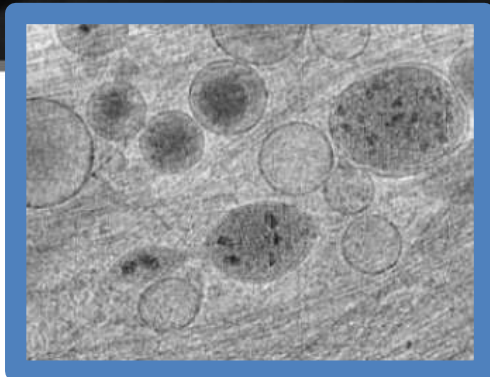
CRYO FIB MILLING



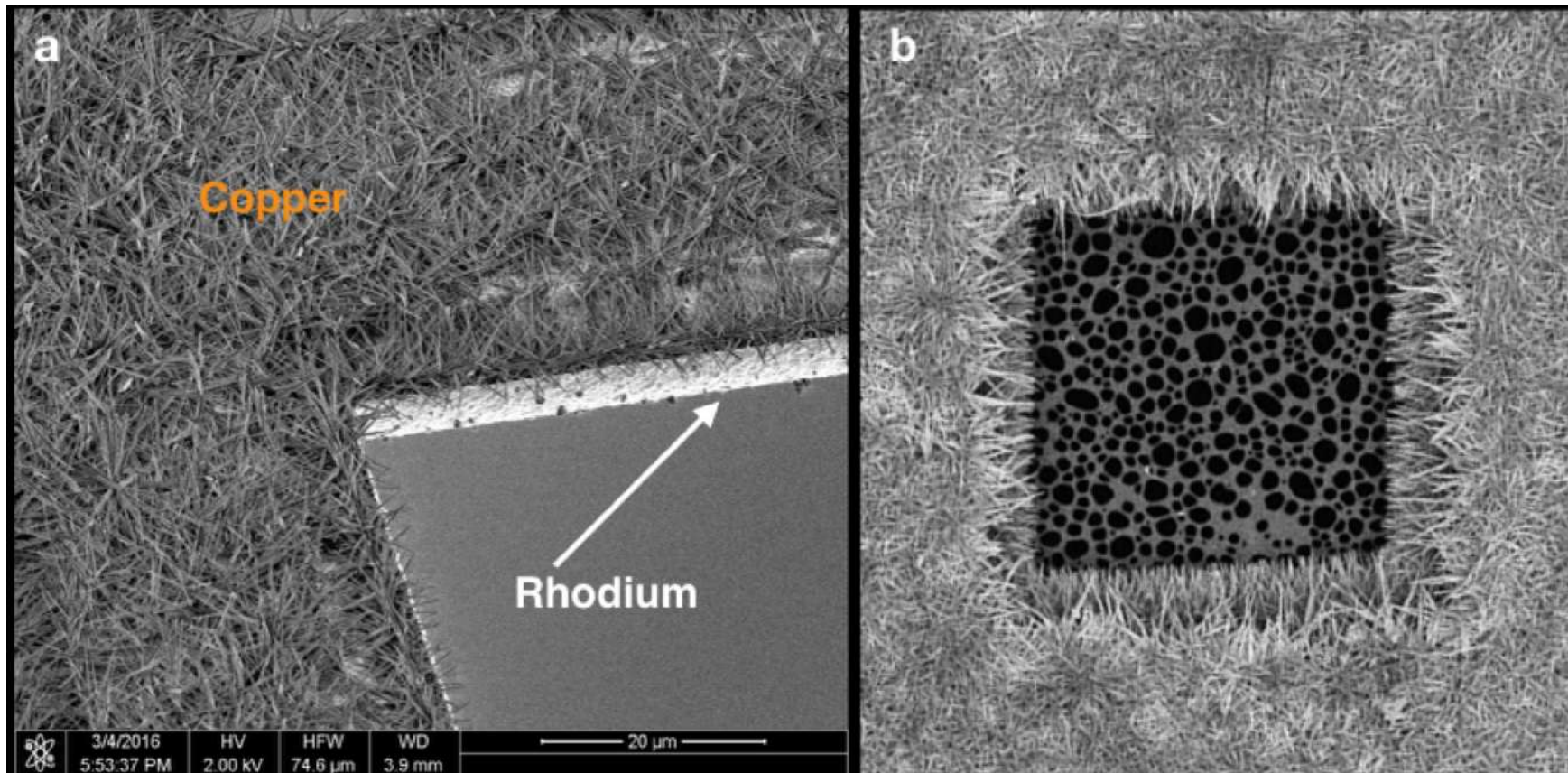
CRYO FIB MILLING



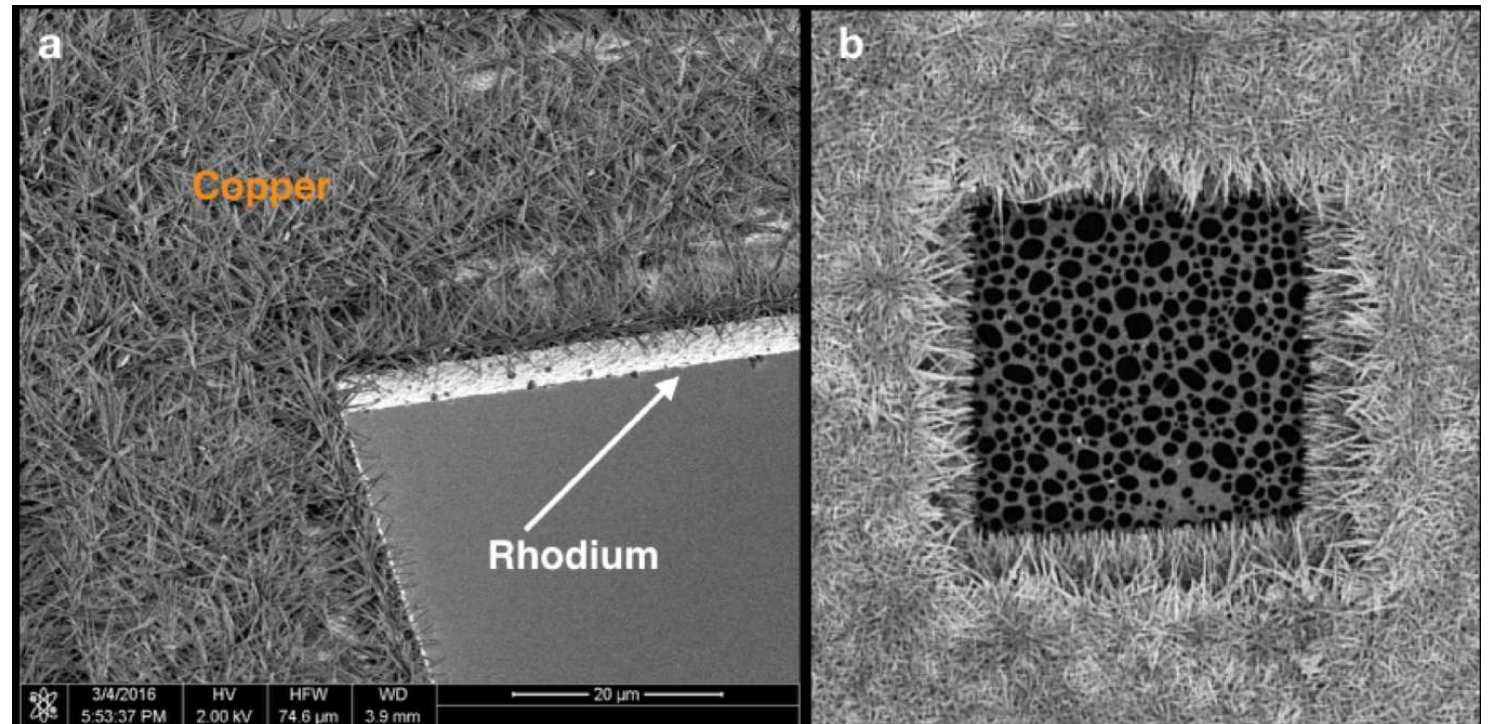
Scale bar: 1 μm



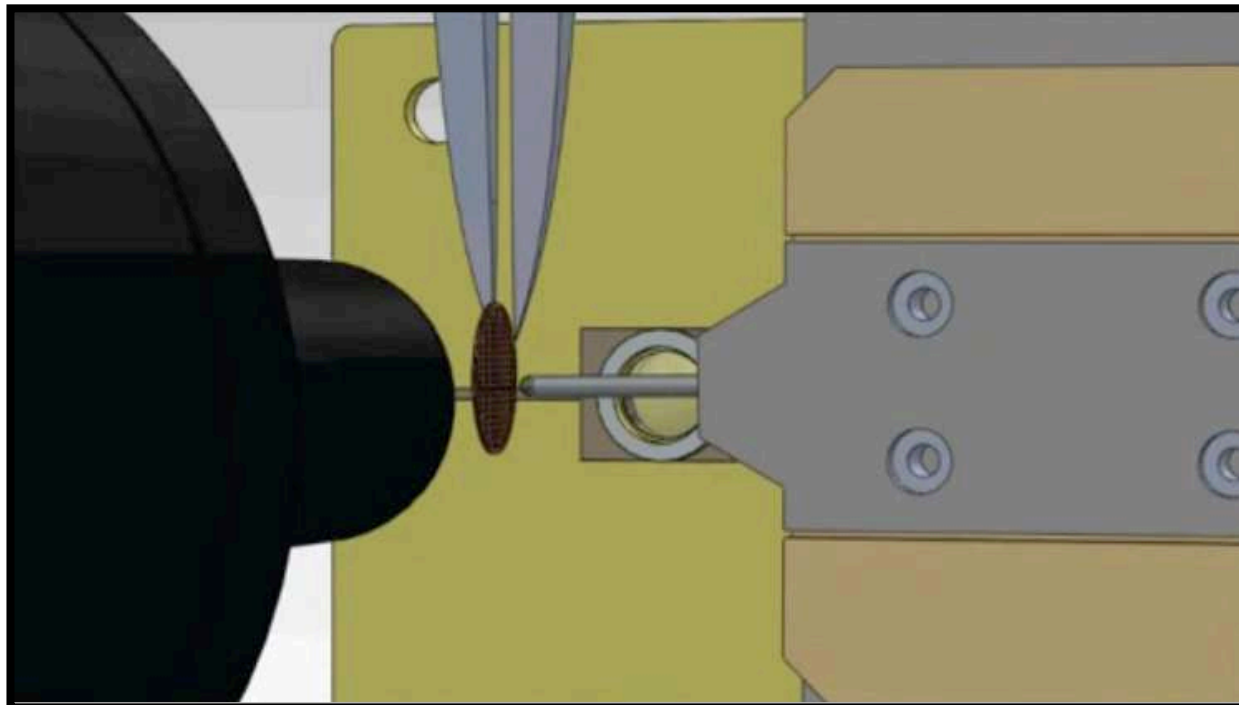
BLOT FREE VITRIFICATION



SPOTITON | CHAMELEON



SPOTITON | CHAMELEON



WHAT NEXT?

cryoEM 001 : Single Particle Masterclass

1. Building a cryoEM toolkit
2. EM compatible samples
3. EM support films and grids
4. Sample preparation
5. Tools of the trade:
microscopes and detectors
6. Microscope operations
7. Data collection strategies
8. Data assessment & QC
9. Data processing:
 - cryoEM IT infrastructure
 - On-the-fly feedback
 - 3D Reconstruction
10. Visualization and validation

